

Health Care-Associated Infections: Controlled Delivery of Cationic Antiseptics from Polymeric Excipients

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1 Abstract

2 Nowadays, the treatment of health care-associated infections represents a serious issue, due to the
3 increasing number of bacterial strains resistant to traditional antibiotics. The use of antiseptics like
4 quaternary ammonium salts and biguanides is a viable alternative to face these life-threatening
5 infections. However, their inherent toxicity as well as the necessity of providing a sustained release
6 to avoid the formation of pathogen biofilms are compelling obstacles towards their assessment in the
7 hospitals. Within this framework, the role of polymeric drug delivery systems is fundamental to
8 overcome the aforementioned problems. Biocompatibility, biodegradability and excipient-drug
9 interactions are crucial properties determining the efficacy of the formulation. In this work, we
10 provide an in-depth analysis of the polymer drug delivery systems that have been developed or are
11 under development for the sustained release of positively charged antiseptics, highlighting the crucial
12 characteristics that allowed to achieve the most relevant therapeutic effects. We reported and
13 compared natural occurring polymers and synthetic carriers to show their pros and cons and
14 applicability in the treatment of health care-associated infections. Then, the discussion is focused on
15 a particularly relevant class of materials adopted for the scope, represented by polyesters, which gave
16 rise, due to their biodegradability, to the field of resorbable drug delivery devices. Finally, a specific
17 analysis on the effect of the polymer functionalization over the formulation performances for the
18 different types of polymeric carriers is presented.

19

20 **Keywords:** Health care-associated infections; Cationic antiseptics; Polymeric drug delivery systems;
21 Controlled delivery; Sustained release.

22

23 1. Introduction

24 In the recent years, biocompatible polymers attracted an increasing interest for the production of drug
25 delivery devices, stimulated by the wide range of thermomechanical and physico-chemical properties
26 accessible with these materials. Biodegradability, tunable mechanical properties and easiness of
27 functionalization are attractive characteristics for the application of these materials to various clinical
28 treatments¹⁻³. Among them, an emerging field of application is represented by the sustained release
29 of antiseptic compounds for the treatment of health care-associated infections (HCAIs)⁴.

30 HCAs are pathologies that cause long hospital stays, high morbidity and mortality risk, development
31 of antibacterial resistant microorganisms and additional costs for the health system, specifically for
32 intensive health care units^{5,6}.

33 The systemic literature review of national and multicenter studies in high-income countries published
34 by the World Health Organization (WHO) presents a HCAI prevalence value, which refers to the
35 hospitalized patients who acquired at least one HCAI, ranging from 3.5% to 12%⁷. In order to clarify
36 the implications of those values, according to the European Centre for Disease Prevention and Control
37 (ECDC), every year in Europe more than 4 million people are affected by HCAI, accounting for direct
38 costs of approximately €7 billion⁸. Among these, surgical site infections (SSIs), the category in which
39 the pathogen is acquired during the surgical procedure, deserve significant attention being the second
40 most reported HCAs in Europe and in United States, as well as the most frequent in low- and middle-
41 income countries^{5,9-13}.

42 The main cause of this phenomenon is the abuse and misuse of antibiotics done by the society in the
43 past decades. Indeed, these are among the most prescribed medicines in human history, with an
44 estimation of 50% of unneeded prescriptions¹⁴. Consequently, this pushed the microbial evolution
45 towards the spreading of resistant strains¹⁵. As a matter of fact, in the last decades, *Staphylococcus*
46 *aureus* developed the resistance against methicillin (MRSA) first and then against vancomycin
47 (VRSA)^{16,17}. *Staphylococcus epidermidis* also showed an increased number of strains resistant to
48 rifampicin¹⁸. It is then evident that antibiotics are progressively losing their reliability in the fight
49 against these strains¹⁹. The situation is worsened by the fact that the total amount of antibiotics
50 intended for human use is outnumbered by that used by food industries, which contribute significantly
51 to antibiotic resistance, as animals serve as carriers for the development of drug-resistant strains^{14,20}.

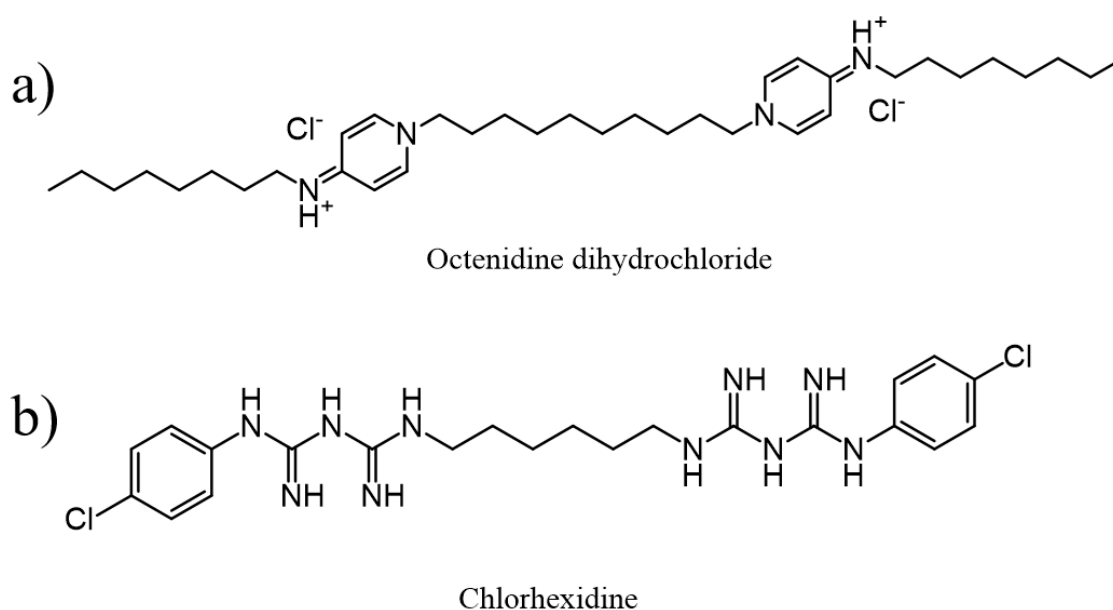
52 In this context, the choice of antiseptics as therapeutics for topical application is gaining an increasing
53 interest in order to reduce the use of antibiotics and promote an efficient treatment of resistant strains.
54 The positive outcome of the antiseptic-based treatments relies on the careful control of the dosage. In
55 fact, the antiseptics, unlike antibiotics, do not work on specific cellular pathways but rather exert their
56 toxicity on membrane efflux pumps, intracellular and nuclear functions, thus presenting a broader
57 spectrum of activity²¹⁻²³. A wide range of active chemical agents, known as biocides, including
58 biguanides, bisphenols, halogen-releasing agents, heavy metal derivatives and quaternary ammonium
59 compounds may be present in antiseptic products²⁴. Among them, this review is focused on the
60 formulation of two interesting classes of positively charged compounds, namely quaternary
61 ammonium compounds (QAC) and biguanides²⁵⁻²⁸.

62 Their efficacy against a specific bacterium is expressed in terms of Minimal Inhibitory Concentration
63 (MIC) and Minimal Bactericidal Concentration (MBC), which represent the minimal concentration
64 required to inhibit bacterial growth or to kill bacteria, under *in vitro* conditions, respectively²⁹.

65 In particular, QACs are broad-spectrum antibacterial agents applied in medical, industrial and
66 household contexts. Their general structure comprises a positively charged nitrogen “head”, linked
67 to alkyl chains of different length²⁴. QACs tend to be quite stable and water-soluble compounds. In
68 solution, ionization results in the production of a cation, the substituted nitrogen part, which provides
69 surface activity. They are characterized by an amphiphilic behavior provided by a polar head group
70 and hydrophobic hydrocarbon chains, resulting in a detergent-like mechanism against microbes²⁴.

71 Being the electrostatic interaction the driving force for non-specific binding, QACs present an
72 inherent toxicity also to mammalian cells. Indeed, their toxic effects are hampering their use for
73 systemic applications, which would necessitate a careful control on the amount of drug administered.
74 However, they are widely accepted for topical applications³⁰. Among them, octenidine
75 dihydrochloride (OCT) is one of the most used in the treatments for skin, mucous membrane and
76 wound management decolonization³¹. It is a cationic surfactant bearing two cationic centres separated
77 by a long aliphatic C10 chain, as shown in **Figure 1a**. OCT is effective against both Gram-positive
78 and Gram-negative bacteria (e.g. *S. aureus*, *E. coli*), fungi, yeasts, dermatophytes, enveloped viruses
79 and echinococcal cysts and no resistance is known yet^{31,32}.

80 Regarding biguanides, these are naturally occurring cations that generally exhibit antibacterial
81 activity against Gram-positive and Gram-negative microorganisms³³. Among these, chlorhexidine
82 (CHX) is a widely applied therapeutic due to its broad-spectrum efficacy, substantivity properties and
83 low irritation. It is present in a wide range of products to prevent infections, such as preoperative skin
84 cleansing, surgical site preparation and personal hand antiseptic products. Specifically, CHX is a
85 chlorinated biguanide that is present as bis-cation at physiological pH, as shown in **Figure 1b**³⁴. CHX
86 positive charge is capable of interacting via non-specific binding with the phospholipid membrane of
87 the negatively charged bacteria, causing a damage in the outer layers of the cell^{35,36}. Despite
88 advantages such as a wide antimicrobial spectrum and substantivity, CHX activity is pH dependent.
89 Moreover, *in vitro* studies have demonstrated that CHX, as many commonly used antiseptics such as
90 OCT and povidone-iodine, presents specific dose-dependent and time-dependent cytotoxicity profiles
91 on fibroblasts, myoblasts, and osteoblasts³⁷⁻⁴⁰.
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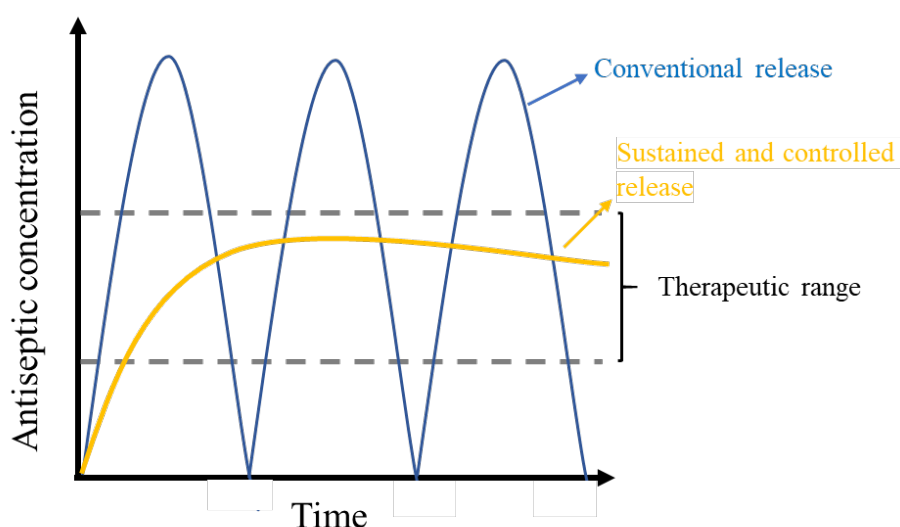
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94 Figure 1: Chemical structure of (a) octenidine dihydrochloride and (b) chlorhexidine.

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96 In this frame, it is worth mentioning also that for the antiseptics used in postoperative wound
97 management, generally the treatment is considered successful if it is able to provide antimicrobial
98 activity in a time window within 2 to 6 weeks⁴¹. In order to pursue this prolonged effect as well as to
99 avoid potentially dangerous drug concentration peaks, drug delivery systems able to slowly release
100 the antiseptic during time are extensively researched. Among these systems, those based on polymeric
101 materials are the most appreciated in terms of tunable properties and versatility. Different antiseptic
102 formulations based on polymer excipients have been developed, with some of them already applied
103 in the clinics. Given their importance in favoring the applicability and diffusion of the antiseptics
104 among the patient population, a systematic analysis of the requisites and characteristics for a
105 successful polymeric drug carrier able to modulate the release of biocides is urgently required.
106 This review is intended to specifically describe the polymeric drug delivery systems able to efficiently
107 encapsulate and release CHX and OCT as positively charged antiseptic compounds, focusing on the
108 critical attributes enabling to maximize the therapeutic index of these antiseptics. A review and
109 critical comparison of natural and synthetic polymers used as drug carriers is performed, with
110 particular emphasis to polyesters, which due to their biocompatibility and biodegradability gave rise
111 to the important family of resorbable devices. Finally, the most studied and efficient polymer

112 functionalization strategies able to enhance the therapeutic index of the drug by slowing down its
113 release and to minimize the number of repeated administrations as well as concentration-related side
114 effects are described.

115 2. Sustained Release of OCT and CHX

116 The pharmacological treatment of diseases is traditionally based on multiple dosing of a therapeutic
117 agent specifically formulated (i.e., tablets, capsules, and injections) in order to maintain the drug
118 concentration in the addressed site within a given therapeutic window. Regarding antiseptics,
119 following the traditional administration approaches, which consist typically in saline or water and
120 alcohol solutions and lotions, a large amount of drug is released in a short period as shown in **Figure**
121 **2** (blue curve)⁴². This determines a significant fluctuation in the drug concentration, leading to the
122 necessity for repeated administrations as well as to difficult control over the instantaneous drug
123 concentration, which may cause adverse reactions for the patient⁴³.

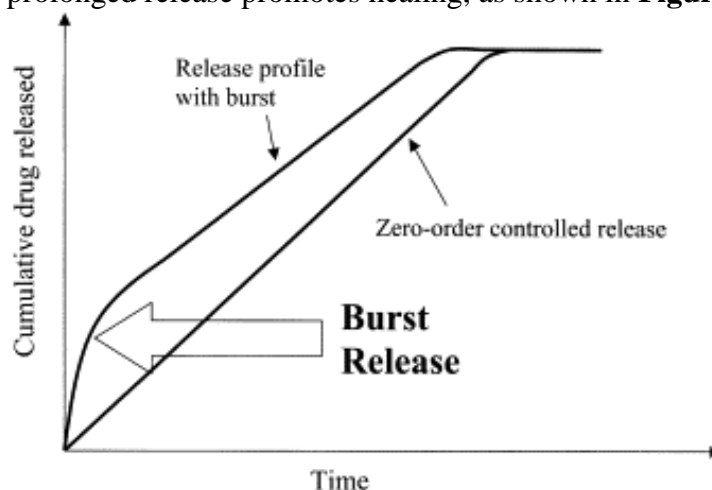


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125 *Figure 2: General antiseptic concentration profile in the targeted area for a conventional drug administration, consisting*
126 *in a in situ antiseptic administration after the surgery, with respect to a sustained and controlled delivery.*

127 These problems may be effectively overcome by the use of controlled drug delivery
128 systems, releasing the active principle slowly during time,
129 thus allowing a better predictability and reproducibility of its concentration in the targeted area. Due to
130 this higher control, an overall reduced amount of active principle in the final formulation is often
131 required, together with less administrations, leading to an improved compliance for the patient^{44,45}.
132 This represents the goal motivating the development of delivery systems acting as antiseptic
133 excipients for topical treatment, such as wound management. Indeed, sustained and local
134 antimicrobial agent release may overcome not only high dosage-related side effects but also infection
135 recurrence due to the premature suspension of the treatments and low compliances due to frequent
136 administration required⁴. There is not a standard therapeutic window in surgical wound management
137 ⁴¹. However, the time scales of the different wound healing phases, namely hemostasis, inflammation,
138 proliferation, matrix formation and remodeling, were rationalized⁴⁶. In this context, prolonged drug
139 release (over a window of at least two weeks) able to guarantee the sterility until the matrix formation
140 phase is desirable in order to avoid bacterial burden renewal as well as frequent dress change⁴⁷.
141 Moreover, an ideal release profile is represented by an initial controlled burst release followed by a
142 zero-order kinetic able to maintain an effective and safe drug concentration (i.e. between the
143 minimum effective concentration (MEC) and the maximum toxic concentration (MTC))⁴⁸. Indeed,

144 the initial burst release is aimed at providing immediate and strong antibacterial activity, while the
145 subsequent milder and prolonged release promotes healing, as shown in **Figure 3**⁴⁹.



146
147 *Figure 3: Trend of the ideal profile in the release of the antiseptics with respect to a typical zero-order release.*
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149 Being CHX and OCT the most studied and commonly used representatives of biguanides and
150 quaternary ammonium compounds, respectively, hereinafter drug delivery systems able to provide
151 long lasting release of those antimicrobial agents will be reported.

152 There are two main categories in which polymer excipients for sustained drug release could be
153 divided, namely natural and synthetic drug delivery systems. The following sections will provide an
154 overview of the polymeric drug delivery systems developed for CHX and OCT in both categories and
155 pros and cons will be analyzed in detail.

156

157 **2.1. Natural Drug Delivery Systems**

158 Natural drug delivery systems have been widely researched for the sustained release of antiseptic
159 compounds, due to their high biocompatibility to the human body. Indeed, different CHX and OCT
160 delivery systems based on cellulose⁵⁰⁻⁵³, polysaccharides⁵⁴⁻⁶⁰, collagen⁶¹ can be found in the
161 literature, as described in **Table 1**.

162 Among polysaccharides, chitosan is widely appreciated in the medical and pharmaceutical fields, due
163 to its appealing properties, such as biodegradability, biocompatibility and ability to form films and
164 gels⁶². Moreover, it provides an acceleration in the granulation phase during the wound healing
165 process while having a bacteriostatic effect and bio-adhesive properties due to its positive charges^{63,64}.
166 Currently, it represents one of the most exploited excipient for wound dressing, since it is already
167 present in the market with various products, namely ChitoSamTM, ChitoFlex, ChitoGauze XR pro and
168 Axiostat®. Therefore, chitosan-based delivery systems in the forms of film⁶⁰, hydrogel⁵⁶, micro-⁵⁸ and
169 nano-⁵⁹ particles have been widely investigated as CHX excipients.

170 Drug release kinetics from chitosan hydrogels, resulting from both drug diffusion and matrix
171 degradation, proved to be dependent and consequently controllable by the crosslinking density of
172 chitosan network thus allowing an adjustable therapeutic window⁵⁶.

173 However, since both chitosan and the antiseptic are positively charged at physiological pH, polymer-
174 drug interactions and in turn the loading efficiency are hampered by electrostatic repulsion. To
175 overcome this problem, functionalization with negatively charged groups increasing the complexity
176 of the system are required to enhance the drug loading and to improve the characteristics of the
177 formulation⁵⁹. In this context, CHX loaded hexametaphosphate (HMP) nanoparticles have been
178 exploited to provide antimicrobial properties to the commercially known Savlon® Alginate

179 Dressings⁶⁵. HMP NPs gained attention due to synthesis process easiness and the NP ability to form
 180 a stable colloidal suspension exploitable in dip coating. Colloidal stability as well as capability of fast
 181 adherence to different surfaces are both obtained thanks to NP surface charges.
 182 As a matter of fact Barbour *et al.*⁶⁵ successfully dip coated medical grade titanium with HMP NPs.
 183 The colloidal coating showed a long-lasting antimicrobial effect with a sustained CHX release over
 184 a period of 99 days. Overall, the use of chitosan may be particularly advantageous in wound dressing
 185 because of the many beneficial properties that this compound provides for the application.
 186 However, the intrinsic solubility of this polymer prevents *in vivo* long-term exposure to the exudating
 187 wounds, since their biological fluids may hamper the integrity of the device. At the same time, the
 188 polymer-drug electrostatic attraction needs to be carefully controlled in order to efficiently
 189 encapsulate and release the drug. As a matter of fact, in the case of pectin, a widely used anionic
 190 polysaccharide, strong drug-polymer interactions result in an inefficient antimicrobial activity, due to
 191 undesired drug retention. Indeed, pectin microparticle synthesis involving CHX as cross-linking
 192 agent as well as encapsulated active ingredient resulted in a drug release kinetic reaching its plateau
 193 after 20 minutes and releasing only 7% of the total drug after 6 hours. This could be solved by the
 194 simultaneous presence of another di-cation (*i.e.* zinc) able to compete with CHX as gelation agent
 195 and providing a less tight microparticle network⁵⁷. A possible solution, in order to reduce the
 196 drawbacks in the clinical application and to pave the way to a long term wound dressing device is
 197 represented by films made of a network of alginate and pectin (SA-PC) crosslinked with calcium
 198 chloride, in order to improve the physicochemical and mechanical properties of the formulation⁶⁶. In
 199 this context, promising tests performed both *in vitro* and *in vivo* validated this approach for wound
 200 dressings encapsulating antidiabetic compounds^{66,67}.

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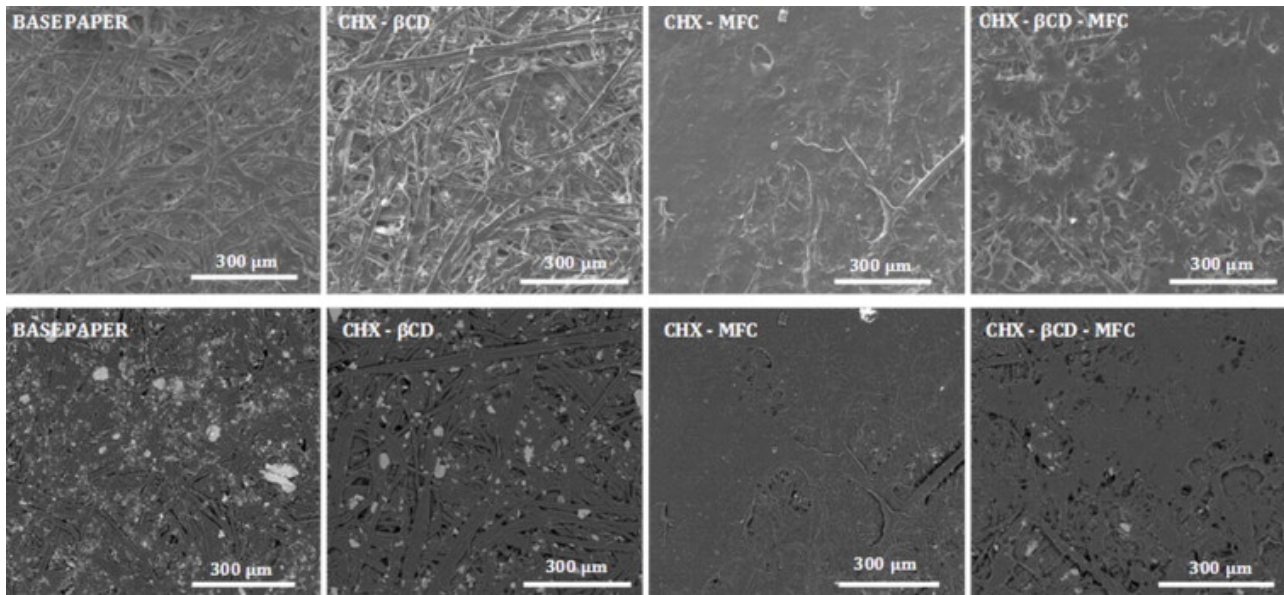
Table 1: Advantages, drawbacks and possible functionalization to improve the formulation performances for the different natural polymers used as excipients for positively charged antiseptics.

| Polymeric excipient | Formulation form | Advantages | Drawbacks | Functionalization |
|---------------------------------|--|--|---|---|
| Chitosan ^{54,56,58-60} | Hydrogel, microspheres and nanoparticles | Biodegradability, bioadhesive properties, granulation phase acceleration | Hampered polymer-drug interactions, low loading efficiency, fast drug release | Peptidomimetic derivative glutathione (GSH) ⁵⁹ , tripolyphosphate or a,b-glycerophosphate cross-linking ^{56,60} |
| Pectin ^{57,68,69} | Hydrogel, microparticles | Biodegradability, easiness of gelation | Excessive drug retention | Divalent cations (Zn ²⁺ Ca ²⁺) as gelation agents ^{57,68} |
| Collagen ^{47,61,70-72} | Film composed by a collagen matrix | Biodegradability, stimulation of healing process, polymer-drug affinity | Batch safety and reproducibility for bovine or porcine derived | Glutaraldehyde crosslinking ⁷² |
| Cellulose ⁷³⁻⁷⁶ | Matrix for wound dressing | Efficient drug loading, high stability, prevention of wound drying | Fast drug release | Addition of cyclodextrins or poloxamers ^{73,74} |

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Another natural polymer commonly applied in the medical formulation of antiseptics is collagen, since it is not only biocompatible and biodegradable, but also stimulates wound healing enhancing

207 fibroblast synthesis and endothelial cell migration^{47,70}. Denatured collagen (gelatin) matrices
208 containing 2.5 mg of chlorhexidine gluconate (CHG), commercially known as PerioChip™, are
209 widely used in the treatment of periodontal environment bacterial colonization. This delivery system
210 is able to maintain high CHG concentration for 72 h, due to both diffusion mechanism and initial
211 matrix degradation which occurs typically in a range between 7-10 days⁶¹.
212 Nonetheless, issues regarding batch safety and reproducibility of collagen needs to be overcome for
213 this type of formulations. In fact, bovine or porcine derived collagen poses transgenic transmission
214 risks besides its fast degradation that hinder the possibility of providing a long lasting release⁷⁷.
215 Towards this direction, fish-derived or denatured collagen preventing transgenic risks may
216 represent safe alternatives to conventional collagen formulations^{72,77}.
217 In this context, cellulose represents the most studied natural polymer. In fact, the highly porous nature
218 of this organic macromolecule may result in a high drug loading capacity⁷⁶. In the past, cellulose-
219 based delivery systems such as bacterial nanocellulose (BNC), gained attention owing to their unique
220 properties. BNC fulfills the requirements for usage as wound dressing, among them biocompatibility,
221 mechanical and thermal stability, softness, exudate absorbance preventing wound drying and proper
222 gaseous exchange⁷³.
223 The addition of antimicrobial properties to nanocellulose through the encapsulation of an antiseptic
224 compound leads to an active wound dressing able to improve acute and chronic wound treatment
225 outcomes.
226 As a matter of fact, Moritz *et al.*⁷⁵ successfully loaded OCT into a BNC fleece thanks to capillary and
227 diffusional forces, obtaining an 8 hours initial burst effect and 96 hours lasting sustained release.
228 Specifically, it is worth mentioning that after 24 hours, more than 80% of the loaded drug was
229 released, which may result in a frequent wound dressing change and in turn, lower patient compliance.
230 For this reason, in order to improve the capacity to sustain the release of the formulation, other
231 excipients need to be added. Specifically, different studies investigated nanocellulose formulations
232 comprising cyclodextrins (CD), which are cyclic structures of D-glucose oligosaccharides^{51,52,74}. CD
233 has a 3D truncated cone structure featuring a hydrophilic external surface and a hydrophobic inner
234 cavity. Hydrophobic or Van der Waals weak interactions allow CD to entrap guest molecules as
235 CHX⁵⁵, resulting in inclusion complexes able to delay the drug release kinetics^{78,79}.
236 Interestingly, in the study by Lavoine *et al.*⁷⁴ a combination of micro-fibrillated cellulose and CDs
237 coating successfully resulted in cellulosic substrates-based drug delivery systems of CHG as shown
238 in **Figure 4**.
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Figure 4: Scanning electron microscopy (SEM) images of the different surfaces on the cellulosic substrate varying the wound dressing formulation (from left to right): cellulosic substrate, CHX and CD, CHX and micro-fibrillated cellulose (MFC), CHX, MFC and CD. The upper panels show the topography of the samples measured with Everhart–Thornley detector (ETD) while the bottom ones provide information about the homogeneity of the coating through the back-scatter electron (BSE) mode in order to highlight the distribution of both the cellulosic parts and the fillers. Reproduced with permission from ⁷⁴. Copyright Elsevier 2014.

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Particularly, the coupling of these two excipients produced a synergistic combination of effects: the micro-fibrillated cellulose reduced the initial burst effect, while the presence of a CD drug reservoir resulted in prolonged release. In this case, the results were observed for over 50 hours. Overall, the main advantages of natural drug delivery systems include their biocompatibility and biodegradability ensuring patient safety. In addition, concerning the wound dressing application, their capacity to absorb the wound exudate guaranteeing a sufficient air permeability while promoting the healing process represents characteristics of paramount importance in the clinical application. Moreover, being produced from renewable sources, natural drug carriers may reduce the carbon footprint in the medical and pharmaceutical economy. However, their low stability, batch-to-batch variability, possibility of transgenic transmission and the difficulty in reaching a long-lasting release make them less versatile drug carriers compared to their synthetic counterparts. This is even more so considering the poor control over their chemical and mechanical properties during the synthesis. The lack of control during the production process and the drop in the physicochemical characteristics of the formulation during time hinder the possibility to provide a long-term sterility of the wound, fundamental in the future in the fight against HCAs infections. In order to overcome these issues, synthetic materials often represent the solution, owing to their versatility and high reproducible as well as tunable properties.

266 2.2. Synthetic Drug Delivery Systems

267 Concerning CHX and OCT releasing systems based on synthetic polymers, in this chapter the focus
268 will be mainly on the non-biodegradable excipients while the biodegradable ones will be treated
269 separately in the next sections.

270 Among synthetic polymers, the most studied delivery systems are represented by methacrylates⁸⁰⁻⁸⁵,
271 polyurethanes⁸⁶⁻⁸⁹, silicon^{90,91}, alkoxy silanes⁹², ethylene-vinyl acetate copolymers⁹³⁻⁹⁵, self-
272 assembled polyelectrolytes⁹⁶⁻⁹⁸.

273 These synthetic polymers, differently from the natural-derived ones, are mainly applied as long-
274 lasting antimicrobial releasing carriers in order to respond to the rising demand of HCAs from dental
275 as well as orthopedic surgery. In this field, an important aspect is related to the mechanical properties
276 of the formulation, which should be kept for the whole duration of the release. As matter of fact, CHX
277 and OCT are widely applied in dental restorative formulations, oral fungal infections and in surgical
278 procedures as active ingredients inside bone cements.

279 Poly(methyl methacrylate) (PMMA) for example has been widely used as antibiotic releasing bone
280 cement since 1960⁸⁴. Addressing the threat of antibiotic-resistant bacteria, OCT loaded PMMA bone
281 cements were investigated as an effective solution in the treatment and prevention of infections due
282 to orthopedic surgical procedures. As a matter of fact, Weckbach *et al.* mixed together cement and
283 OCT powder to obtain the commercially known bone cement Palacos®, achieving a low but sustained
284 release up to 6 weeks with a prolonged antimicrobial efficacy against *S. Aureus*⁸⁵.

285 Methacrylate-based systems in different formulations have been further investigated as CHX loading
286 systems⁸⁰⁻⁸⁵. Specifically, in dental applications, the drug delivery system may be produced via *in*
287 *situ* polymerization exploiting a photo-initiator, where the drug is entrapped in the matrix during the
288 radical-driven polymerization process. However, the main problem found in the application of
289 cationic antiseptics as biguanide and QACs as active principles for methacrylate-based formulations
290 is the protonation of the amine groups of the drug that is thought to consume free radicals, interfering
291 with the radical polymerization⁸¹. This could result in the lack of desired properties as well as side
292 effects due to an uncontrolled drug release, making this type of polymers not particularly suitable for
293 the application.

294 As an alternative, polyurethane-based CHX carriers under different forms such as films or three-layer
295 systems^{86,87}, antimicrobial patches⁸⁸ and nanoporous webs⁸⁹ have been investigated.

296 Since polyurethanes (PU) are a class of polymers synthesized from the polymerization of di-
297 isocyanates and diols (or polyols), they can be obtained from a wide range of available building
298 blocks, paving the way to the fine tuning of their mechanical and physico-chemical properties⁹⁹.

299 Polyurethane-based releasing systems are widely appreciated in biomedical industry: BIOPATCH®,
300 a polyurethane-based patch for the release of CHX, replaced every week, is proved to be more
301 effective in reducing pin-tract infection than topical antibiotic treatment performed twice a day with
302 three different antibiotics, owing to its 7 days sustained release⁸⁸.

303 In the study of Huynh *et al.*⁸⁶, a medical grade polyurethane under different forms (e.g. film and three
304 layer system) was studied *in vitro* as chlorhexidine diacetate (CDA) long lasting delivery system. The
305 drug release rate and tunability were demonstrated to be dependent from the structure of the drug
306 carrier. Indeed, the simplest formulation consisting in the PU film showed an initial burst directly
307 dependent on the drug loading and a kinetic of release strongly dependent on the salt concentration
308 in aqueous phase. These limitations, which prevent a long-term release of the compound may be
309 overcome by using a three-layer system.

310 In fact, a three-layer delivery system featuring two outer unloaded PU layer serving as rate-controllers
311 provided a 29 days lasting release following zero-order kinetics in distilled water as well as in
312 physiological saline solution.

313 Indeed, PU multilayer system attracted the interest from researchers as coating material for their
314 controlled release and higher drug loading capacity compared to the single layer reservoir.

315 As a matter of fact, the release kinetics of two different CHX salts loaded in a three-layer polyurethane
316 drug carrier proved to last 42 days with an initial 24 hours burst effect⁸⁷. The authors reported the

317 delaying effect of the two unloaded layers and the different CHX salt solubility as key factors
318 influencing the diffusion-driven drug release¹⁰⁰.

319 Among the different efforts towards infection prevention and management, the synthesis of
320 polyelectrolyte films proved to be particularly interesting as antimicrobial agent releasing wound
321 dressing^{96,97}. A step forward in this direction is attempted by Agarwal *et al.*⁹⁶ who provided sustained
322 antimicrobial activity to a biological wound dressing by stamping a polyelectrolyte multilayer film
323 incorporating CDA on top of it.

324 Layer-by-layer ultrathin nanostructures serving as matrices for localized drug release proved to last
325 for 8 days *in vitro*. However, the stamping process may undermine the local stability of CDA loaded
326 thin film, resulting in more rapid elution of CDA.

327 Following this discussion, it should be clear that the achievement of localized, controlled and
328 sustained antiseptic release able to reduce the bacterial colonization and increase the treatment
329 compliance poses many constraints to be satisfied. Biocompatibility, process reproducibility and
330 mechanical properties tunability are the fundamental performances to be provided by the formulation
331 for these applications. However, even if the synthetic polymers aforementioned are able to guarantee
332 these properties, a further important point of discussion is the fate of the drug carrier after the
333 complete release of the drug. In order to avoid accumulation of plastic in the environment and, even
334 worse, in the human body, the biodegradability of the carrier represents a key characteristic. This is
335 the rationale behind the wide research and investigation towards polyester-based drug carriers, which
336 will be discussed in the following section.

337

338 **3. Polyester-Based Drug Delivery Systems**

339 Synthetic polyesters as polylactide (PLA), polyglycolide (PGA), polycaprolactone (PCL) and their
340 copolymers gained increasing interest in the development of drug delivery systems for their
341 biocompatibility, tunable properties, and degradability into non-toxic and well resorbed molecules,
342 which avoids further surgery for the removal of the implants^{101,102}. In those devices, the release of the
343 drug is often achieved by diffusion, whose rate is strongly influenced by the progressive degradation
344 of the polymer matrix, in which the therapeutic is incorporated. At the same time, the high control
345 over the synthesis processes enables reproducibility in the final product as well as tunable degradation
346 rates and customizable mechanical characteristics for a good control over the drug release^{103,104}.
347 Compared to natural polymers that undergo enzymatic degradation only, polyesters degrade through
348 both enzymatic or hydrolytic cleavage of the labile ester bonds, leading to a progressive reduction in
349 the molecular weight¹⁰². Specifically, the degradation of insoluble polyester particles in water may
350 occur via bulk or surface erosion¹⁰⁵. The former consists in a roughly homogeneous decay of the
351 molecular weight and density involving the whole polymer volume, with the produced fragments that
352 diffuse outside the polymer matrix. On the other hand, the surface erosion is described by a
353 progressive degradation occurring mainly on the surface and causing a progressive shrinking in the
354 polymer volume¹⁰⁶. Normally the surface erosion rate is constant until the complete dissolution of the
355 polymer, while the bulk erosion one depends on different factors and may vary during time¹⁰⁷. The
356 occurrence of one or the other mechanism is mainly influenced by two processes, namely the water
357 diffusion into the polymeric matrix, the subsequent chain cleavage and the retro diffusion of the
358 fragments from the bulk¹⁰⁸. Indeed, if the water and fragment diffusion inside and from the bulk is
359 faster than the chain cleavage, the material is characterized by bulk erosion degradation, otherwise the
360 surface erosion is predominant¹⁰⁸. It is also worth mentioning that these phenomena depend also on
361 the polymer size. Each polymer is characterized by a critical size above which the bulk erosion is

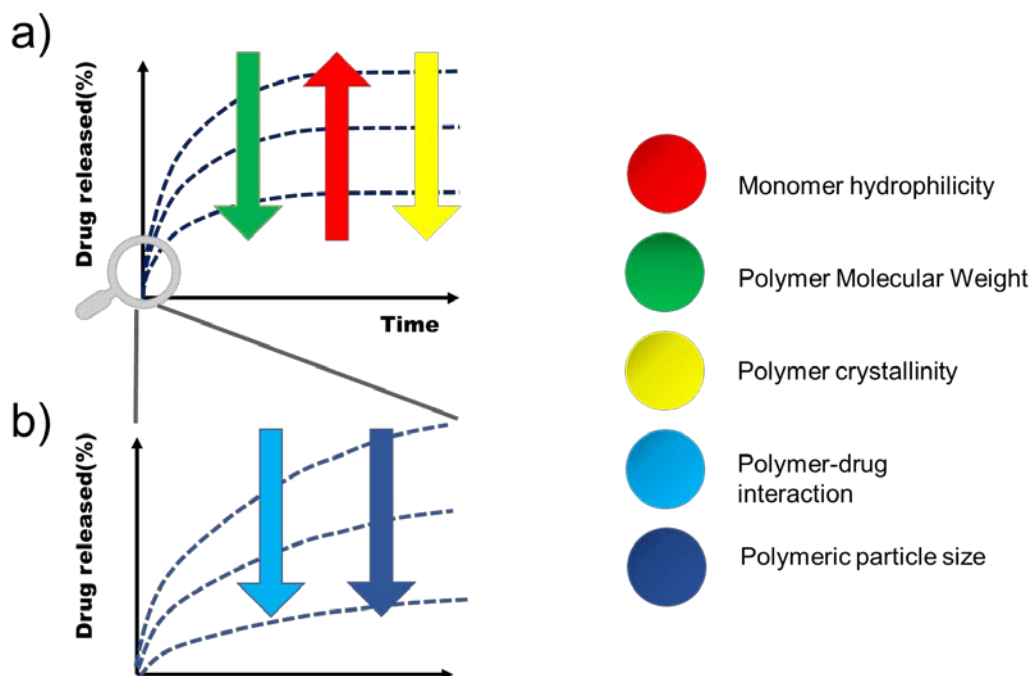
362 lost. In this context, aliphatic polyesters typically show a bulk erosion behavior consisting in 10^{-1} m
363 as characteristic size and their characteristic degradation time can be easily controlled. Furthermore,
364 another factor that plays a key role in the aliphatic polyester degradation process is represented by
365 the autocatalysis. Indeed, the breakage of the ester bonds inside the polymer matrix leads to the
366 formation of acidic products that lowers the microenvironment pH, thus catalyzing the further
367 hydrolysis of the ester bonds in the polymer backbone^{109,110}. This phenomenon, in the drug delivery
368 application may be triggered not only by the polymer fragments but also by certain type of drugs that
369 are loaded inside the carrier. In fact, the local increase or decrease of the pH inside the polymeric
370 matrix caused by the active principle molecules represents a common effect measured in the
371 polymeric carrier¹¹¹⁻¹¹³. However, this should not represent an issue when dealing with quaternary
372 ammonium salts. In fact, their pKa is above the pH of both the water and the human body, therefore
373 their protonation state is negligible during the production and the application of the formulation, not
374 influencing the microenvironmental pH of the matrix. Finally, the other important parameters
375 involved in the polymer degradation consist in the copolymer composition, molecular weight,
376 crystallinity and stereochemistry of the monomers, whose tuning makes it is possible to adapt the
377 formulation characteristics and degradation rates to the different applications^{104,114}. Indeed, the slow
378 degradation rate of PLA makes it one of the most common choices for the production of long-term
379 orthopedic implants such as plates and screws. Whereas, PGA and its copolymers with lactic acid
380 (PLGA) present faster degradation, thus making it particularly suitable for the fabrication of sutures
381 and drug delivery devices¹¹⁵. The timespan of PLGA hydrolytic degradation is dependent on
382 glycolide to lactide (G/L) ratio. Since glycolide is less hydrophobic than lactide, by increasing its
383 proportion in the PLGA it is possible to significantly speed up the degradation. As an example,
384 increasing the G/L weight ratio from 25:75 to 50:50 can accelerate the degradation by two-fold from
385 100 to 50 days^{103,115,116}.

386 Another advantage in using polyesters is the possibility of providing a broad range of mechanical
387 properties. Indeed, it is possible to obtain polyesters with a relatively brittle structure like PGA as
388 well as highly flexible ones as PCL, despite this is typically characterized by a low tensile modulus,
389 which may hamper its application in several fields¹¹⁷.

390 Following the aforementioned promising features, the research is increasing its efforts in the
391 application of these compounds as excipients to achieve a sustained and controlled release of
392 antibiotics and antiseptics^{115,118,119}. Specifically, in this section, a focus on the role played by the
393 different polyester-based delivery devices as fibers and films for wound dressing, patch, micro- and
394 nanoparticles is reported in the delivery of antiseptics.

395 Young *et al.* investigated the use of injectable adhesives made of lactic acid (LA) and propylene
396 glycol (PG) dimethacrylates for the sustained release of CHX during bone repair. The drug release
397 study from polymeric disks reported a direct correlation between the LA and PG block molecular
398 weights and the CHX release, due to a reduction in cross-linking and to the interaction between the
399 degradation product of the polymer and the drug¹¹⁹. However, the low affinity between the polymeric
400 excipient and the drug caused a significant early release of the active principle due to its low
401 encapsulation efficiency inside the polymeric matrix. Moreover, the possible dependency of the
402 antiseptic release from the disc size may create problems in the final application of the formulation.
403 For these reasons, a more extensive investigation focused on the application of different types of
404 polyesters, namely PCL and PLA as fiber-based delivery systems was performed. Indeed, PCL
405 sutures endowing CDA were realized by Scaffaro *et al.* to prevent infections on sutured wounds. The
406 polymer monofilaments were spun thanks to a capillary rheometer and showed a sustained drug
407 release up to one week with a burst release that increased with the drug concentration¹²⁰. Instead,
408 PLA fibers were mainly used to encapsulate antiseptics through electrospinning. Different works

409 investigated the characteristics of PLA fibers containing CHX used either alone or to reinforce PCL
 410 films^{121,122}. Specifically, Luo *et al.* showed a fast drug diffusion from the fibers in the first day, with
 411 a prolonged release evaluated for up to 650 hours. When the same drug particles were encapsulated
 412 with a layer-by-layer strategy, the release rate significantly decreased. On the other hand, when the
 413 PLA fibers were used as a reinforcement to PCL films, a significantly faster drug release and lower
 414 infection rate was observed when both the fibers and the film were loaded, with complete bacterial
 415 growth inhibition¹²¹. Therefore, the use of the electrospinning technique provided a versatile tool able
 416 to tailor the suture fibers drug release and initial burst for the application. Moreover, the possibility
 417 to electrospin different materials together may be particularly advantageous for the wound dressing
 418 application. Indeed, the use of this technique with PLA or PLGA coupled with natural polymers as
 419 collagen and chitosan may guarantee at the same time the mechanical properties and the strict control
 420 over the drug release provided by biodegradable polyesters and the promotion of the wound healing
 421 of these natural polymers^{123,124}. In this context, another promising formulation is that of thermogelling
 422 polymers. In fact, triblock polymers made of PLGA-PEG-PLGA are able to provide the correct
 423 exudate absorption and air permeability, beside the tunable release of the drug and the easiness of
 424 application¹²⁵. In this way, by exploiting the physicochemical characteristics of the formulation (*e.g.*
 425 hydrogel or fibers) it is possible to achieve the set of properties required by the application.
 426 Despite these raising applications of polyesters, they are mostly used in the production of micro- and
 427 nanoparticles for drug delivery. Indeed, biodegradable particles are already a well-established method
 428 to efficiently deliver therapeutics for the treatment of various pathologies like diabetes and cancer
 429 and for vaccine delivery due to their interesting properties¹²⁶⁻¹²⁸.
 430 In fact, polymeric microparticles (MPs) are extensively employed as biomaterial because of their
 431 favorable characteristics in terms of simple production and design, good biocompatibility and broad
 432 structure variety¹²⁹⁻¹³¹.



433
 434
 435 *Figure 5: Schematic representation on the effect of the polyester characteristics on the release of the active principle. a)*
 436 *Influence of the most significant polymer and monomer characteristics on the kinetic of release of the antiseptic from the*
 437 *formulation; b) Effect of the formulation characteristics as the polymeric particle size and polymer-drug interaction on*
 438 *the drug encapsulation and initial burst.*

439

440 “Microparticle” is the term used for spherical particles with diameters in the micrometer range
441 (typically from 1 μm to 1000 μm)^{126,132}. There are different interesting features of MPs that make
442 them particularly suitable for antiseptic release: (i) controlled release of encapsulated materials, (ii)
443 protection of the encapsulated materials against degradative reactions (e.g., oxidation, dehydration,
444 UV, heat, acids and bases) in the external environment (iii) improved shelf life, (iv) delivery of
445 therapeutics into the target location with high efficiency and for long time periods, (v) enhancement
446 of solubility profile of poorly soluble active agents, (vi) enhancement of bioavailability of the loaded
447 drug and reduced side effects^{127,128,131}. MPs are usually formed by a polymer matrix in which an active
448 compound can be immobilized, either on the surface or entrapped inside the bulk. With respect to the
449 distribution of the active compound, two different categories of MPs can be distinguished:
450 microspheres and microcapsules. Microspheres refer to MPs composed of a homogeneous mixture
451 comprising the active principle and the excipient, while in microcapsules the core is either empty or
452 liquid (e.g. vesicles) in order to efficiently encapsulate and tune the release of hydrophilic compounds.
453 Furthermore, one or more discrete domains of active compound may be found in the microcapsule
454 core¹³³.

455 Drug molecules can be released from polymeric MPs through water filled pores, following two
456 different processes: convection produced by the osmotic pressure and the diffusion caused by random
457 movements due to the concentration gradient. In addition to diffusion, the drug molecules can be
458 released from the polymer matrix by erosion¹³⁴. As such, the drug release can be influenced by
459 morphological properties of the polymer selected, such as molecular weight, molecular weight
460 distribution, copolymer composition, presence of end functional groups, crystallinity and solubility,
461 as described in **Figure 5**^{135–141}.

462 In the recent years, a significant effort was done in order to apply these delivery systems to
463 compounds as QACs and biguanides.

464 Specifically, in order to efficiently encapsulate antiseptics in MPs, the polymer-drug interaction has
465 to be enhanced. In fact, PLGA MPs providing a sustained release of the antiseptic were investigated
466 in a formulation of povidone-iodine, as an intramammary disinfectant delivery system¹⁴². Despite the
467 sustained diffusion of the antiseptic compound over 28 days, the moderate drug encapsulation and
468 the significant initial burst lowered the formulation efficacy. Furthermore, in this case, the method of
469 production of the formulation (i.e. solvent evaporation method) provoked a low process yield
470 complicating an eventual subsequent scale-up.

471 Instead, the presence of molecules able to improve the affinity between the polymeric carrier and the
472 antiseptic may significantly increase the drug retention and encapsulation. As an example, Yue *et al.*
473 produced a CHX delivery system composed by PLGA MPs able to provide the sustained release of
474 the active principle. Specifically, the prolonged drug retention was obtained through the interaction
475 of CHX with cyclodextrins characterized by a different lipophilicity. It was proved also that
476 increasing the hydrophilicity of the cyclodextrin (e.g. *hydroxypropyl beta-cyclodextrin*) a better
477 encapsulation and a faster release were obtained¹⁴³. In this context, the biocompatibility of the
478 polyester formulation can be also further improved through the addition of natural polymers in a
479 blend^{144,145}. Indeed, PLGA-based MPs with a shell of glycol-chitosan were synthesized by Chen *et al.*
480 Specifically, CHX was released together with different growth factors in order to promote wound
481 regeneration together with antibacterial activity. In these systems, the rate of the drug was proved to
482 be dependent on the copolymer composition. In fact, 85:15 PLGA formulations showed a sustained
483 release up to one week with efficient antibacterial activity, while worse results were found for 50:50
484 PLGA MPs¹⁴⁵.

485 However, depending on the final application, particles characterized by a smaller size, as
486 nanoparticles (NPs) may be preferred. Indeed, NPs generally range from 10 nm to 1000 nm in

487 size^{126,146}. Exploiting their size at the nanoscale, these nanostructures can circulate in the blood for
488 prolonged periods of time, with a controlled release that causes lower drug level fluctuations in the
489 plasma¹⁴⁷. Moreover, similarly to microparticles, they offer a wide variety of advantages compared
490 to conventional therapies, such as targeted drug delivery, sustained release, protection of labile groups
491 from degradation, low toxicity and increased drug specificity¹⁴⁸.

492 It is worth mentioning that, as the size of the nanoparticles is reduced, the maximum drug loading per
493 particle decreases. However, since the surface to volume ratio increases, the diffusion of the drug out
494 of the particles is favored as shown in **Figure 5**. Consequently, the burst release in polymer
495 nanoparticles is higher and overall, the release of the drug is mainly driven by diffusive phenomena.
496 In this context, biodegradable polyester-based NPs, using a core of PLA and its copolymers, represent
497 and interesting tool to achieve a sustained release of antiseptics compounds for oral or topical
498 applications and different studies are focusing on this aspect in the recent years.

499 Indeed, Baier *et al.* studied the enzyme-triggered release of OCT from PLA-based nanoparticles
500 stabilized by different surfactants. In this type of drug delivery systems, the release of the drug was
501 triggered only by the action of specific enzymes, which caused the degradation of the polymer matrix,
502 allowing a sustained antimicrobial activity of the formulation¹⁴⁹. However, the use of a significant
503 amount of surfactant may negatively influence the biocompatibility of the formulation. Furthermore,
504 the use of preformed polymers decreases the tunability of the particle characteristics.

505 In order to improve the different properties of the formulation as drug encapsulation efficiency and
506 degradation time of the polymeric matrix, copolymers of PLA with different monomers have been
507 investigated¹⁵⁰. In this context, the choice of the co-monomer is crucial in the characteristics of the
508 final formulation.

509 In fact, the possibility to copolymerize PLA with a hydrophilic compound (*e.g.* PEG) leads to an
510 increase of the drug loading efficiency and a higher control to the release of the active principle due
511 to the higher porosity of the colloidal system. Two studies report the use of self-assembled bi- and
512 tri-layered nanoparticles made of poly(ethylene glycol)-b-poly(lactic acid) (PEG-b-PLA) block
513 copolymers. Due to the presence of both hydrophilic and hydrophobic moieties, the particles could
514 effectively entrap CHX in the interior, showing higher degradation rates. Overall, through these
515 strategies, sustained antimicrobial activity up to 21 days were obtained^{34,151}.

516 Copolymers of PLA and PGA are also commonly studied as carriers for antiseptic delivery. Indeed,
517 PLGA was employed in different studies for the nanoencapsulation of CHX in restorative dentistry¹⁵².
518 Priyadarshini *et al.* prepared nanoparticles *via* emulsion/evaporation, with high drug entrapment
519 efficiency (up to 88%). The *in vitro* CHX release showed a bi-phasic trend, with an initial burst in the
520 first 3 hours followed by a slow progressive escape of the drug via a combination of diffusion and
521 polymer degradation up to 28 days. Moreover, exploiting the feature of the PLGA copolymer it is
522 possible to tune the release of the drug by varying the copolymer composition.

523 Nonetheless, other polyesters, *e.g.* PCL may be used in the synthesis of polymeric NPs. Indeed,
524 Lboutounne *et al.* developed a drug delivery system consisting of CHX encapsulated in PCL NPs for
525 the treatment of nosocomial infections. This drug delivery system was proposed as a substitute to
526 antiseptic topical formulations (hand-washing agents) that could lead to skin problems such as
527 dermatitis or allergies. The authors investigated the effect of the drug carrier on the porcine skin
528 compared with the traditional CDA solution. It was proved that CDA absorption in the skin was three
529 times higher from nanocapsules than from the corresponding solution. Thanks to their size, the
530 particles could adhere to bacterial surface, inhibiting the bacterial growth¹⁵³. However, the
531 formulation achieved a maximum encapsulation efficiency of 60 %, due to a low affinity between the
532 polymer and the drug.

533 Indeed, one of the main drawbacks that is limiting the application of polyester-based formulations is
534 the poor affinity to the cationic antiseptic, with its amphiphilic and electrostatic character. Due to

535 their molecular structure, the drug molecules tend to locate at the interface between the formulation
536 and the aqueous environment, leading to a fast release. Consequently, polymer functionalization able
537 to increase the affinity between the polymer and the charged antiseptic may significantly enhance the
538 drug loading in the formulation.
539

540 **4. Current Trends for the Control of the Antiseptic Concentration** 541 **within a Therapeutic Window**

542 One of the major issues of polyester devices in the controlled delivery of therapeutics is the so-called
543 “burst release”, where a large amount of drug is released in the first 24 hours^{154,155}. This fast delivery
544 of therapeutics results in possible toxicity for the patient, due to high drug concentrations and short
545 release profiles¹³⁴. While a strong antibacterial activity is required in the first hours after the surgery,
546 also a sustained antiseptic release over the targeted timeframe of sterility is necessary⁴⁹. In this
547 context, a variety of possible functionalizations can be carried out in order to enhance the interaction
548 between the polymer matrix and the drug, thus lowering the initial burst depending on the specific
549 application¹⁵⁶.

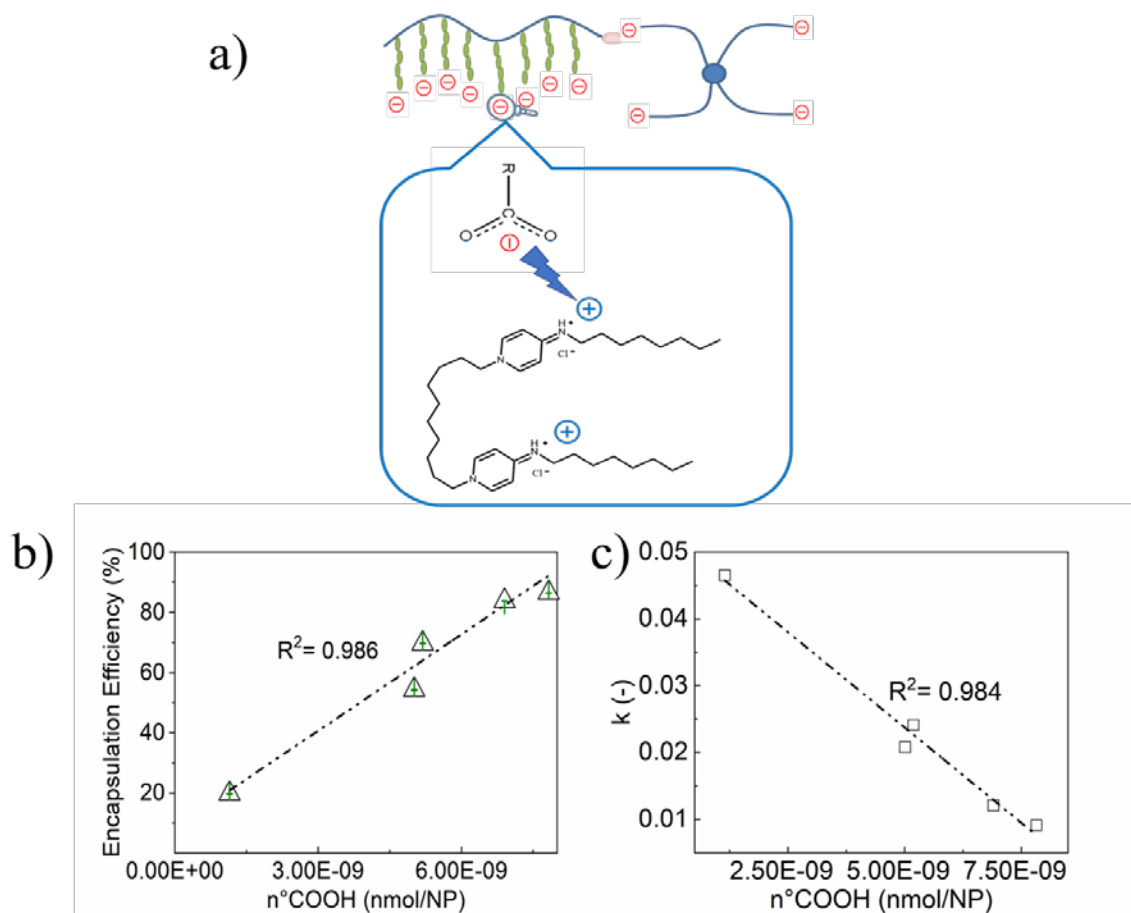
550 Biodegradable polyesters can be functionalized following post-polymerization or copolymerization
551 routes. In the former case the polymer is first synthesized and then enriched with functional groups
552 devoted to the retention of the drug. However, the possibility of racemization or scission of the ester
553 bonds as side reaction represent the main limit of this strategy. Therefore, a more frequently used
554 method involves the copolymerization of functional monomers with commercially available ones.
555 The moieties bearing functional groups are protected during the polymerization to avoid side
556 reactions, so a deprotection step is usually required to obtain functional polymers¹⁵⁷.

557 In this context, an efficient strategy employed to enhance the active ingredient loading and to slow
558 down the drug release is the conjugation between the therapeutic molecule and the pendant groups of
559 the polymer backbone^{134,158}. By changing the strength of the chemical bond it is possible to tune the
560 release rate depending on the application¹⁵⁹. Specifically, the presence of a covalent bond may allow
561 to achieve linear release kinetics. Indeed, Yoo *et al.* showed how the release of Doxorubicin-PLGA
562 conjugated nanoparticles was independent from drug diffusion but relied only on polymer
563 degradation, with a zero-order profile¹⁶⁰. A similar strategy may be envisioned for the controlled
564 delivery of antiseptics. However, a potential drawback of covalently binding the drug to the polymer
565 chain is the loss of biological activity¹⁶¹. Moreover, a further limitation of the technique is the need
566 for active groups both in the drug molecule and the polymer chains.

567 Starting from this strategy, modified antibacterial agents usually presenting functional groups such
568 as carboxy-, hydroxy- and amino- moieties may be functionalized through the covalent linkage to a
569 polymerizable monomer, mainly acrylic^{144,162–164}. In this way the final monomer is composed by a
570 ionizable end group, providing the desired therapeutic effect (*e.g.* ammonium, pyridinium), a
571 polymerizable group (*e.g.* methacrylate) and an alkyl chain spacer¹⁶⁵. For example, Beyth *et al.*
572 realized dental composites in which a quaternary ammonium compound was attached to a
573 polymerizable monomer. As the polymerization was carried out, the bacterial agent was incorporated
574 in the resin, thus avoiding its fast release. However, even if the antibacterial activity was sustained
575 for 3 months, antiproliferative effects were shown only against bacteria in contact with the
576 antibacterial molecules¹⁶⁵.

577 In order to overcome the issue related to the loss of biological activity, many groups, instead of
578 covalently binding the antiseptic to the polymer, exploited their electrostatic interaction relying on

579 oppositely charged species to obtain polymeric devices with the possibility of entrapping a large
580 quantity of drug^{137,166}. Indeed, the electrostatic interaction between the therapeutic and the polymer
581 allows the formation of complexes that can significantly reduce the release rate¹⁶⁶⁻¹⁶⁸. Most of the
582 times, because many polymers are neutral under physiological conditions, the functionalization with
583 ionic species is required to obtain charged polymeric chains. In this case, the drug release depends
584 not only on the electrostatic interaction with the polymer, but also on the strength of the bond
585 introduced by the charged functionality¹⁶⁹. Specifically, this type of functionalization proved to be
586 particularly efficient in the encapsulation and release of cationic amphiphilic compounds as QACs
587 and biguanides. For example, two groups reported the use of cyclodextrin as a functionalizing agent
588 to increase the affinity with the released antiseptic drugs, particularly benzalkonium chloride and
589 chlorhexidine. In the first study, CD is immobilized on cotton gauzes by using citric acid as a linker.
590 High encapsulation efficiencies of benzalkonium chloride could be obtained, due to the formation of
591 inclusion complexes with CDs. Moreover, being the drug positively charged, an ionic interaction with
592 free carboxylic groups of citric acid was observed, thus increasing the loading values. The release of
593 benzalkonium chloride was sustained for 2 weeks, with higher antibacterial properties with respect
594 to commercial products¹⁷⁰. The second study instead, employs the same strategy to prolong the release
595 of chlorhexidine from cellulosic substrates¹⁷¹. Finally, it is worth mentioning that the presence of the
596 electrostatic interaction is able to significantly improve the drug loading also for synthetic polyester-
597 based formulations containing cationic antiseptics. For example, some studies on PLGA
598 microparticles reported the influence of polymer end groups on OCT release¹⁷². Indeed, deprotonated
599 carboxylic groups are able to establish an electrostatic interaction with the drug once in an aqueous
600 environment, which prevents the fast diffusion of the amphiphilic active principle outside the
601 polymeric carrier. Moreover, the application of this functionalization to NPs composed of complex
602 polyester architectures (*e.g.* star-shaped and brush-like structures) allowing to modulate the number
603 of carboxylate groups in the carrier, univocally showed that there is a linear correlation between the
604 drug encapsulation efficiency and the number of anionic groups, as shown in **Figure 5**¹⁷³.
605
606



607
608 *Figure 6: Mechanism of electrostatic interaction between negatively charged polyesters of different structure and the*
609 *positively charged octenidine dihydrochloride (a); dependency of encapsulation efficiency (b) and kinetic constant (c)*
610 *from the number of ionizable groups per particle. Reproduced with permission from¹⁷³.*

611

612 This is counterbalanced by the enhanced release rate of the drug for these types of systems. In fact,
613 PLGA with free carboxyl termini is more hydrophilic at physiological pH and it can absorb more
614 water compared to capped species with esterified termini^{103,141,174,175}.

615 Overall, the polymer-drug electrostatic interaction showed to be a promising route to the
616 encapsulation and release of QACs and biguanides since it allows at the same time a higher
617 encapsulation efficacy and a controlled release due to the degradation of the polymeric matrix that
618 occurs in the timespan required for the application. Moreover, the presence of an acidic end group in
619 the polymeric chain should not influence the polymer toxicity. In fact, from different tests performed
620 both *in vivo* and *in vitro* the comparison between the polymers functionalized with acidic end groups
621 and their ester counterparts (i.e. RESOMER RG H and S) shows the absence of a significant
622 difference in the cellular toxicity between these polymers^{176,177}. Since, the ester PLA and PLGA-
623 based formulations are already present in the market, these preliminary results may pave the way to
624 the clinical application of these functionalized materials also for the release of positively charged
625 antiseptics. In this context, the use of these polymers as excipients for biguanides and quaternary
626 ammonium salts allows their application for a wide range of HCAIs. Among these, they may be
627 particularly beneficial in the case of wound dressing. In fact, the possibility to use them in synergy
628 with the aforementioned natural polymers could promote wound healing and exudate adsorption
629 while maintaining the tunability and mechanical properties of polyesters. Finally, the carboxylic
630 functionalization could provide a sustained drug release over time, thus addressing the issue regarding
631 the dressing change frequency. Indeed, the possibility to reduce the number of medical treatments

632 through a sustained and control release of the drug from the polymeric excipients may decrease the
633 pressure and the costs on the national health systems while reducing the risks of complications for
634 the patient¹⁷⁸.

635

636 **5. Conclusions**

637 Surgical site infections represent one of the most urgent problems to be solved in healthcare. At the
638 same time, the increase in the number of bacterial strains resistant to antibiotics led to the
639 consideration of antiseptics to fight against this type of infections. However, the use of antiseptics,
640 most of them charged at physiological conditions, brings about the necessity of providing an efficient
641 encapsulation in polymer excipients and their long-lasting release in order to avoid the bacterial
642 colonization of the site even after hospitalization. Based on this, polymeric carriers are promising
643 tools thanks to their versatile characteristics provided by the polymer structure. Specifically, in this
644 review the most promising natural and synthetic polymers were discussed as drug delivery systems.
645 While the formers are characterized by an enhanced biocompatibility, the versatility provided by the
646 synthetic polymers, especially polyesters, able to couple it with their biodegradability proved to be a
647 key point in the production of efficient formulations. Moreover, the combination of the advantages
648 given by the synthesis of polyesters with specific functionalities able to enhance the affinity between
649 the excipient and the drug may represent a crucial aspect to significantly increase the performances
650 of the final formulations. Among these, regarding the categories of quaternary ammonium
651 compounds and biguanides analyzed in this review, the electrostatic interaction between the
652 positively charged drug and the polymer showed to be a promising feature to be exploited in the future
653 in order to improve the treatment of these infections.

654

655 **6. References**

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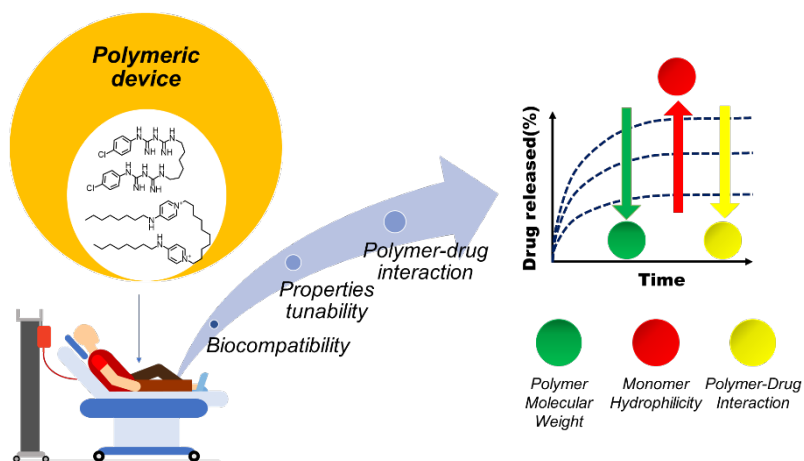
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1221 **Table of Contents**



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1224 This review provides the guidelines to efficiently produce a polymeric formulation able to
1225 guarantee a sustained release of positively charged antiseptic for the long-term sterility in the
1226 treatment of health care-associated infections.

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