

Review

Polymeric nanogels and hybrid platforms for redox modulation from ROS biology to therapeutic design

Giuseppe Nunziata,¹ Kotaro Fujishiro,² Greta Avancini,³ Ryan M. Porter,⁴ Roy Morello,⁵ Yuta Kurashina,² Ehsan Nazarzadeh Zare,^{6,7,8} Paolo Decuzzi,³ Hassan Karimi-Maleh,^{9,*} and Filippo Rossi^{1,*}

¹Department of Chemistry, Materials and Chemical Engineering “Giulio Natta”, Politecnico di Milano, Via Bassini 6, 20133 Milan, Italy

²Institute of Engineering, Tokyo University of Agriculture and Technology, 2-24-16 Nakamachi, Koganei City, Tokyo 184-8588, Japan

³Laboratory of Nanotechnology for Precision Medicine, Fondazione Istituto Italiano di Tecnologia, Via Morego 30, 16163 Genova, Italy

⁴Department of Orthopedic Surgery and Rehabilitation, University of Arkansas for Medical Sciences, 4301 W. Markham Street, Little Rock, AR 72205, USA

⁵Department of Physiology & Cell Biology, University of Arkansas for Medical Sciences, 4301 W. Markham Street, Little Rock, AR 72205, USA

⁶School of Chemistry, Damghan University, Damghan 36716-45667, Iran

⁷Centre for Research Impact and Outcome, Chitkara University Institute of Engineering and Technology, Chitkara University, Rajpura, Punjab, 140401, India

⁸Center for Theoretical Physics, Khazar University, 41 Mahasti St, Baku, AZ1096, Azerbaijan

⁹Zhejiang Province Engineering Research Center for Endoscope Instruments and Technology Development, Quzhou People's Hospital, The Quzhou Affiliated Hospital of Wenzhou Medical University, 2 Zhongludi, Kecheng District, Quzhou City 324000, Zhejiang Province, China

*Correspondence: h.karimi.maleh@gmail.com (H.K.-M.), filippo.rossi@polimi.it (F.R.)

<https://doi.org/10.1016/j.xcrp.2026.103446>

SUMMARY

Reactive oxygen species (ROS) play a dual role in biological systems. At physiological levels, they act as essential signaling mediators regulating immune responses, metabolism, proliferation, and differentiation. Conversely, excessive ROS production overwhelms antioxidant defenses, driving oxidative stress, inflammation, and tissue damage. Understanding the generation, spatial confinement, and kinetics of ROS is therefore critical for designing effective therapies. Conventional small-molecule scavengers show limited translational success due to poor stability, rapid clearance, and lack of targeting, while synthetic antioxidants, although more versatile, still face delivery constraints. In this context, polymeric nanogels provide hydrated nanoscale networks capable of protecting labile scavengers, hosting catalytic redox centers, and responding dynamically to ROS-rich microenvironments. However, their standalone application is often limited by insufficient retention in dynamic tissues. To overcome these limitations, hybrid systems integrating nanogels within hydrogels or composite matrices have emerged, enabling improved localization and sustained redox modulation.

INTRODUCTION

Reactive oxygen species (ROS) are a category of highly reactive molecules derived from elementary oxygen.¹ These oxidants can be in the form of free radicals, like superoxide ($O_2^{\cdot-}$), hydroxyl radicals ($\cdot OH$), and peroxy radicals ($ROO\cdot$), as well as non-radical compounds, such as hydrogen peroxide (H_2O_2) and singlet oxygen molecules (1O_2).² They can be triggered by endogenous (mitochondrial and cellular metabolism or through inflammation) or exogenous (environmental or by delivery of nanomaterial) stimuli.³ Depending on their concentration in living organisms, they can have dual roles.⁴ At a low level, their presence is beneficial for proper physiological processes as signaling molecules that regulate, for example, immune response and cell growth.⁵ Through highly specific and reversible oxidation of protein cysteine residues, particularly the conversion of $Cys-S^-$ to sulfenic acid ($Cys-SOH$), ROS modulate the structure and activity of key regulatory proteins.⁶ These redox-dependent modifica-

tions influence transcriptional programs, phosphorylation cascades, metabolic fluxes, and multiple intracellular signaling pathways.^{7–9} Controlled ROS production is therefore required for optimal immune response activation, where an increase in mitochondrial or nicotinamide adenine dinucleotide phosphate oxidase (NOX)-derived ROS promotes the release of pro-inflammatory cytokines, including interleukin- 1β (IL- 1β), tumor necrosis factor alpha (TNF- α), and interferon- β (IFN- β), enabling adequate host defense.¹⁰ Similarly, finely tuned redox signals regulate cell proliferation and differentiation. Cancer cells and several stem cell populations maintain slightly elevated ROS concentrations to activate pathways involving hypoxia-inducible factors (HIFs), phosphatidylinositol 3-kinase (PI3K), nuclear factor κB (NF- κB), and mitogen-activated protein kinases (MAPKs), thereby supporting self-renewal and growth.¹¹ In stem cells, low ROS levels preserve proliferative potential, whereas higher levels activate ataxia-telangiectasia mutated (ATM)-dependent checkpoints that limit renewal.¹²



The therapeutic goal of ROS-modulating biomaterials should not be interpreted as complete ROS elimination. Physiological ROS are essential signaling mediators involved in immune activation, metabolic regulation, proliferation, differentiation, angiogenesis, and tissue remodeling. Therefore, excessive or non-selective ROS depletion may impair host defense, delay tissue repair, alter stem cell behavior, or interfere with redox-dependent signaling cascades. This consideration is particularly relevant for catalytic scavengers, whose activity may persist beyond the initial oxidative insult.¹³ Future polymeric systems should therefore be designed to restore redox homeostasis within a defined therapeutic window rather than to suppress ROS indiscriminately. Spatial confinement, disease-triggered activation, degradable architectures, and tunable catalytic or sacrificial capacity are key parameters to minimize interference with physiological signaling while preserving therapeutic antioxidant efficacy. Overall, these observations establish ROS not only as damaging oxidants but as indispensable regulators of cellular homeostasis, proliferation, immunity, and development, provided that their concentrations remain within the physiological redox-signaling range. When antioxidant defenses fail to maintain redox equilibrium, ROS accumulate beyond the physiological threshold and shift from signaling mediators to cytotoxic agents.¹⁴ Under these conditions, highly reactive species, such as hydroxyl radicals, can directly oxidize DNA, RNA, proteins, and membrane lipids, compromising structural integrity and impairing essential cellular functions.¹⁵ The degree of damage strongly depends on the specific reactivity of each ROS and on the availability of labile Fe^{2+} , which catalyzes hydroxyl-radical formation through the Fenton reaction.¹⁶ This iron-dependent amplification of oxidative reactions represents one of the major determinants of ROS-induced toxicity.¹⁷

Although mammalian cells possess an extensive antioxidant network (including catalase, peroxidases, and glutathione-based systems) these defenses become insufficient when ROS production outpaces scavenging capacity.¹⁸ The resulting accumulation of oxidized nucleic acids and proteins must be counteracted by repair pathways or removed via proteasomal degradation and autophagy.¹⁹ When these compensatory mechanisms are overwhelmed, cellular injury ensues, and oxidative stress becomes a driving force in pathological inflammation, promoting cytokine release and accelerating tissue degeneration.²⁰ Interestingly, the extreme resilience of organisms such as *Deinococcus radiodurans* illustrates that ROS toxicity is not solely dictated by the amount of oxidants but by the cell's ability to control iron homeostasis, limit endogenous ROS formation, efficiently scavenge radicals, and repair or replace damaged biomolecules.²¹ These systems enable some microorganisms to survive even extraordinarily high ROS levels.²² In mammals and plants, similar principles apply; ROS-induced cellular damage is tightly modulated by antioxidative enzymes, iron-sequestering mechanisms, and robust DNA/protein repair pathways.^{23,24} When these layers of protection fail, ROS no longer act as metabolic signals but instead trigger regulated death programs such as ferroptosis or necrosis-like pathways rather than simply causing passive oxidative collapse.^{25,26} Excessive ROS accumulation shifts the system from physiological redox signaling to oxidative stress, leading to macromolecular damage and triggering inflammatory

cascades that aggravate disease progression.²⁷ This redox imbalance has motivated intense research into biomaterials able to neutralize ROS or restore homeostasis.^{28–30} In recent years, ROS-scavenging technologies have gained growing attention due to their ability to attenuate oxidative injury.³¹ Numerous studies have demonstrated the antioxidant efficiency of these materials against oxidative stress as a crucial pathological driver.^{14,32}

Targeting ROS provides two complementary opportunities. On one hand, scavenging excessive oxidants can restore redox balance, attenuate inflammation, and protect vulnerable cells, including stem cells, which are particularly sensitive to oxidative microenvironments.³³ On the other hand, the abnormally high ROS levels characteristic of inflamed or tumor tissues can be harnessed for selective and on-demand activation of therapeutic agents, enabling more precise drug delivery while minimizing off-target toxicity.³⁴ Polymeric nanotechnologies have rapidly emerged as ideal platforms to implement both strategies.³⁵ Nanoparticles, nanogels, and hydrogels can be engineered to neutralize ROS through catalytic or antioxidant functionalities or to undergo ROS-triggered degradation and drug release via cleavable linkers such as thioketals or boronic esters.^{36–38} These smart systems enhance the solubility, stability, and circulation time of therapeutics while providing precise spatiotemporal control over drug activation.³⁹ Their modular design allows them to function as protective carriers, diagnostic tools, or ROS-responsive prodrugs capable of activating exclusively within oxidatively stressed tissues.⁴⁰ For these reasons, polymer-based nanoplat-forms are emerging as some of the most powerful strategies to modulate, exploit, or correct ROS-driven biological processes.⁴¹ However, despite rapid progress, the field remains fragmented, with ROS-scavenging systems, ROS-responsive prodrugs, and polymeric nanoarchitectures often discussed separately without a unified framework linking their mechanisms, design principles, and therapeutic potential.^{42,43} This review addresses this gap by providing a comprehensive and integrated overview of polymeric nanogels, hydrogels, and hybrid systems engineered either to neutralize excessive ROS or to harness pathological oxidative environments for precise, spatiotemporally controlled drug delivery (Figure 1).

We highlight converging design strategies, structure-property relationships, and emerging biomedical applications across inflammation, cancer, and regenerative medicine, offering a consolidated perspective that is increasingly needed to guide future development of next-generation ROS-targeted therapies.

ROS IN MAJOR HUMAN DISEASES

Given the pervasive involvement of ROS across a wide spectrum of pathological conditions, an effective redox-oriented therapeutic strategy requires a precise understanding of the nature of the oxidant species involved, their spatiotemporal dynamics, and the molecular mechanisms through which they drive tissue damage.⁴⁴ Accordingly, the following section outlines how distinct ROS and redox-dependent pathways contribute to major human diseases, not as an exhaustive pathological survey but as a conceptual framework to inform the rational design of polymer-based nanogels, hydrogels, and hybrid systems tailored to

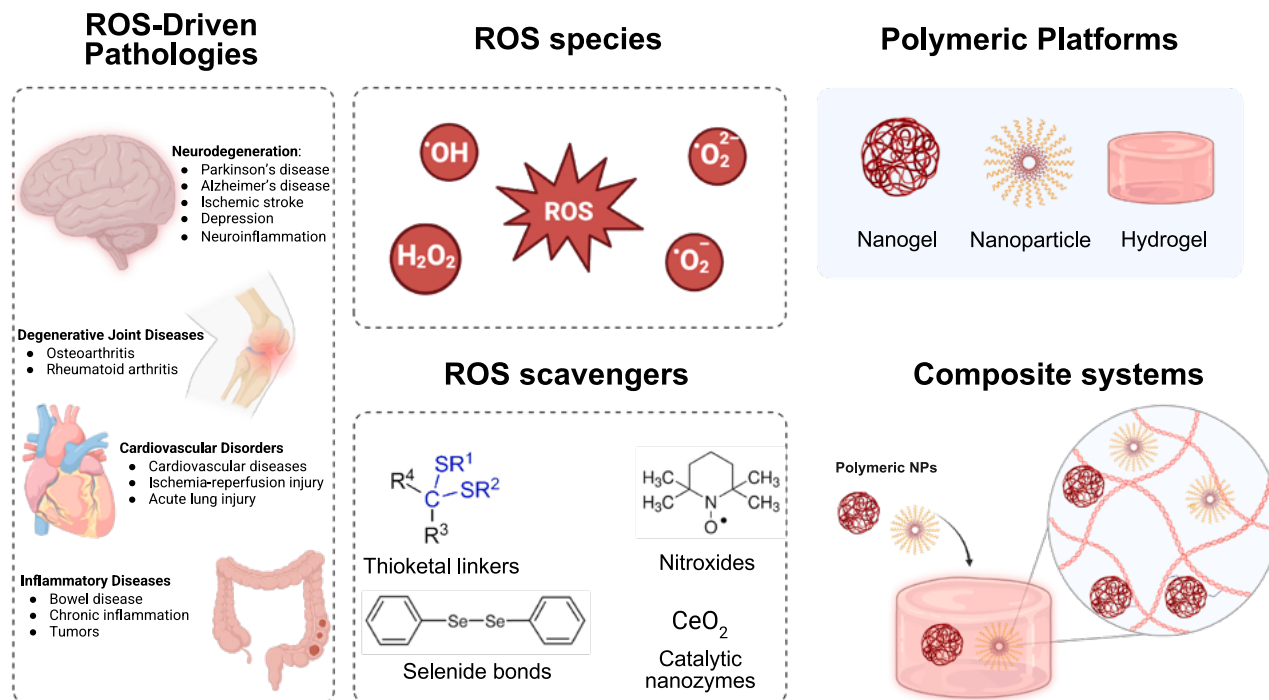


Figure 1. Overview of ROS-driven pathologies and polymeric platforms for redox modulation

Schematic representation of the main elements discussed in the review, including ROS-associated pathological conditions, representative ROS species, ROS-scavenging chemistries, and polymeric delivery platforms.

The figure summarizes how nanogels, nanoparticles, hydrogels, and composite systems can be engineered with antioxidant or ROS-responsive motifs, such as thioketal linkers, nitroxides, selenide bonds, and catalytic nanozymes, to modulate oxidative stress in disease-relevant microenvironments.

specific oxidative microenvironments. ROS are highly reactive oxidants generated during cellular metabolism and in response to environmental cues.⁴⁵ When their production exceeds the buffering capacity of endogenous antioxidant systems, oxidative stress arises, leading to damage of nucleic acids, proteins, and membrane lipids.⁴⁶ This redox imbalance disrupts signaling networks, sustains inflammation, and directly contributes to the initiation and progression of a broad spectrum of human diseases, including neurodegenerative, inflammatory, metabolic, skeletal, and cardiovascular disorders.⁴⁷ Beyond nonspecific macromolecular damage, excessive ROS actively drive regulated cell death programs.⁴⁸

Ferroptosis is a paradigmatic example of ROS-dependent cytotoxicity, relying on iron-catalyzed lipid peroxidation rather than caspase activation.^{49,50} The abstraction of hydrogen atoms from polyunsaturated phospholipids initiates self-propagating lipid radical chain reactions that culminate in membrane destabilization.⁵¹ Under physiological conditions, enzymatic antioxidants and lipid-soluble scavengers restrain this process; however, glutathione depletion, glutathione peroxidase 4 (GPX4) dysfunction, or impaired cystine uptake via system Xc[−] abolish peroxide detoxification, while expansion of the labile iron pool accelerates radical formation.⁵² These converging events establish ferroptosis as a central mechanism linking oxidative stress to tissue degeneration across multiple pathological contexts.⁵³ Neurodegenerative disorders exemplify the pathological consequences of sustained redox imbalance.^{54,55} In Alzheimer's dis-

ease, Parkinson's disease, and related dementias, high neuronal oxygen consumption combined with limited antioxidant capacity promotes the accumulation of ROS and reactive nitrogen species, impairing mitochondrial function and proteostasis.⁵⁶ In Parkinson's disease, iron deposition in the substantia nigra amplifies oxidative injury and lipid peroxidation, while ROS-mediated modification of α -synuclein enhances its aggregation.⁵⁷ Aggregated α -synuclein further compromises mitochondrial respiration, generating a self-reinforcing cycle of oxidative stress, energetic failure, and neuronal loss.⁵⁸ Similar interactions between iron dyshomeostasis, mitochondrial dysfunction, and oxidative injury are observed across neurodegenerative conditions.⁵⁹ Chronic inflammatory diseases are likewise characterized by persistent, localized oxidative stress.⁶⁰ In osteoarthritis and rheumatoid arthritis, excessive ROS production within joint tissues impairs mitochondrial function, induces chondrocyte apoptosis, and sustains inflammatory signaling.^{61–64} Upregulation of nicotinamide adenine dinucleotide phosphate (NADPH) oxidase isoforms and insufficient antioxidant responses create a redox microenvironment that promotes cartilage degradation and synovial damage.^{65–67}

Importantly, disease severity correlates with oxidative dysregulation, underscoring ROS as active drivers rather than secondary byproducts of inflammation.⁶⁸ Metabolic disorders further illustrate the pathological impact of prolonged oxidant overload.⁶⁹ In diabetes, chronic hyperglycemia enhances mitochondrial ROS generation and activates redox-sensitive pathways,

damaging endothelial cells, reducing nitric oxide bioavailability, and accelerating the formation of advanced glycation end products.⁷⁰ These processes contribute to microvascular complications, including nephropathy, neuropathy, and retinopathy, while excessive ROS also impair fibroblast migration and angiogenesis, delaying wound healing.⁷¹ Similarly, in chronic inflammatory bowel diseases, excessive oxidant production by intestinal epithelial cells and infiltrating immune cells overwhelms local antioxidant defenses.⁷² ROS and reactive nitrogen species damage mitochondrial enzymes, oxidize lipids, and activate NF- κ B and MAPK pathways, sustaining cytokine release, epithelial barrier dysfunction, and chronic inflammation, ultimately increasing the risk of colitis-associated colorectal cancer.⁷³ Across these diverse pathological settings, a common theme emerges: ROS-driven damage is spatially and temporally confined to diseased tissues, sustained over time, and poorly addressed by systemic antioxidant supplementation. These features highlight the need for strategies capable of locally modulating redox homeostasis, buffering excessive oxidants, and responding dynamically to pathological ROS levels, thereby providing a strong rationale for the development of polymer-based nanogels, hydrogels, and hybrid systems designed to scavenge or exploit ROS. Despite the central role of ROS in disease, direct comparison across studies remains challenging due to the intrinsic heterogeneity of ROS detection methods.⁷⁴

Many commonly used assays rely on indirect or proxy readouts, such as fluorogenic probes or downstream oxidative markers, which differ in sensitivity, specificity, and temporal resolution.⁷⁵ As a result, measured ROS levels often reflect integrated oxidative stress rather than individual ROS species or localized redox dynamics.⁷⁶ This methodological variability has direct consequences for the interpretation of therapeutic efficacy. Intracellular probes such as dichloro-dihydro-fluorescein diacetate (DCFH-DA) are widely used to estimate global oxidative activity, but their oxidation is indirect; can be influenced by peroxidases, metal ions, light exposure, and cellular uptake; and does not selectively identify a single ROS species.⁷⁷ Similarly, dihydroethidium (DHE) and mitochondrion-targeted derivatives such as MitoSOX are commonly used to assess superoxide production, but their interpretation requires caution because non-specific oxidation products can be generated unless appropriate analytical controls are included.⁷⁸ Cell-free assays such as 2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid) (ABTS) and 2,2-diphenyl-1-picrylhydrazyl (DPPH) provide rapid screening of electron- or hydrogen-donating antioxidant capacity, but they are performed under simplified chemical conditions that do not reproduce the spatial confinement, enzymatic regulation, protein corona formation, diffusion barriers, or continuous ROS fluxes present in biological tissues.⁷⁹ Conversely, lipid peroxidation readouts such as malondialdehyde (MDA) and thiobarbituric acid reactive substances (TBARS) report downstream oxidative damage rather than the primary ROS species responsible for material activation and can be affected by sample preparation, secondary reactions, and matrix-dependent artifacts.⁸⁰ Therefore, a reduction in DCF fluorescence, MDA/TBARS levels, or ABTS/DPPH absorbance should not automatically be interpreted as direct and species-specific ROS neutralization.⁸¹ Future studies should combine complementary readouts,

including species-oriented probes, cell-free antioxidant tests, lipid/protein oxidation markers, inflammatory biomarkers, and direct measurement of material degradation or release kinetics.

Importantly, assay conditions should be reported in detail, including ROS source, oxidant concentration, exposure time, probe concentration, calibration strategy, and biological model, to enable meaningful comparison among redox-responsive nanogels and hybrid platforms. In this context, it is also important to recognize that ROS are not interchangeable therapeutic triggers. Although they are often discussed collectively, their biological effects and suitability for material activation depend strongly on chemical identity, lifetime, diffusion range, and subcellular localization.⁸² $O_2^{\cdot-}$ is primarily generated by mitochondria and NADPH oxidases and has a short lifetime and limited diffusion because of its charged nature and rapid dismutation.⁸³ It therefore acts mainly near its site of production and is particularly relevant for catalytic systems able to mimic superoxide dismutase activity, such as manganese-based complexes, cerium oxide, Prussian blue, and nitroxide-bearing materials. H_2O_2 is the most quantitatively tractable species because it is comparatively more stable than radical ROS, can diffuse across biological membranes, and acts as a central mediator of redox signaling.⁸⁴ Physiological H_2O_2 -dependent signaling is generally associated with low steady-state concentrations in the nanomolar range, commonly reported to be around 1–10 nM, whereas higher concentrations, above approximately 100 nM, are typically associated with adaptive stress responses or oxidative distress.⁸⁵ However, even for H_2O_2 , these values should be interpreted as order-of-magnitude references rather than universal thresholds because the concentration experienced by a molecular target depends on the compartment; distance from the source; local antioxidant buffering; membrane permeability; enzymatic sinks such as peroxiredoxins, glutathione peroxidases, and catalase; and the analytical method used for detection.⁸⁶ For shorter-lived oxidants, this limitation is even more pronounced. $\cdot OH$ are extremely reactive and react almost immediately at the site where they are formed, typically through Fenton-type chemistry involving labile iron or copper. Therefore, direct “targeting” of $\cdot OH$ by a material is intrinsically difficult; more realistic strategies aim to prevent its upstream formation by reducing H_2O_2 availability, chelating redox-active metals, or introducing radical-quenching motifs such as nitroxides or polyphenols. Lipid peroxides and lipid-derived radicals are instead spatially confined to membranes and lipoprotein-rich domains and are central mediators of ferroptosis, making lipophilic radical-trapping antioxidants, selenium-containing motifs and GPX-like catalytic systems more appropriate than generic hydrophilic scavengers.⁸⁷ Finally, peroxynitrite ($ONOO^-$), generated from the diffusion-controlled reaction between $O_2^{\cdot-}$ and nitric oxide, combines oxidative and nitrative reactivity and is particularly relevant in inflammatory and neuroinflammatory environments, where oxidative and nitrosative stress coexist; in this case, dual ROS/reactive nitrogen species (RNS)-scavenging chemistries may be required.⁸⁸ From a materials design perspective, this species-dependent behavior has two important consequences. First, polymeric platforms should not be described generically as “ROS responsive” without specifying the oxidant species, concentration range, exposure time, and detection method

used to demonstrate responsiveness. Many polymer degradation or release studies employ exogenous H_2O_2 challenges in the micromolar to millimolar range to accelerate cleavage, swelling, or payload release.^{89,90} These assays are useful to prove chemical oxidizability, but they do not necessarily reproduce physiological redox signaling or the local oxidative microenvironment of diseased tissues. Second, the selection of the responsive chemistry should reflect the spatial and temporal properties of the target oxidant. Catalytic systems are more suitable for sustained and continuously regenerated oxidative stress, whereas sacrificial chemistries such as thioketals, boronic esters, thioethers, selenides, and diselenides are more appropriate when oxidative bursts are intended to trigger network degradation or drug release.

Therefore, the rational design of ROS-modulating polymeric systems requires matching oxidant identity, local concentration, lifetime, compartmentalization, and material-response kinetics rather than relying only on broad antioxidant capacity.

MOLECULES WITH ROS-SCAVENGING ACTIVITY

Understanding the mechanisms through which ROS-scavenging and ROS-responsive molecules operate is essential to rationally design and functionalize polymeric nanosystems. Identifying how different chemical motifs interact with specific oxidant species provides the foundation for tailoring nanogels, hydrogels, and hybrid platforms capable of modulating pathological redox microenvironments. ROS scavengers comprise a wide spectrum of molecules capable of neutralizing oxidative stress, restoring redox balance, and limiting cellular injury. Classically, they are divided into endogenous and exogenous antioxidants.⁹¹ Endogenous scavengers constitute the intrinsic defense network of the organism and include enzymatic systems such as superoxide dismutase, catalase, and glutathione reductase as well as non-enzymatic components such as glutathione, bilirubin, uric acid, melatonin, polyamines, and metal-binding proteins.^{92,93} Exogenous antioxidants are predominantly obtained from the diet and encompass vitamins A, C and E together with polyphenols, carotenoids, flavonoids, and trace minerals.⁹⁴ Beyond physiological and nutritional defenses, a broad class of synthetic ROS scavengers has gained increasing relevance in biomedical engineering.⁹⁵ These compounds are rationally designed to mimic or enhance natural antioxidant functions, with mechanisms that depend on their molecular structure, redox activity, and catalytic properties. Based on their composition, synthetic scavengers can be broadly categorized into inorganic materials and organic small molecules.^{96,97}

Importantly, this distinction is not merely chemical but reflects fundamentally different modes of ROS interaction, stability, and turnover, which are directly relevant for their integration into nanoscale delivery systems.⁹⁸ Inorganic scavengers introduce a level of catalytic robustness and redox tunability that extends beyond classic organic antioxidants.⁹⁹ Manganese-based compounds are a well-established family of catalytic scavengers in which porphyrin ligands stabilize the $\text{Mn}^{2+}/\text{Mn}^{3+}$ redox cycle, enabling efficient superoxide dismutation and peroxide reduction.¹⁰⁰ Cationic manganese porphyrins have been shown to attenuate oxidative stress and apoptosis in models of spinal

cord injury, although without full functional recovery.¹⁰¹ A related compound, $\text{MnTnBuOE-2-PyP}^{5+}$ (BMX-001), demonstrated pronounced anti-inflammatory and antioxidant effects in ischemic stroke by suppressing macrophage-driven oxidative bursts with minimal local toxicity, highlighting the translational potential of catalytic metal-based scavengers.¹⁰² Among inorganic nanomaterials, Prussian blue nanoparticles exhibit enzyme-mimetic activities resembling superoxide dismutase, catalase, and peroxidase, enabling broad-spectrum ROS detoxification.¹⁰³ These materials significantly reduce lipid peroxidation and suppress inflammatory mediators such as TNF, IL-1 β , IL-6, and inducible nitric oxide synthase (NOS2), protecting tissues from oxidative injury.¹⁰⁴ Similarly, cerium oxide nanoparticles have emerged as highly effective synthetic scavengers owing to their reversible $\text{Ce}^{3+}/\text{Ce}^{4+}$ redox cycling, sustained by oxygen vacancies within the crystalline lattice.¹⁰⁵ This self-regenerating redox behavior enables catalytic neutralization of superoxide, hydrogen peroxide, and hydroxyl radicals, conferring prolonged activity compared to conventional antioxidants, which are rapidly consumed under oxidative stress.¹⁰⁶ In parallel, organic antioxidants encompass a diverse group of small molecules capable of directly quenching radical species or indirectly restoring redox balance through metabolic and signaling pathways.¹⁰⁷

Nitroxides such as (2,2,6,6-tetramethylpiperidin-1-yl)oxyl (TEMPO) efficiently neutralize hydroxyl radicals, superoxide, and hydrogen peroxide via a superoxide dismutase-like catalytic cycle without being irreversibly consumed.¹⁰⁸ However, their clinical application is limited by rapid diffusion and a short biological half-life.^{109,110} To overcome these limitations, strategies such as tethering TEMPO to biomaterials or confining it within polymeric matrices have been developed.¹¹⁰ For instance, TEMPO-functionalized hydrogels engineered for cardiovascular implants enable sustained nitroxide retention, reduce hydroxyl radical accumulation, and attenuate pro-inflammatory cytokine secretion, illustrating how material integration can extend antioxidant efficacy.¹¹¹ Subcellular targeting further enhances nitroxide performance.¹¹² Mito-TEMPO, designed to accumulate within mitochondria, mitigates oxidative damage associated with severe hypoglycemia by preserving mitochondrial function, reducing blood-brain barrier leakage, and improving ATP production.¹¹³ Its derivative Tempol, characterized by enhanced water solubility and biocompatibility, has demonstrated protective activity in osteoarthritis by reducing cartilage degradation, restoring endogenous antioxidant enzyme activity, and suppressing inflammatory cytokines.¹¹⁴ Tempol has also shown neuroprotective effects in cisplatin-induced cerebellar toxicity, where it limits neuronal loss and preserves synaptic organization.¹¹⁵ Recent advances extend beyond single-function scavengers toward dual-action molecules capable of neutralizing both ROS and reactive nitrogen species. These systems combine antioxidant and NO-scavenging moieties within a single molecular framework, enabling more effective attenuation of oxidative and nitrosative stress.¹¹⁶ In selenized and thiolated systems encapsulating *trans*-resveratrol for gastrointestinal applications, selenium and thiol groups participate in redox cycles that directly scavenge peroxides and peroxynitrite, reducing inflammation and cytotoxicity in both cellular and animal models.^{117,118}

Table 1. Quantitative benchmarking of representative ROS-scavenging and ROS-responsive systems

System	Strategy	Assay/model	Experimental conditions	Quantitative benchmark	Comment
MnTnBuOE-2-PyP ⁵⁺ /BMX-001 manganese porphyrin ¹⁰²	catalytic ROS/RNS-scavenging redox modulator	mouse intracerebral hemorrhage model	BMX-001 administered immediately after collagenase-induced ICH; analysis at 72 h	Macrophages in hemorrhagic area decreased from 48 ± 10 to 33 ± 8 cells; Prussian-blue-positive cells decreased from 65 ± 22 to 41 ± 15 .	<i>In vivo</i> anti-inflammatory/iron-associated oxidative injury endpoint, not direct ROS-scavenging potency.
poly(DMA-co-TEMPO) copolymer, 40T ¹⁰⁹	polymeric ROS-scavenging system	carrageenan-induced air pouch model	5 mg polymer/animal; TEMPO dose matched at 8.48 $\mu\text{mol}/\text{animal}$	After 6 h, retention was 0% for Tempol, ~12% for 40T, and ~94% for 100T. 40T reduced O ₂ ⁻ and TNF- α levels in the air pouch by ~90% and ~83%, respectively.	Shows that optimized TEMPO density, rather than maximum TEMPO loading, controls <i>in vivo</i> efficacy.
TEMPO-loaded hydrogel-modified pericardium, PSD5:5GM-TEMPO/VEGF ¹¹⁰	ROS-responsive release + ROS scavenging	H ₂ O ₂ -triggered TEMPO release and Fenton-based OH scavenging	TEMPO released under 10 μM H ₂ O ₂ ; OH scavenging measured using Fenton reaction	TEMPO loading capacity: $10.84 \pm 3.79 \mu\text{g cm}^{-2}$. Under 10 μM H ₂ O ₂ , cumulative TEMPO release reached ~60% at 96 h and ~85% at 12 days. Maximum OH scavenging approached ~70%.	Combines mild H ₂ O ₂ -triggered release with radical scavenging.
Tempo-FPF Janus hydrogel ¹¹¹	ROS-scavenging hydrogel	DPPH, ABTS, and OH scavenging assays	Hydrogel concentration: 18 mg mL ⁻¹ ; DPPH at 37°C for 60 min; ABTS for 10 min; OH at 37°C for 30 min	DPPH scavenging: $77.8 \pm 0.1\%$; ABTS scavenging: 98.2% ; OH elimination: $81.3 \pm 0.2\%$. PFP control: $19.9 \pm 0.8\%$, 24.0% and $24.9 \pm 0.6\%$, respectively.	Strong cell-free antioxidant benchmark.
Selenized thiolated alginate nanogel loaded with <i>trans</i> -resveratrol, Ngel-3@Res ¹¹⁷	ROS-scavenging nanogel	cell-free OH, DPPH, and ABTS assays after simulated gastrointestinal digestion	OH assay based on Fenton reaction; DPPH and ABTS radical assays; <i>C. elegans</i> oxidative stress model with 40 mM H ₂ O ₂	After digestion, Ngel-3@Res showed OH clearance of $99.11\% \pm 0.61\%$, DPPH clearance of $55.11\% \pm 0.56\%$, and ABTS clearance of $59.88\% \pm 0.80\%$. Mean survival under H ₂ O ₂ stress increased from 2.00 ± 0.36 h to 3.20 ± 0.32 h.	Good radical-scavenging benchmark, but cell-free and biological survival assays are not directly comparable.

(Continued on next page)

Table 1. Continued

System	Strategy	Assay/model	Experimental conditions	Quantitative benchmark	Comment
Celecoxib-loaded CMCs/Alg microbeads ¹¹⁸	anti-inflammatory delivery with indirect redox modulation	LPS-stimulated HCT-116 cells	Cele/CMCs/Alg tested at 100–250 μ M; NO measured by Griess assay; ROS assessed by H ₂ -DCFDA	Mucoadhesivity: 59.32 $\pm$ 0.74%. Non-toxic range: 100–250 μ M. Cele-250 μ M/CMCs/Alg reduced NO generation by 61.14% $\pm$ 3.67%. Drug release sustained up to 24 h.	Not a direct ROS scavenger; useful as inflammatory/redox-modulating delivery benchmark.

Values are reported as assay-specific benchmarks and should not be interpreted as directly comparable potency rankings because the cited studies used different ROS/RNS sources, concentrations, exposure times, detection methods, and biological models.

Building on the systems discussed above, ROS-scavenging and ROS-responsive strategies can be regarded as two complementary but mechanistically different approaches to redox modulation. ROS-scavenging systems are designed to directly neutralize excessive oxidants or catalytically convert them into less reactive species, thereby restoring redox homeostasis. This category includes small-molecule antioxidants, nitroxides, polyphenols, manganese porphyrins, cerium oxide, and Prussian blue nanozymes as well as polymeric systems bearing antioxidant or catalytic motifs. Their main advantage is the direct attenuation of oxidative stress, which is particularly relevant in chronic inflammation, neurodegeneration, osteoarthritis, cardiovascular injury, and wound healing. However, excessive or non-localized scavenging may interfere with physiological ROS signaling, and catalytic systems require careful evaluation of persistence, off-target activity, and long-term safety. ROS-responsive systems, in contrast, do not primarily act by continuously removing oxidants but exploit pathological ROS as biochemical triggers for bond cleavage, network degradation, swelling, solubility changes, payload release, or prodrug activation. Typical chemistries include thioketals, boronic esters, thioethers, selenides, diselenides, and other oxidizable motifs. These systems are especially useful when ROS-rich microenvironments are used to achieve spatially and temporally controlled therapy; for example, in inflamed tissues, tumors, or acute oxidative bursts. Their main limitation is that responsiveness depends strongly on the local ROS concentration, exposure time, and oxidant identity, and some systems are activated only under exogenous H₂O₂ concentrations higher than those present in physiological redox signaling. Importantly, the two strategies are not mutually exclusive. Several advanced platforms combine ROS-cleavable linkers with catalytic or sacrificial scavenging motifs, enabling simultaneous oxidative stress attenuation and on-demand therapeutic release. Representative systems and quantitative assay-specific benchmarks are summarized in Table 1.

Collectively, these examples illustrate the chemical diversity and mechanistic versatility of molecular ROS scavengers while also highlighting their intrinsic limitations, including limited stability, rapid clearance, and suboptimal localization. These constraints have driven the development of polymeric and nanostructured ROS-scavenging systems, which offer improved retention, controlled presentation of scavenging motifs, enhanced catalytic turnover, and selective activity within pathological microenvironments.

POTENTIAL POLYMERIC DELIVERY SYSTEMS FOR ROS SCAVENGERS

In recent years, a wide variety of biomaterials have been explored for their ability to scavenge ROS and alleviate oxidative stress-related damage.^{119–121} Among these, polymeric systems such as nanoparticles, nanogels, and hydrogels have attracted particular attention (Figure 2).

Their three-dimensional networks and tunable compositions indeed allow incorporation of ROS-scavenging moieties or catalytic nanoparticles while providing controlled therapeutic release.

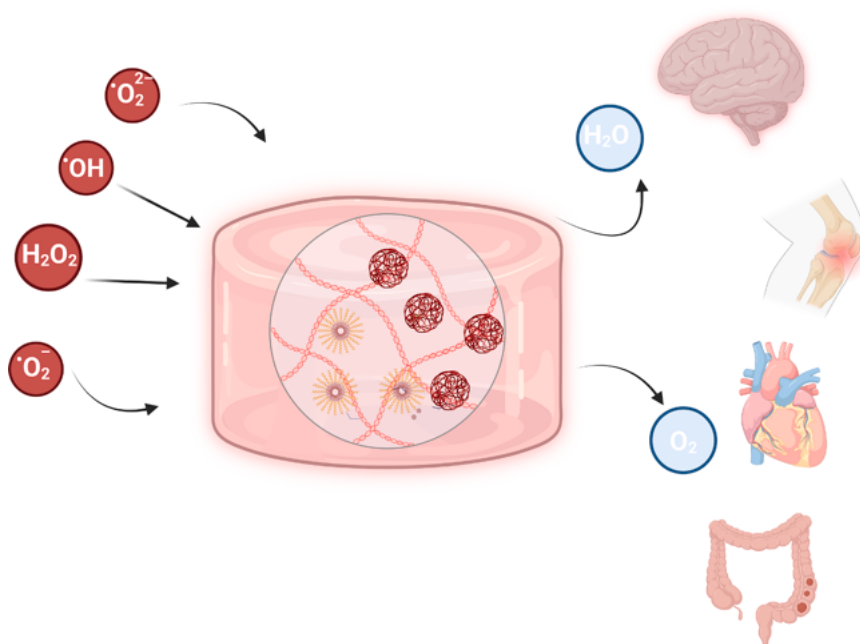


Figure 2. ROS-scavenging composite hydrogel for redox homeostasis restoration

Schematic representation of a polymeric hydrogel/nanogel composite designed to regulate oxidative stress.

Reactive oxygen species, including hydroxyl radicals ($\text{OH}^\bullet$), superoxide ($\text{O}_2^{\bullet -}$), and hydrogen peroxide (H_2O_2), are intercepted by the redox-active network containing embedded nanogels and antioxidant/catalytic components. The platform promotes ROS scavenging or conversion into less reactive species, such as H_2O and O_2 , supporting localized redox modulation in pathological microenvironments associated with neurodegeneration, joint inflammation, cardiovascular injury, and intestinal inflammatory diseases.

Metal oxide and polymer-based nanoparticles can mimic antioxidant enzyme activity, and nanogel- or hydrogel-based platforms offer hydrated networks that enable localized and sustained ROS regulation.^{122,123} The same polymeric architectures can be readily engineered to respond to additional pathological cues, including pH variations, temperature changes, or enzymatic activity, enabling the development of multi-responsive platforms that integrate redox regulation with broader microenvironmental sensing.^{124,125} Before discussing polymeric delivery systems, it is useful to frame the diversity of ROS involved in different pathological contexts and to relate each ROS type to its dominant biological effects, experimental models, and available scavenging strategies. As summarized in Table 2, distinct ROS species differ in reactivity, lifetime, and cellular localization, imposing specific constraints on the chemistry and architecture of polymeric systems designed to modulate oxidative stress.

Nanogels as redox interfaces

Within the broader landscape of redox-modulating strategies, nanogels occupy a specific position that should be distinguished from other antioxidant delivery or ROS-regulating approaches. Small-molecule antioxidants and redox modulators remain attractive because of their chemical simplicity, defined structures, and established pharmacological profiles, but their therapeutic performance is often limited by rapid clearance, poor localization, and insufficient control over the spatial and temporal dynamics of ROS-rich microenvironments.¹²⁶ Liposomes and polymeric micelles can improve the solubility and delivery of hydrophobic antioxidants, although premature leakage, dilution-induced instability, and limited control over ROS-triggered degradation may reduce their efficacy unless specific responsive motifs are introduced.^{127,128}

Exosomes and extracellular vesicles offer biological communication properties and potential tissue tropism, but their hetero-

geneity, loading reproducibility, storage stability, and large-scale production remain challenging.¹²⁹ Inorganic nanoparticles and nanozymes, including cerium oxide, Prussian blue, and manganese-based systems, provide robust catalytic ROS-scavenging activity, but their long-term persistence, biodegradation, metal-related toxicity, and fate after repeated administration require careful assessment.¹³⁰ Finally, gene-based strategies targeting antioxidant pathways such as Nrf2, GPX4, or NOX can modulate redox homeostasis upstream but involve more complex delivery, safety, and regulatory issues and may be less suited to transient, localized ROS buffering.¹³¹ In this context, nanogels offer genuine advantages when redox modulation requires a hydrated, deformable, and chemically tunable nanoscale network rather than only a molecular antioxidant or a passive carrier. Their cross-linked architecture allows the simultaneous incorporation of catalytic centers, sacrificial ROS-cleavable motifs, antioxidant molecules, and therapeutic payloads, while mesh size, charge, degradability, and surface chemistry can be adjusted to control diffusion, retention, and release.¹³² Compared with liposomes or micelles, nanogels can provide stronger structural integrity and a more direct coupling between oxidative cues and network swelling, degradation, or payload release.^{133,134} Compared with inorganic nanozymes, polymeric nanogels can improve colloidal stability, reduce uncontrolled aggregation, and introduce degradable or excretable components.¹³⁵ Compared with free antioxidants, they can enhance local residence time and protect labile scavengers from premature degradation. However, nanogels are not universally superior. Their advantages become most relevant when localized, sustained, and disease-responsive redox modulation is required, particularly in tissues where free molecules would be rapidly cleared and where macroscopic matrices such as hydrogels or scaffolds can further improve retention.

For these reasons, nanogels have emerged as some of the most versatile polymeric platforms for the modulation of oxidative stress.^{136–138} Nanogels are nanoscale, three-dimensional polymer networks that combine the high-water content, softness, and molecular permeability of hydrogels with the colloidal

Table 2. ROS in physiology and disease: Detection, modulation strategies, and polymeric delivery platforms

ROS species	Physiological role/ pathological context	Representative diseases or conditions	Common <i>in vitro</i> ROS models/stimuli	Representative scavengers or redox motifs	Polymeric platforms highlighted in this review
Superoxide (O_2^-)	signaling in immune response and mitochondrial function, excessive levels drive oxidative stress and inflammation	Parkinson's disease, Alzheimer's disease, cardiovascular diseases, ischemia-reperfusion injury	mitochondrial stress, rotenone or MPTP treatment, LPS stimulation	Mn-porphyrins, nitroxides (TEMPO, Tempol), CeO_2 nanozymes	nitroxide-bearing nanogels, ceria-loaded nanogels, nanogel-hydrogel composites for sustained scavenging
Hydrogen peroxide (H_2O_2)	second messenger in redox signaling, high concentrations induce oxidative damage and trigger inflammatory cascades	osteoarthritis, rheumatoid arthritis, depression, acute lung injury, chronic wounds	direct H_2O_2 exposure, NOX activation, inflammatory cytokine stimulation	thioetheral linkers, diselenide bonds, boronic esters, selenium-containing motifs	ROS-cleavable nanogels (thioetheral, diselenide), H_2O_2 - responsive hydrogels, nanogel-hydrogel hybrids
Hydroxyl radical (OH)	no physiological signaling role; highly cytotoxic, induces lipid peroxidation and DNA damage	ischemic stroke, neurodegeneration, acute inflammatory injury	Fenton reaction (Fe^{2+}/H_2O_2), oxidative burst models	nitroxides, polyphenols, catalytic nanozymes	nanogels with sacrificial scavenging motifs, catalytic nanozyme-based polymeric systems
Lipid peroxides/lipid radicals (ROO, LOOH)	minimal physiological role, central mediators of ferroptosis and membrane damage	neurodegenerative diseases, ischemic injury, cardiovascular disorders	erastin-induced ferroptosis, iron overload, oxidative stress models	selenium-based motifs, GPX4-supporting systems; polyphenols	diselenide-crosslinked nanogels, nanozyme-loaded nanogels targeting ferroptotic pathways
Reactive nitrogen species (e.g., $ONOO^-$)	modulate inflammatory signaling; excessive levels cause nitrosative stress	inflammatory bowel disease, rheumatoid arthritis, neuroinflammation	iNOS induction, inflammatory cytokines, NO donors	dual ROS/RNS scavengers, selenium- and thiol- containing systems	dual-function nanogels and hybrid polymeric platforms
Mixed ROS microenvironments	controlled redox signaling vs. pathological oxidative imbalance	tumors, chronic inflammation, diabetic wounds, osteoarthritis	LPS, hypoxia- reoxygenation, chronic inflammatory stimulation	combined antioxidant and catalytic motifs	combined nanogel-hydrogel systems, nanogel-nanofiber scaffolds for prolonged retention and redox control

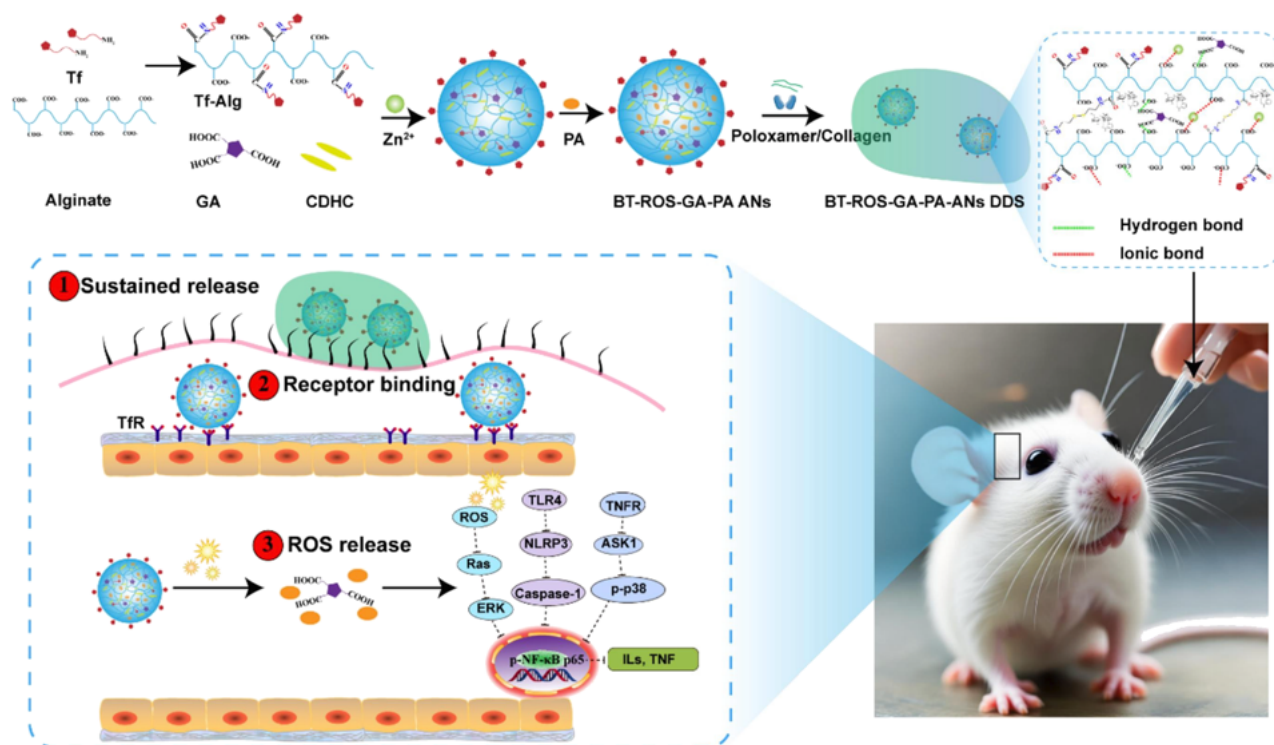


Figure 3. Brain-targeted ROS-responsive polysaccharide nanogels for antidepressant therapy

Schematic representation of a transferrin-modified polysaccharide nanogel designed for nose-to-brain delivery and ROS-responsive drug release in the hippocampal microenvironment.

The nanogel promotes antioxidant and anti-inflammatory effects by reducing ROS levels and inflammatory mediators, thereby contributing to the improvement of depression-related pathological features.

Reprinted with permission from Elsevier.¹⁵⁰

dimensions and transport properties of nanoparticles.¹³⁹ Their highly hydrated crosslinked networks can protect labile antioxidants from premature degradation, while tunable nanoscale dimensions and chemistry enable access to disease-specific microenvironments characterized by elevated superoxide, hydrogen peroxide, and lipid peroxides.^{140–142} From a synthesis perspective, nanogels are commonly prepared from natural polymers such as alginate, chitosan, hyaluronic acid, and gelatin or from synthetic building blocks such as polyethylene glycol (PEG), polyvinyl alcohol (PVA), polyethylenimine (PEI)/polyamidoamine (PAMAM) dendrimers, and poly(meth)acrylates, using preparation methods that include covalent or dynamic crosslinking, ionic gelation, and emulsion/inverse emulsion polymerization.¹⁴³ Across the available studies, nanogels are not simply passive depots. They function as active redox interfaces that bind, transform, or spatially confine ROS while simultaneously coordinating therapeutic release in response to oxidative or inflammatory cues that characterize each disease setting.¹⁴⁴ Several contributions first establish “platform” nanogels in which the network is intrinsically ROS responsive and already endowed with antioxidant capacity.¹⁴⁵ A closely related strategy is represented by dendrimer-based nanogels in which the crosslinker itself confers ROS responsiveness and intrinsic radical-scavenging activity.¹⁴⁶ In the thioketal-cross-linked platform, generated via flash nanoprecipitation of PAMAM dendrimers

with an N-hydroxysuccinimide (NHS)-activated thioketal linker, the resulting compact nanogels undergo fragmentation upon exposure to peroxide-rich environments and simultaneously quench 2,2-diphenyl-1-picrylhydrazyl (DPPH $\cdot$), hydroxyl radicals, and hydrogen peroxide in dose-dependent scavenging assays.

These nanogels display negligible cytotoxicity in macrophage and lens epithelial cells, confirming that the redox-responsive crosslinking motif enables both ROS-triggered destabilization and antioxidant action without compromising biocompatibility. In a different architecture, nitroxide-bearing reversible addition-fragmentation chain transfer (RAFT)-derived nanogels and related diselenide-crosslinked systems translate the same idea into soft, water-swollen networks.¹⁴⁷ In these platforms, nitroxide or Se–Se motifs act as sacrificial sinks for superoxide and peroxides, while the nanogel matrix behaves as a depot for drugs or genes. Beyond intrinsic ROS-scavenging activity, these nanogels stabilize low-density lipoproteins (LDLs) against oxidation for up to 1 month, a functional outcome that places them among promising candidates for early-stage cardiovascular intervention, particularly in atherosclerosis, where oxidized LDL is a primary pathogenic trigger. These platform studies mainly focus on physicochemical characterization and cell-level oxidative challenges, but they define a design space in which nanogels act as combined sensors and scavengers, loosening or

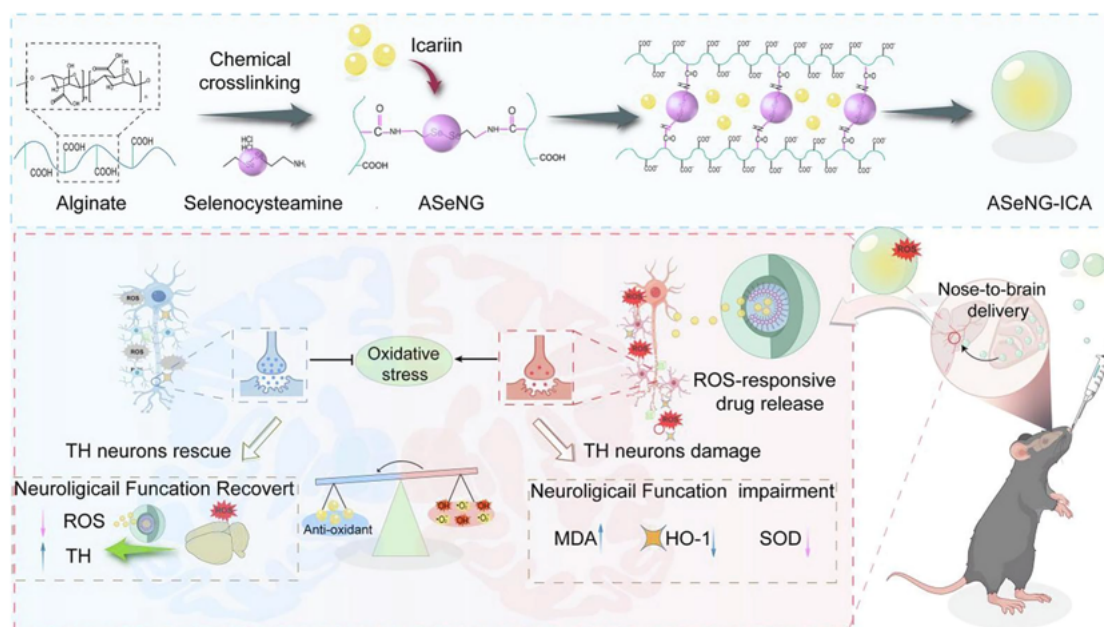


Figure 4. ROS-responsive selenized-alginate nanogels for nose-to-brain delivery in Parkinson's disease

Schematic representation of selenized-alginate nanogels loaded with icariin (ASeNG-ICA) and designed for intranasal administration and brain-targeted delivery. The selenium-containing nanogel network enables ROS-responsive behavior in oxidative microenvironments, supporting drug release, attenuation of oxidative stress and neuroinflammation, and protection of dopaminergic neurons in Parkinson's disease-related pathological conditions.

Reprinted with permission from Elsevier.¹⁵²

disassembling their network as ROS levels rise. Other studies extend this toolbox by matching nanogel chemistry to organ-specific oxidative microenvironments.^{148,149} In the central nervous system, a transferrin- and disulfide-modified alginate nanogel (BT-ROS-GA-PA ANs; biotin-conjugated [BT], reactive oxygen species [ROS]-responsive nanogel designed to deliver two active compounds: glycyrrhizic acid [GA] and paeoniflorin [PA]) was engineered to deliver glycyrrhizic acid and paeoniflorin to the hippocampus in a chronic unpredictable mild stress (CUMS) model of depression.¹⁵⁰ Disulfide bridges impart H_2O_2 -sensitive swelling and drug release, while transferrin grafting promotes receptor-mediated transport across the blood-brain barrier and accumulation in ROS-enriched hippocampal regions. Intranasal administration is particularly advantageous for neurological diseases, as it enables direct nose-to-brain delivery while bypassing the blood-brain barrier and reducing systemic exposure.¹⁵¹

In vivo, intranasal administration of the thermosensitive BT-ROS-GA-PA nanogel increases hippocampal drug levels compared to oral delivery (Figure 3), lowers ROS content, and decreases IL-6, TNF- α , and prostaglandin E_2 (PGE $_2$) together with behavioral endpoints of depression, indicating that ROS-responsive release *in situ* can rebalance both oxidative and inflammatory components of the CUMS phenotype.

A complementary nose-to-brain strategy is exemplified by the selenized alginate nanogel developed for brain-targeted delivery of icariin in Parkinson's disease.¹⁵² Here, diselenide crosslinks function as ROS-responsive gates. In peroxide-rich milieu, Se-Se bonds undergo selective cleavage, accelerating drug

release while generating transient selenium intermediates that contribute to neutralizing oxidants (Figure 4). This redox-coupled release is paired with improved brain penetration, with bio-distribution indicating higher accumulation of icariin in the striatum and substantia nigra compared to the free drug.¹⁵³ At the cellular level, the formulation restores mitochondrial homeostasis, recovering membrane potential, complex I activity, and ATP production while reducing DCF-detected ROS and malondialdehyde and increasing superoxide dismutase (SOD) and glutathione peroxidase. In the mitochondrial permeability transition pore (MPTP) mouse model, intranasal administration attenuates dopaminergic neuron loss, preserves helper T cells (TH)-positive fiber density, and improves motor performance across rotarod, pole, and open field tests.

Beyond neurodegeneration, diselenide-rich nanogels have also been adapted to tumor immunotherapy.¹⁵⁴ The platform leverages the dynamic cleavage and re-formation of Se-Se and Se-O bonds to couple ROS-responsive behavior with immunomodulation.¹⁵⁵ Under oxidative stress, abundant in MC38 colorectal tumors, selenium-based bonds undergo reversible scission, promoting siPD-L1 release while attenuating local ROS (Figure 5). This dual action enhances PD-L1 silencing, suppresses autophagy-mediated immune evasion, and boosts CD8 $^+$ T cell infiltration. *In vivo*, systemic administration results in significant tumor growth inhibition with reduced oxidative burden and minimal off-target toxicity.

Cerium-based nanosystems appear in mechanistically distinct configurations addressing neurodegeneration and pulmonary inflammation, respectively.^{156,157} A catalytic variation of this

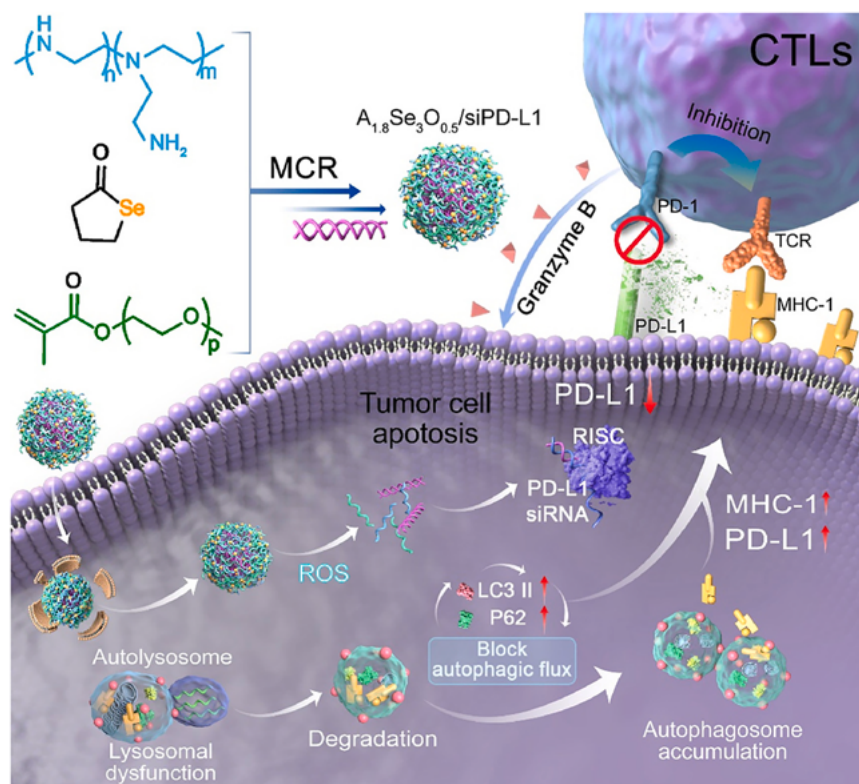


Figure 5. Schematic illustration of the preparation and antitumor mechanisms of the nanogels

Nanogels with three features (ROS sensitivity, proton sponge ability, and biocompatibility) were synthesized using selenol chemistry-mediated multicomponent reaction (MCR) technology with LMW PEI, γ -SBL, and PEGMA. After electrostatic complexation with siRNA, nanogels were formed. These nanogels demonstrated improved uptake by tumor cells and underwent endosomal escape and siRNA release in an intracellular oxidative environment, resulting in the downregulation of PD-L1 and increased expression of major histocompatibility complex class I (MHC-I). Ultimately, CD8+T cells are activated and induce tumor cell death by secreting granzyme B. ROS-sensitive $A_{1.8}Se_3O_{0.5}/siPD-L1$ nanogels: synthesis and antitumor mechanism.

Reprinted with permission from Elsevier.¹⁵⁵

ethyl acetate fraction of *Olea ferruginea* (EAF-AC-NGs) are designed to stabilize polyphenolic antioxidants and to target ROS-driven cardiac hypertrophy.¹⁶¹ Encapsulation increases particle size from about 50 to 110 nm and yields high entrapment efficiency, while *in vivo*, the formulation restores superoxide dismutase

and catalase activities suppressed in isoproterenol-induced hypertrophy, reduces myocardial H_2O_2 and malondialdehyde, and lowers BAX expression more effectively than the free plant fraction, with histology confirming improved preservation of myocardial architecture (Figure 7). From a formulation perspective, such size ranges and dispersity could be further refined using microfluidic or flow-assisted fabrication strategies, enabling tighter control over nanogel dimensions and batch-to-batch reproducibility.¹⁶²

In a murine middle cerebral artery occlusion (MCAO) model, these nanogels accumulate in the ischemic penumbra and thrombus regions enriched in P-selectin.¹⁶³

Upon H_2O_2 exposure, phenylboronic ester and Se-Se motifs undergo sequential cleavage, triggering urokinase release alongside peroxide scavenging.¹⁶⁴ In *in vivo* models, treatment reduces infarct volume on 2,3,5-triphenyltetrazolium chloride (TTC) staining, lowers tissue iron and lipid peroxidation, restores GPX4 levels, and improves neurological deficit scores, indicating combined thrombolytic and anti-ferroptotic activity. Barrier tissues provide further examples of how ROS-responsive nanogels can be adapted to local delivery and retention constraints. In the cornea, a supramolecular hyaluronic acid nanogel assembled through cyclodextrin-adamantane host-guest interactions and thioketal-containing PEG arms yields a dexamethasone (DEX)-loaded construct whose network is cleaved by pathologically high H_2O_2 .¹⁶⁵ The formulation remains stable in storage and releases minimal drug in the absence of oxidants but reaches near-complete release in millimolar H_2O_2 while maintaining low cytotoxicity in human corneal epithelial cells (Figure 8). In a rabbit

rationale is represented by hyaluronic acid nanogels co-loading citric acid-coated cerium nanozyme with the neuropeptide chimeric antigen receptor T-cell (CART) for Alzheimer's disease.¹⁵⁸ Here, CeO_2 domains exploit Ce^{3+}/Ce^{4+} cycling to decompose superoxide and hydrogen peroxide, while hyaluronic acid contributes targeting and CART provides neuroprotective signaling (Figure 6). The formulation suppresses ABTS⁺ and DPPH radical signals and decreases intracellular ROS and lipid peroxidation. It also mitigates ferroptosis-associated readouts such as HO-1 upregulation and iron-induced neuronal loss. In APP/PS1 transgenic mice, treatment reduces amyloid-associated oxidative stress, improves synaptic integrity, and rescues cognitive performance, illustrating how nanozyme-based ROS modulation can be integrated with peptide therapeutics in a nanogel scaffold.¹⁵⁹

A second, mechanistically distinct cerium-based platform addresses the rapidly escalating oxidative burden characteristic of acute lung injury.¹⁶⁰ Curcumin-loaded ceria nanoenzymes combine Ce^{3+}/Ce^{4+} -mediated detoxification of superoxide and peroxides with the anti-inflammatory action of curcumin, improving curcumin dispersion and retention in inflamed pulmonary tissue. In lipopolysaccharide-induced acute lung injury, the formulated nanoenzymes reduce DHE-detected superoxide and malondialdehyde, restore SOD and catalase activities, and suppress TNF- α , IL-1 β , and IL-6. Histologically, they alleviate alveolar wall thickening, edema, and inflammatory infiltration more effectively than free curcumin or CeO_2 alone. Cardiovascular applications highlight the importance of tuning both release and radical selectivity. Alginate-chitosan nanogels loaded with an

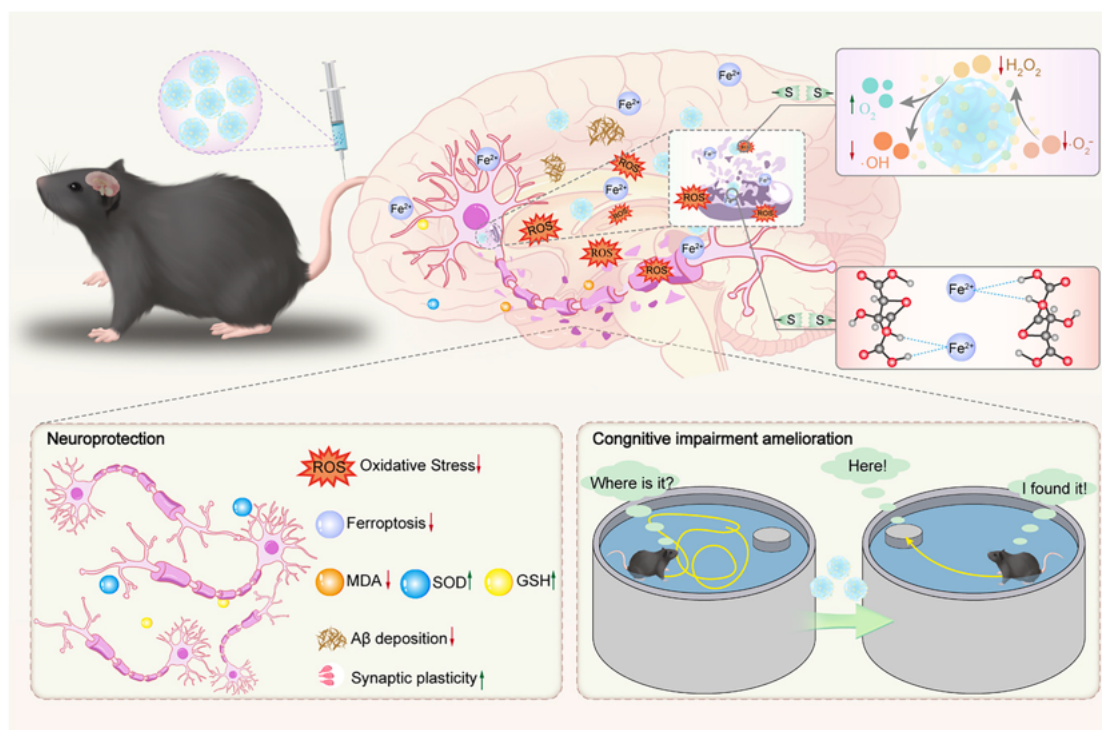


Figure 6. Nanogel-based strategies for the treatment of Alzheimer's disease

Schematic illustration of nanogel platforms designed to modulate Alzheimer's disease-related pathological processes, including oxidative stress, neuro-inflammation, and impaired neuronal homeostasis.

The figure highlights how nanogels can be engineered to improve brain delivery, protect therapeutic agents, and promote localized modulation of disease-associated microenvironments.

Reprinted with permission from Elsevier.¹⁵⁸

chemical burn model, once-daily instillation inhibits corneal neovascularization and reduces local pro-inflammatory cytokines, with efficacy comparable to twice-daily conventional DEX drops.

In the lungs, zinc-alginate curcumin (ZA-Cur) nanogels exploit ionic crosslinking and hydrogen bonding to integrate curcumin within a negatively charged network that can be nebulized as a fine aerosol.¹⁶⁶ The system displays faster release at 37°C, at a mildly acidic pH, and in the presence of monovalent cations, conditions intended to mimic inflamed airways (Figure 9). *In vitro*, ZA-Cur quenches DPPH radicals and reduces intracellular ROS in LPS- or H₂O₂-challenged alveolar cells more efficiently than free curcumin at the same dose.

Finally, non-classic models underline how the same logic can be exported beyond canonical human pathologies. ACC@TBO nanogels, based on azidobenzoyl-modified carboxymethyl chitosan crosslinked under light and loaded with toluidine blue O (TBO), have been used in plant protection as dual photodynamic/Fenton-like antibacterial agents.¹⁶⁷ The mesh architecture protects the photosensitizer, while Fe²⁺/H₂O₂ in the infected microenvironment promotes network changes and ROS generation, which eradicate phytopathogenic bacteria without damaging the host plant. Although the biological context differs, the design principle remains aligned with the biomedical nanogels above: an oxidatively stressed niche is exploited as both a trigger for release and a target for controlled ROS production or quenching. Taken

together, these studies show that nanogels can be programmed to read and reshape highly specific oxidative signatures. Their activity spans hippocampal ROS surges in depression, mitochondrial and neuroinflammatory stress in Parkinson's and Alzheimer's disease, peroxide-driven injury in cardiac hypertrophy and stroke, corneal neovascularization and acute lung injury, and even oxidatively stressed niches in plant infections. By combining ROS-cleavable crosslinkers, catalytic redox centers, and targeting ligands with formulation choices that respect the constraints of each organ and route of administration, nanogels move from generic "antioxidant carriers" to redox devices that localize, modulate, and sometimes exploit ROS in a disease-selective manner. Beyond therapeutic delivery and ROS scavenging, an increasingly important direction is the development of theranostic redox-responsive nanogels, in which diagnostic and therapeutic functions are integrated within the same platform.¹⁶⁸ In this context, imaging is not only used to track nanogel biodistribution but also to report on local oxidative activation, payload release, and treatment response.¹⁶⁹

ROS-responsive nanogels can be engineered with activatable fluorescent or chemiluminescent probes, ratiometric redox sensors, magnetic resonance contrast agents, photoacoustic components, or catalytic nanozymes with imaging capability.^{170,171} In this context, redox-sensitive nanoscale coordination polymers provide a useful example of how diagnostic and therapeutic

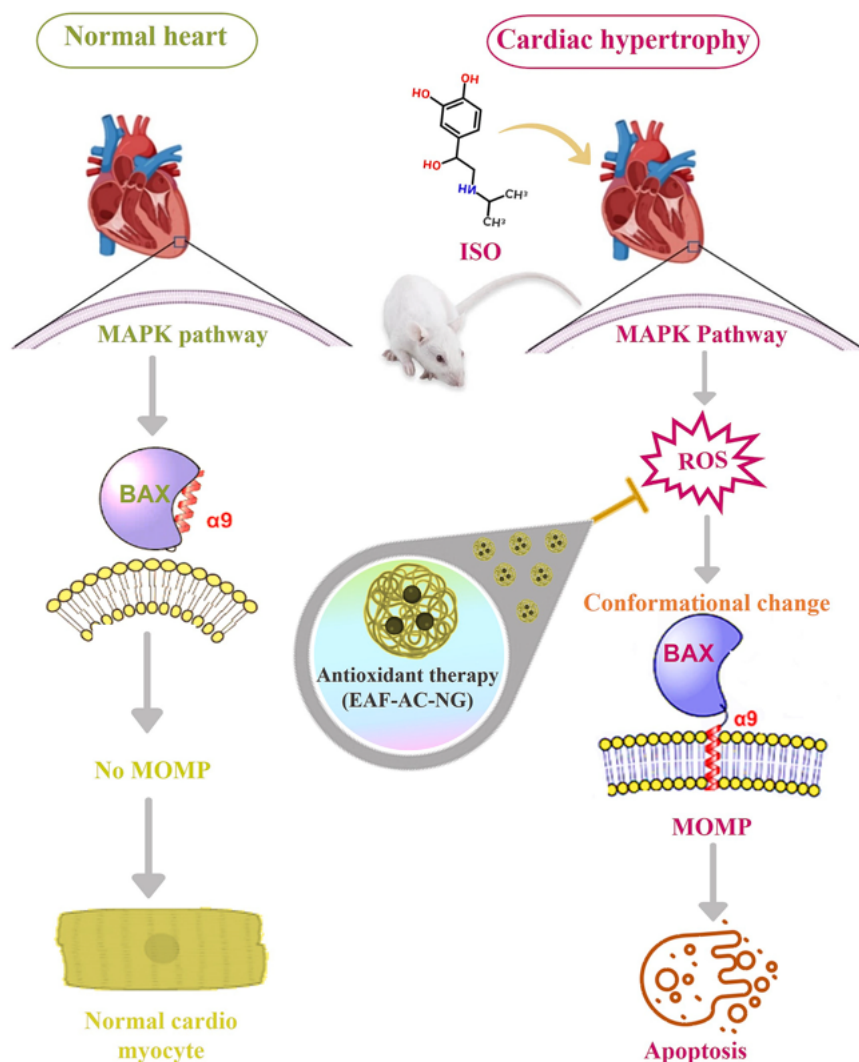


Figure 7. Nanogel-mediated modulation of isoproterenol-induced oxidative stress

Schematic illustration of the oxidative stress pathway activated by isoproterenol and the proposed protective mechanism of EAF-AC-NG.

The nanogel system is presented as a redox-modulating platform able to counteract ROS generation, oxidative damage, and downstream inflammatory/cellular stress responses, thereby supporting tissue protection in the proposed pathological model.

Reprinted with permission from Elsevier.¹⁶¹

sufficiently present, or whether the therapeutic response correlates with local redox changes. By incorporating imaging functions, redox-responsive nanogels and related polymeric platforms could be used to monitor accumulation, activation, release, and treatment outcome.

However, this integration also introduces additional challenges, including the need to quantitatively correlate imaging signals with biologically meaningful ROS/RNS or glutathione (GSH) levels, avoid background activation, preserve biodegradability and clearance, and prevent imaging components from increasing long-term toxicity or formulation complexity.

Combined systems: Hydrogels as macroscopic redox matrices

However, standalone nanogels face practical limitations.^{174,175} Owing to their small size, which allows rapid vascular/lymphatic clearance, free nanogels can exhibit short residence times following

functions can be combined within a single platform. Zhao et al. developed Mn-SS/DOX@PDA-PEG nanoparticles based on manganese ions and dithiodiglycolic acid as a disulfide-containing bridging ligand.¹⁷² The disulfide bonds within the coordination network were cleaved in the presence of intracellular glutathione, leading to nanoparticle disassembly and accelerated doxorubicin release under reducing conditions. At the same time, the Mn²⁺ centers acted as T1 magnetic resonance imaging contrast agents, allowing the *in vivo* visualization of nanoparticle accumulation in the tumor after systemic administration. This example highlights a key principle of redox theranostics: the same structural element responsible for redox responsiveness can be integrated with imaging-active components, enabling both stimulus-responsive therapy and image-guided monitoring of delivery. Such approaches are important for precision redox modulation because oxidative and reductive stress are spatially heterogeneous, disease dependent, and temporally dynamic.¹⁷³ A purely therapeutic nanogel or nanocarrier can scavenge ROS or release a drug, but it cannot indicate whether the platform has reached the pathological site, whether the redox trigger is

local injection (e.g., intra-articular injection into the synovial joint), diminishing their efficacy at the target site.¹⁷⁶ Moreover, without a supporting matrix, nanogels may lack mechanical integrity under physiological stress and can exhibit burst release.¹⁷⁷ These constraints have motivated hybrid constructs in which ROS-responsive nanogels are embedded within larger biomaterial platforms (hydrogels, nanofibrous scaffolds, or composite dressings), thereby improving retention, robustness, and release control. Hydrogels in particular provide a robust framework. These three-dimensional, crosslinked polymer networks mimic soft tissues due to their high water content, biocompatibility, and tunable mechanical properties, and ROS-scavenging hydrogels have shown promise across multiple oxidative stress-related pathologies.¹⁷⁸ For example, a hyaluronic acid hydrogel chemically conjugated with epigallocatechin gallate scavenged ROS in osteoarthritic joints, supported mesenchymal stem cell viability, and modulated local inflammation by promoting M2 macrophage polarization.¹⁷⁹ Encapsulating nanogels within bulk hydrogel matrices can mitigate premature burst release, enhance mechanical robustness, and prolong intra-

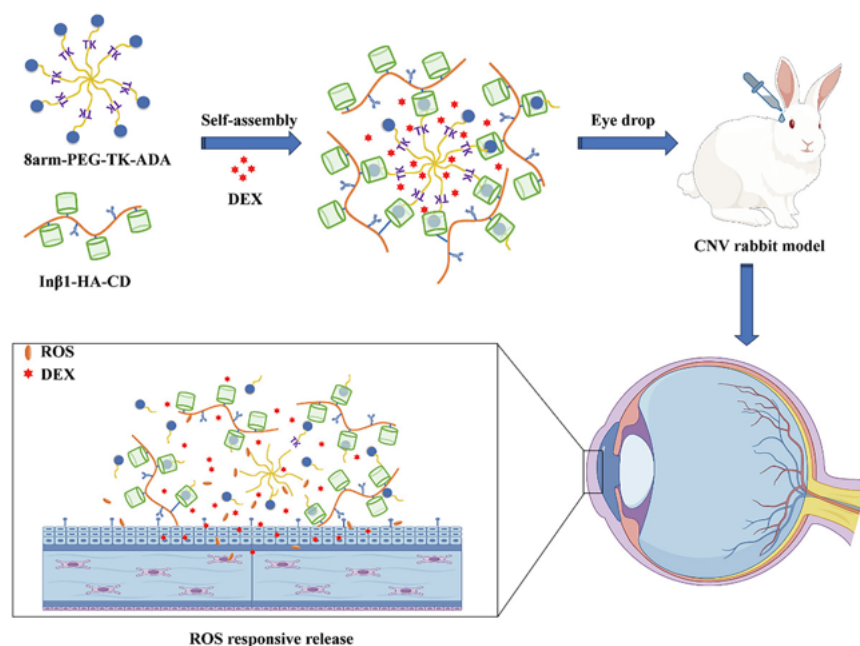


Figure 8. Host-guest-assembled nanogel for ROS-stimulated payload release

Schematic representation of nanogel formation through host-guest interactions and subsequent ROS-triggered release of the encapsulated therapeutic payload.

The figure illustrates how ROS-responsive motifs within the supramolecular nanogel network enable structural changes or degradation under oxidative conditions, promoting controlled drug release in ROS-enriched pathological microenvironments. Reprinted with permission from Elsevier.¹⁶⁵

tissue retention.¹⁸⁰ In joint environments, thermosensitive or adhesive hydrogel systems can further improve outcomes by resisting washout and enabling sustained activity over days.¹⁸¹

This concept is exemplified by He et al., who developed a bioadhesive and lubricating hydrogel for rheumatoid arthritis, combining dopamine and sulfonated hyaluronic acid networks

in situ. The gradual release of kartogenin supports chondrogenic differentiation and cartilage repair. *In vivo*, this multifunctional matrix modulated inflammation, reduced oxidative damage, and reversed cartilage erosion in both early- and late-stage rheumatoid arthritis, enabling extended joint residence and mechanical durability.

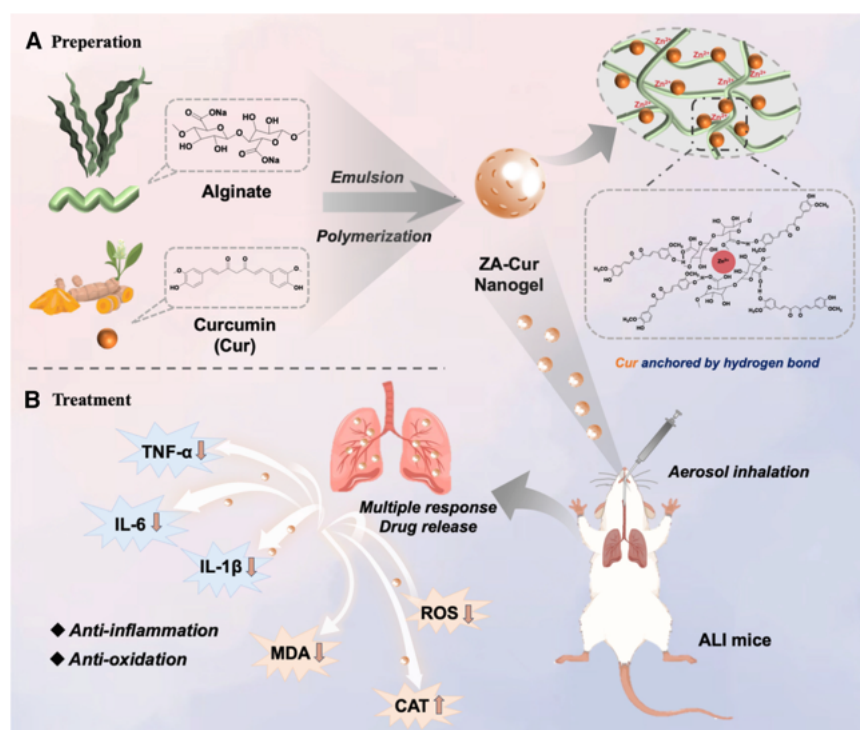


Figure 9. Nanogel synthesis and nanogel-based treatment of acute lung injury

Schematic illustration of the preparation of therapeutic nanogels and their application in a mouse model of acute lung injury.

The figure highlights how the nanogel platform can modulate ROS-associated inflammatory responses, reduce oxidative damage, and support tissue protection in the injured lung microenvironment.

Reprinted with permission from Elsevier.¹⁶⁶

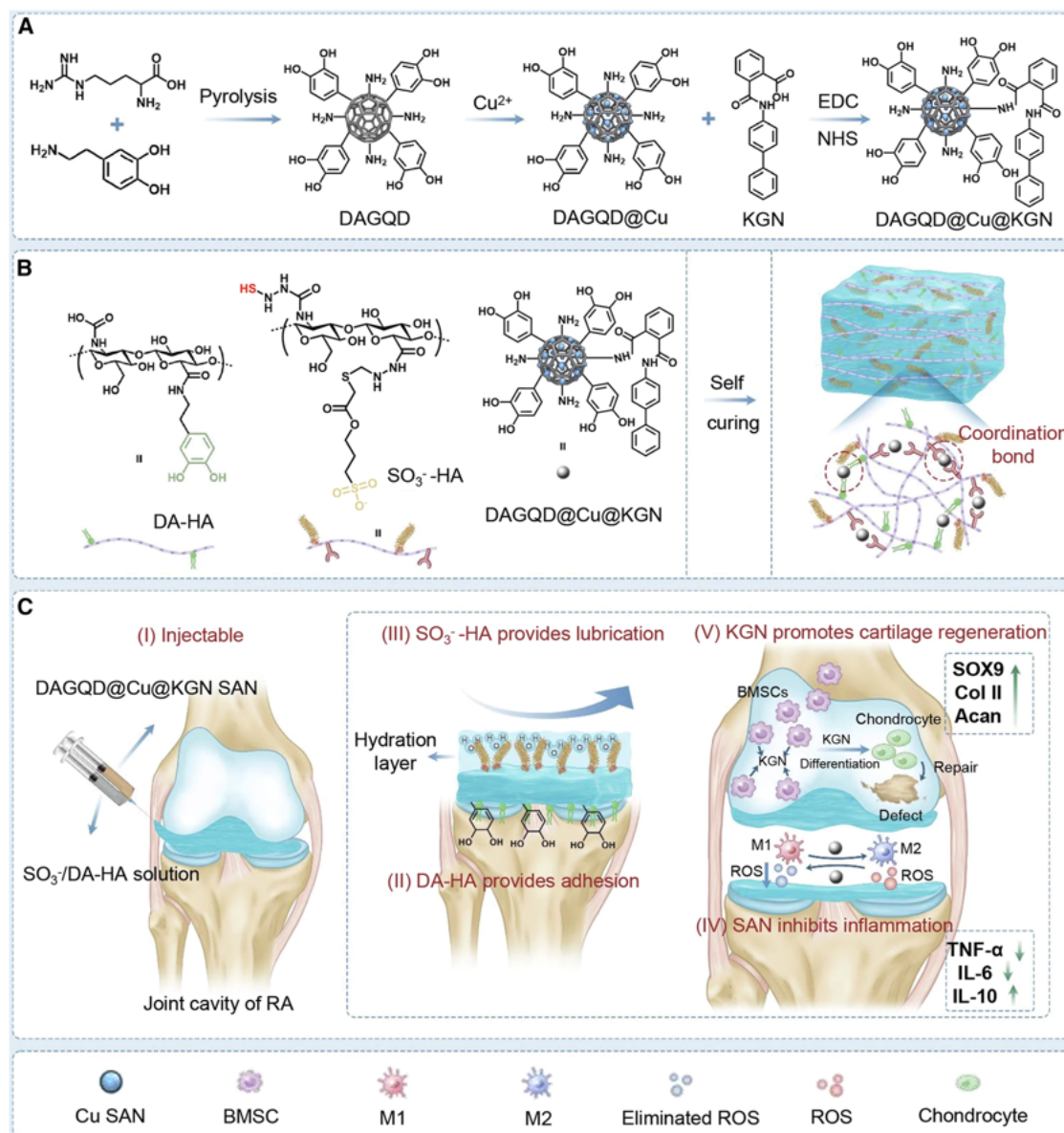


Figure 10. Injectable ROS-scavenging hydrogel for rheumatoid arthritis therapy

(A) Synthesis of DAGQD@Cu SAN and DAGQD@Cu@KGN SAN.

(B) Synthesis of DAGQD@Cu@KGN-SO₃⁻/DA-HA hydrogel.

(C) DAGQD@Cu@KGN-SO₃⁻/DA-HA hydrogel for RA therapy. (I) Hydrogel self-cures *in situ* within the joint cavity; (II, III) DAGQD@Cu@KGN-SO₃⁻/DA-HA hydrogel displays lubrication and adhesion properties; (IV) regulation of inflammatory microenvironment facilitated by SAN with remarkable ROS-scavenging capabilities; (V) kartogenin released from the hydrogel promotes the recruitment of bone marrow mesenchymal stromal cells (BMSCs) and induces their differentiation into chondrocytes, facilitating the repair of cartilage damaged by rheumatoid arthritis.

Reprinted with permission from Nature Publishing Group.¹⁸²

The combination of nanogels with scaffolds can also leverage multi-modal triggers for controlled release, pairing hydrogel responsiveness to physiological cues (pH, enzymes, and temperature) with nanogel ROS responsiveness.¹⁸³ Zhou et al. exemplified this strategy with a dual-responsive nanohydrogel system (KPP@PELE) for osteoarthritis therapy, integrating a PAMAM dendrimer functionalized with MMP-13-cleavable peptide linkers and loaded with kartogenin,

encapsulated within a thermosensitive poly(D,L-lactide)-poly(ethylene glycol)-poly(D,L-lactide) (PELE) hydrogel.¹⁸⁴ The *in situ* gelling PELE hydrogel prolongs intra-articular retention, while MMP-13-sensitive linkers enable localized enzyme-triggered release within the osteoarthritis (OA) micro-environment. *In vitro*, the system was biocompatible, suppressed inflammatory responses, and promoted chondrogenic differentiation.

Table 3. Platform for ROS scavenging

Platform	Primary function	ROS interaction mechanism	Main advantages	Key limitations	Representative applications in this review
Polymeric nanoparticles	systemic or local delivery of redox-active agents	encapsulation or surface presentation of scavengers, catalytic nanozyme activity	small size, tunable chemistry, scalable synthesis	limited tissue retention, rapid clearance	catalytic nanozyme delivery in neurodegeneration and acute inflammation
Nanogels	local and responsive redox modulation	ROS-cleavable crosslinkers, sacrificial or catalytic scavenging motifs	high hydration, chemical versatility, protection of labile scavengers	short residence time <i>in vivo</i> , potential burst release	neurodegeneration, cardiovascular disease, ischemic stroke
Hydrogels	local retention and tissue support	immobilized scavengers or redox-active moieties within bulk networks	prolonged retention, mechanical stability, tissue mimicry	limited penetration, slower response kinetics	osteoarthritis, rheumatoid arthritis, wound healing
Nanogel-hydrogel composites	sustained and localized therapy	multi-level ROS responsiveness combining nanogel sensing and hydrogel retention	synergistic redox control, extended therapeutic window	higher formulation complexity	osteoarthritis, rheumatoid arthritis, chronic wounds
Nanogel-nanofiber systems	structural reinforcement and controlled release	ROS-responsive nanogels embedded within fibrous scaffolds	improved mechanical integrity, prolonged retention	fabrication complexity, scale-up challenges	cartilage repair and advanced wound healing

In an OA rat model, it achieved superior cartilage penetration and extended joint residence compared to free kartogenin or hydrogel alone, resulting in enhanced cartilage regeneration and slower OA progression. Beyond joints, numerous studies have demonstrated the effectiveness of ROS-neutralizing hydrogels in promoting wound healing,¹⁸⁵ where sustained oxidative stress impairs regeneration by maintaining inflammation, degrading extracellular matrix components, and inhibiting angiogenesis.¹⁸⁶ Hybrid nanogel-hydrogel dressings can translate antioxidant activity into improved closure rates and tissue quality by combining local retention, infection control, and microenvironmental modulation. In a murine full-thickness wound model, an hyaluronic acid (HA)/PEG hydrogel incorporating chlorhexidine-loaded nanogels (Gel@CLN) promoted markedly faster closure. By day 14, Gel@CLN-treated wounds were essentially healed (~99% closure), whereas wounds treated with unloaded hydrogel or no dressing remained ~85% closed.¹⁸⁷ Histology showed a nearly complete epidermis and organized dermal layers with new capillaries after 2 weeks, in contrast to controls that still lacked continuous epidermis and exhibited persistent inflammatory infiltrates. In diabetic rats, a tannin/small interfering RNA (siRNA)-loaded nanogel-hydrogel composite achieved full closure by day 10 compared to ~18 days in untreated or standard dressing groups, with improved granulation, collagen deposition, and revascularization.¹⁸⁸ Similar trends were reported in additional composite systems, in which reduced inflammatory cytokines and increased angiogenic markers correlated with accelerated granulation and re-epithelialization.¹⁸⁹ Immunohistochemical analyses confirmed that the composite hydrogel significantly reduced proinflammatory cytokines (TNF- α , IL-1 β , IL-6, and MCP-1) and upregulated angiogenic markers (CD31 and α -SMA) in the wound bed, correlating with faster granulation and re-epithelialization.

These outcomes underscore that the multi-pronged therapeutic actions of hybrid nanogel systems (infection control, antioxidant, growth factor modulation, and physical support), translate to significantly improved wound closure rates and tissue quality. Finally, nanofiber scaffolds offer a complementary route to address rapid clearance and burst release by providing structural support while retaining nanogel responsiveness.¹⁹⁰ Embedding ROS-responsive nanogels within electrospun nanofibers can improve mechanical stability and extend retention¹⁹¹ while facilitating controlled release and tissue interaction in dense or highly dynamic environments.¹⁹² Collectively, these hybrid architectures underscore the synergy between structural scaffolds and redox-responsive nanogels. Scaffolds enhance robustness and residence time, while nanogels provide ROS sensing and controlled payload release, improving pharmacokinetics and therapeutic performance in complex oxidative microenvironments. Beyond the individual examples discussed above, several general design principles can be extracted for the rational selection of polymeric redox-modulating platforms. Polymeric nanoparticles are advantageous when systemic administration, small size, and scalable fabrication are required, but their limited tissue retention and rapid clearance may reduce therapeutic efficacy in dynamic or poorly vascularized tissues. Nanogels provide a more versatile redox interface because their hydrated networks can host labile antioxidants, catalytic

nanozymes, or ROS-cleavable linkers while also allowing swelling, degradation, or release in response to oxidative cues. However, free nanogels may still suffer from burst release and insufficient residence time.¹³⁹ Hydrogels address these limitations by providing macroscopic retention, mechanical support, and local confinement, but their larger mesh structure and slower diffusion kinetics may limit penetration into dense tissues. Hybrid nanogel-hydrogel or nanogel-nanofiber systems therefore represent an intermediate strategy, combining nanoscale ROS sensing with macroscopic localization and structural stability.

An additional design criterion concerns the choice between catalytic and sacrificial ROS-scavenging mechanisms. Catalytic scavengers, including cerium oxide, Prussian blue, manganese-based nanozymes, and nitroxide-bearing systems, can undergo repeated redox cycles and are therefore particularly attractive for sustained oxidative microenvironments.¹⁹³ Their main advantage is prolonged activity, but this also requires careful control over dose, biodistribution, redox cycling, long-term persistence, and possible off-target interference with physiological ROS signaling.¹⁹⁴ In contrast, sacrificial chemistries, such as thioethers, boronic esters, diselenide bonds, and oxidizable polymer segments, directly consume ROS or undergo oxidative cleavage, thereby coupling scavenging with drug release or network degradation.¹⁹⁵ These systems are highly useful for transient oxidative bursts and on-demand activation, but their activity is progressively depleted and may be insufficient in chronic diseases unless supported by a depot-like matrix or combined with catalytic elements.¹⁹⁶ Thus, catalytic and sacrificial strategies should not be considered interchangeable but, rather, complementary tools whose selection depends on ROS intensity, exposure duration, and the desired therapeutic output. As highlighted in Table 3, no single polymeric architecture is universally optimal for ROS modulation. Instead, the choice between nanogels, hydrogels, or combined systems should be guided by the dominant ROS species, target tissue, required residence time, and intended therapeutic window. Hybrid platforms emerge as particularly effective when prolonged retention and multi-modal responsiveness are required, bridging the gap between nanoscale redox activity and macroscopic tissue constraints.

CONCLUSIONS

Polymeric nanogels, hydrogels, and hybrid platforms offer a versatile toolbox for the modulation of oxidative stress, combining nanoscale redox responsiveness with tunable drug delivery, tissue retention, and multifunctional therapeutic activity.

Their ability to incorporate catalytic nanozymes, sacrificial ROS-cleavable motifs, antioxidant small molecules, or biologically active macromolecules makes them particularly attractive for diseases in which oxidative stress is spatially localized and dynamically regulated, including inflammation, neurodegeneration, cancer, osteoarthritis, cardiovascular injury, and chronic wounds. However, despite the rapid growth of the field, the translation of these systems remains limited by several conceptual, methodological, and technological challenges. A first major limitation is the lack of standardized methods to quantify and compare antioxidant performance. Different studies rely on disparate ROS detection assays, including DCFH-DA, DHE,

MitoSOX, ABTS, DPPH, lipid peroxidation markers, nitrotyrosine staining, malondialdehyde quantification, or indirect inflammatory readouts. These methods differ in oxidant specificity, sensitivity, subcellular resolution, and susceptibility to artifacts, making it difficult to directly compare the ROS-scavenging capacity of different nanogels or to rank materials across studies. Moreover, many polymer degradation and release assays use exogenous H_2O_2 concentrations in the micromolar to millimolar range to accelerate responsiveness, whereas physiological redox signaling often occurs at much lower and highly compartmentalized oxidant levels. As a result, chemical responsiveness under accelerated *in vitro* conditions does not always predict performance in disease-relevant oxidative microenvironments. Future studies should therefore report not only antioxidant efficiency but also the specific ROS/RNS species targeted, oxidant concentration, exposure time, detection method, material-response kinetics, and the biological context in which redox modulation is evaluated. A second critical issue concerns the balance between therapeutic ROS modulation and preservation of physiological redox signaling. ROS are not only damaging oxidants but also essential mediators of immune activation, angiogenesis, tissue remodeling, proliferation, and differentiation.

Therefore, complete or non-selective ROS depletion may impair host defense, delay tissue repair, or interfere with redox-dependent signaling cascades. This consideration is particularly relevant for catalytic scavengers, whose activity may persist beyond the initial oxidative insult, and for long-retained hybrid matrices designed for chronic diseases. Future redox-modulating platforms should be designed to restore homeostasis within a therapeutic window rather than indiscriminately suppress ROS. Spatial confinement, disease-triggered activation, tunable catalytic turnover, controlled degradability, and localized administration will be essential to minimize off-target interference while maintaining antioxidant efficacy. From a translational perspective, nanogel fate *in vivo* remains a central challenge. Clearance, biodistribution, and tissue retention depend on particle size, surface charge, hydrophilicity, degradability, protein corona formation, and administration route. Very small or highly hydrated nanogels may be rapidly eliminated through renal, hepatic, or lymphatic pathways, whereas larger, poorly degradable, or strongly opsonized systems may accumulate in the liver, spleen, or mononuclear phagocyte system. Immune recognition must also be carefully considered, since surface chemistry, cationic charge, residual monomers or crosslinkers, endotoxin contamination, adsorbed proteins, and repeated dosing can promote complement activation, macrophage uptake, or inflammatory responses. In parallel, long-term toxicity remains insufficiently explored, particularly for redox-active inorganic components or catalytic centers that may persist in tissues, release metal ions, or maintain activity beyond the desired therapeutic window. Accordingly, future investigations should move beyond short-term cytocompatibility and include repeated-dose studies, biodistribution, degradation, excretion, hemocompatibility, complement activation, cytokine release, and chronic toxicity under disease-relevant conditions. Manufacturing feasibility is another decisive barrier.

Many nanogel systems are still prepared using laboratory-scale batch methods that may generate variability in size,

polydispersity, crosslinking density, payload loading, and residual reagents. For clinical translation, critical quality attributes such as particle size distribution, surface chemistry, redox activity, residual monomer/crosslinker content, endotoxin level, sterility, storage stability, and release kinetics must be tightly controlled. Microfluidic and continuous manufacturing approaches may help improve reproducibility, scalability, and batch-to-batch consistency, especially when combined with inline monitoring and standardized purification strategies. However, scale-up should be considered early in material design rather than after biological validation because complex multicomponent architectures may offer excellent laboratory performance but poor manufacturability or regulatory feasibility. Looking forward, the development of next-generation antioxidant polymer materials will likely benefit from the integration of high-throughput experimentation, automated synthesis, advanced characterization, and artificial intelligence-assisted screening. Machine learning and data-driven design can help identify relationships between polymer composition, crosslinking chemistry, particle size, degradability, ROS specificity, payload release, cytocompatibility, and immune response. When coupled with high-throughput ROS assays and disease-relevant biological models, these approaches could accelerate the discovery of materials that respond within physiologically meaningful oxidative windows rather than only under artificially strong oxidant challenges. Importantly, AI-assisted screening will require curated and standardized datasets, including negative results and harmonized reporting of ROS species, oxidant dose, exposure time, and assay conditions. Such tools should therefore complement, rather than replace, mechanistic understanding of ROS chemistry and material-biology interactions.

Overall, polymeric nanogels and hybrid redox-modulating platforms represent a promising direction for antioxidant therapy and ROS-responsive drug delivery. Their future impact will depend not only on increasing chemical sophistication but also on improving comparability, biological relevance, translational safety, and manufacturing robustness. The field should now move from proof-of-concept antioxidant materials toward rationally designed, quantitatively benchmarked, and clinically manufacturable platforms capable of restoring redox homeostasis with spatial, temporal, and disease-specific precision.

ACKNOWLEDGMENTS

This work was partially supported by the European Union's Horizon 2020 Research and Innovation Program under Marie Skłodowska-Curie grant agreement 872648 (RISE-2019 MEPHOS).

AUTHOR CONTRIBUTIONS

G.N., K.F., and G.A. conceptualized, wrote the original manuscript, and edited the manuscript and figures. R.M.P., R.M., Y.K., E.N.Z., and P.D. edited the manuscript. H.K.-M. and F.R. conceptualized, edited the manuscript and figures, supervised the project, and provided funding. All authors contributed to discussions and finalized the manuscript.

DECLARATION OF INTERESTS

The authors declare no competing interests.

REFERENCES

- Fathi, A., Shiade, S.R.G., Saleem, A., Shohani, F., Fazeli, A., Riaz, A., Zulfikar, U., Shabaan, M., Ahmed, I., and Rahimi, M. (2025). Reactive oxygen species (ROS) and antioxidant systems in enhancing plant resilience against abiotic stress. *International Journal of Agronomy* 2025, 8834883.
- Jiang, S., Su, B., Li, G., Liu, X., Liu, P., Yang, M., Xiao, J., Huang, X., Wei, D., Sun, J., et al. (2026). Emodin-derived red-emissive carbon dots: Light-driven ROS generation and antioxidant activities in non-illuminative regimes. *Biomaterials* 324, 123496. <https://doi.org/10.1016/j.biomaterials.2025.123496>.
- Fang, Y., Yang, J., Liang, X., Wu, J., Xie, M., Zhang, K., and Su, C. (2024). Endogenous and exogenous stimuli-triggered reactive oxygen species evoke long-lived carbon monoxide to fight against lung cancer. *J. Nanobiotechnol.* 22, 416. <https://doi.org/10.1186/s12951-024-02688-x>.
- Qi, Y., Rajbanshi, B., Hao, R., Dang, Y., Xu, C., Lu, W., Dai, L., Zhang, B., and Zhang, X. (2025). The dual role of PGAM5 in inflammation. *Exp. Mol. Med.* 57, 298–311.
- Ansari, W.A., Srivastava, K., Nasibullah, M., and Khan, M.F. (2025). Reactive oxygen species (ROS): sources, generation, disease pathophysiology, and antioxidants. *Discov. Chem.* 2, 191. <https://doi.org/10.1007/s44371-025-00275-z>.
- Mittler, R. (2017). ROS Are Good. *Trends Plant Sci.* 22, 11–19. <https://doi.org/10.1016/j.tplants.2016.08.002>.
- Alhaj Sulaiman, A., and Katanaev, V.L. (2025). Beyond antioxidants: How redox pathways shape cellular signaling and disease outcomes. *Antioxidants* 14, 1142.
- Bao, L., Wang, W., Li, M., Liu, J., Liu, J., Alifu, G., Wang, D., Liang, X., Mao, T., and Zhai, Y. (2025). Reactive oxygen species-post translational modifications-central carbon metabolism regulatory loop: coordination of redox homeostasis and carbon flux allocation in plants under abiotic stress. *Front. Plant Sci.* 16, 1637328.
- Liu, S., Liu, J., Wang, Y., Deng, F., and Deng, Z. (2025). Oxidative Stress: Signaling Pathways, Biological Functions, and Disease. *MedComm* 6, e70268.
- Weinberg, S.E., and Chandel, N.S. (2025). Mitochondria reactive oxygen species signaling in immune responses. *Immunity* 58, 1904–1921.
- Michalak, K.P., and Michalak, A.Z. (2025). Understanding chronic inflammation: couplings between cytokines, ROS, NO, Cai 2+, HIF-1 α , Nrf2 and autophagy. *Front. Immunol.* 16, 1558263.
- Wang, Q., Qian, W., Han, Y., Mao, Y., Gao, Z., Chen, Y., Zeng, X., Lu, H., Jiang, L., Li, J., et al. (2025). Ferumoxylol promotes haematopoietic stem cell post-injury regeneration as a reactive oxygen species scavenger. *Nat. Nanotechnol.* 20, 959–969.
- Shen, J., Pan, Y., Han, L., Luo, L., Sun, T., and Yu, Y. (2025). Nanozymes as next-generation ROS scavengers: design strategies, catalytic mechanisms, and therapeutic frontiers. *J. Mater. Chem. B* 13, 8286–8297.
- Manful, C.F., Fordjour, E., Subramaniam, D., Sey, A.A., Abbey, L., and Thomas, R. (2025). Antioxidants and reactive oxygen species: shaping human health and disease outcomes. *Int. J. Mol. Sci.* 26, 7520.
- Jomova, K., Alomar, S.Y., Valko, R., Nepovimova, E., Kuca, K., and Valko, M. (2025). The role of redox-active iron, copper, manganese, and redox-inactive zinc in toxicity, oxidative stress, and human diseases. *EXCLI J.* 24, 880–954.
- Liu, M., Chen, J., Li, L., Zhou, J., Narain, R., Nan, K., and Chen, Y. (2024). Anti-Oxidative Tannic Acid-Based Nanogels Formed via Dynamic Benzoxaborolate Cross-Linking at Physiological pH. *J. Mol. Liq.* 398, 124239. <https://doi.org/10.1016/j.molliq.2024.124239>.
- Tiwari, R., Mondal, Y., Bharadwaj, K., Mahajan, M., Mondal, S., and Sarkar, A. (2025). Reactive Oxygen Species (ROS) and Their Profound Influence on Regulating Diverse Aspects of Cancer: A Concise Review. *Drug Dev. Res.* 86, e70107.

18. Carrasco, E., Blázquez-Castro, A., Calvo, M.I., Juarranz, Á., and Espada, J. (2016). Switching on a transient endogenous ROS production in mammalian cells and tissues. *Methods* 109, 180–189. <https://doi.org/10.1016/j.ymeth.2016.08.013>.
19. Dang, T.V.T., Cho, H.S., Lee, S., and Hwang, I. (2025). Salt stress-accelerated proteasomal degradation of LBD11 suppresses ROS-mediated meristem development and root growth in *Arabidopsis*. *Plant Commun.* 6, 101241. <https://doi.org/10.1016/j.xplc.2025.101241>.
20. García-Domínguez, M. (2025). Pathological and Inflammatory Consequences of Aging. *Biomolecules* 15, 404.
21. Han, J.M., Mwit, G., Yeom, S.J., Lim, J., Kim, W.S., Lim, S., Lim, S.T., and Byun, E.B. (2025). Radiation-Resistant Bacteria *Deinococcus radiodurans*-Derived Extracellular Vesicles as Potential Radioprotectors. *Adv. Healthcare Mater.* 14, 2403192.
22. Sharma, D.K., Soni, I., and Rajpurohit, Y.S. (2025). Surviving the storm: exploring the role of natural transformation in nutrition and DNA repair of stressed *Deinococcus radiodurans*. *Appl. Environ. Microbiol.* 91, e0137124.
23. Singh, C., Singh, R., and Shekhar, A. (2024). Oxidative stress in cadmium toxicity in animals and its amelioration. In *Cadmium Toxicity Mitigation*, A.K. Jha and N. Kumar, eds. (Springer), pp. 391–411.
24. Zhang, L., Shi, W.-Y., Xu, J.-Y., Liu, Y., Wang, S.-J., Zheng, J.-Y., Li, Y.-H., Yuan, L.-X., and Qin, L.-Q. (2024). Protective effects and mechanism of chemical-and plant-based selenocystine against cadmium-induced liver damage. *J. Hazard Mater.* 468, 133812.
25. Anglada, J.M., Crehuet, R., Martins-Costa, M.T.C., Francisco, J.S., and Ruiz-López, M.F. (2025). The reaction of sulfenic acids with OH and HO₂ radicals in different environments. *Phys. Chem. Chem. Phys.* 27, 8856–8867. <https://doi.org/10.1039/D4CP04106B>.
26. Liu, H., Wang, J., Zhao, C., Zhang, R., Wang, M., Jiang, Y., Sun, D., Hou, Y., and Yang, M. (2025). Mesoporous polydopamine@CeO₂ nanoparticles for photoacoustic-guided photothermal therapy in suppressing arthritis. *iScience* 28, 112576. <https://doi.org/10.1016/j.isci.2025.112576>.
27. Bellanti, F., Coda, A.R.D., Trecca, M.I., Lo Buglio, A., Serviddio, G., and Vendemiale, G. (2025). Redox imbalance in inflammation: the interplay of oxidative and reductive stress. *Antioxidants* 14, 656.
28. Wu, M., Liang, B., Zhang, L., Wu, B., and Liu, J. (2025). Cobalt carbonate nanorods enhance chemotherapy via neutralization of acidic tumor microenvironment and generation of carbonate radical anions for necrosis. *Colloids Surf. B Biointerfaces* 250, 114563. <https://doi.org/10.1016/j.colsurfb.2025.114563>.
29. Mao, W., Zhang, X., Li, K., Lin, W., Liu, X., Shi, Y., Liu, T., Pan, C., Liu, J., Wang, H., et al. (2025). Acid-neutralizing zinc-magnesium double hydroxide nanosheets reverse osteoporotic microenvironment by targeting the osteoclast calcium oscillation. *Cell Biomater.* 1, 100015. <https://doi.org/10.1016/j.celbio.2025.100015>.
30. Wang, S.-H., Duan, Q.-Y., Zhang, R., Wang, Z., Wang, Z.-X., Shen, Y., and Wu, F.-G. (2026). A liquid PEG depot capable of triggering intratumoral O₂/H₂O₂ generation and neutralization/calcification for realizing light irradiation-free photodynamic therapy and immunotherapy. *Chin. Chem. Lett.* 37, 111772. <https://doi.org/10.1016/j.cclet.2025.111772>.
31. Wang, D., Mu, D., Mi, Y., Zhang, G., Meng, Q., Yang, Y., Liang, D., and Hou, Y. (2025). Gnetupendin A attenuates ischemic stroke injury by modulating oxidative stress through AMPK/SIRT1 pathway. *Biochem. Pharmacol.* 242, 117294. <https://doi.org/10.1016/j.bcp.2025.117294>.
32. Wang, M., Xiao, Y., Miao, J., Zhang, X., Liu, M., Zhu, L., Liu, H., Shen, X., Wang, J., Xie, B., and Wang, D. (2025). Oxidative Stress and Inflammation: Drivers of Tumorigenesis and Therapeutic Opportunities. *Antioxidants* 14, 735.
33. Min, Z., Zou, Y., Meng, Y., Liu, X., Li, H., Liu, H., and Liu, J. (2025). Harnessing Redox: Biocomposites Modulate Macrophage-Stem Cell Dynamics in Osteo-Inflammation. *Tissue Eng. B Rev.* <https://doi.org/10.1177/19373341251377651>.
34. Fan, Y.-Y., Wu, H., and Xu, C. (2025). ROS-responsive nanoplatforms for targeted tumor immunomodulation: a paradigm shift in precision cancer immunotherapy. *Pharmaceutics* 17, 886.
35. Nunziata, G., Nava, M., Lacroce, E., Pizzetti, F., and Rossi, F. (2025). Thermo-Responsive Polymer-Based Nanoparticles: From Chemical Design to Advanced Applications. *Macromol. Rapid Commun.* 46, 2401127.
36. Li, Z., Zhao, Q., Zhou, J., Li, Y., Zheng, Y., and Chen, L. (2025). A reactive oxygen species-responsive hydrogel loaded with Apelin-13 promotes the repair of spinal cord injury by regulating macrophage M1/M2 polarization and neuroinflammation. *J. Nanobiotechnol.* 23, 12. <https://doi.org/10.1186/s12951-024-02978-4>.
37. Namdar, N., Najafipour, S., Maghbool, M., Goodarzi, A., Heiran, R., Nasiri-Ghiri, M., Alipanah, H., and Osanloo, M. (2025). Multifunctional carboxymethylcellulose-based nanogel containing *Matricaria chamomilla* essential oil: A practical approach to combating skin aging in rats. *Int. J. Biol. Macromol.* 334, 149183. <https://doi.org/10.1016/j.ijbiomac.2025.149183>.
38. Zhang, Y., Jin, Z., Xu, L., Zhong, Z., Wang, X., Gao, C., and Li, L. (2026). ROS-scavenging nanoparticles loaded with tectorigenin protect against acetaminophen-induced hepatotoxicity by interrupting the calcium/ROS-mediated pathogenic endoplasmic reticulum–Mitochondrial signaling cascade. *Bioact. Mater.* 58, 408–421. <https://doi.org/10.1016/j.bioactmat.2025.12.016>.
39. Lacroce, E., Nunziata, G., Cianniello, F., Limiti, E., Rainer, A., Vangosa, F.B., Sacchetti, A., Sponchioni, M., and Rossi, F. (2024). Amphiphilic pH-responsive core-shell nanoparticles can increase the performances of cellulose-based drug delivery systems. *Int. J. Biol. Macromol.* 283, 137659. <https://doi.org/10.1016/j.ijbiomac.2024.137659>.
40. Vasvani, S., Vasukutty, A., Bardhan, R., Park, I.-K., and Uthaman, S. (2024). Reactive oxygen species driven prodrug-based nanoscale carriers for transformative therapies. *Biomater. Sci.* 12, 4335–4353.
41. Zhang, M., Ran, S., Yin, X., Zhang, J., Sun, X., Sun, W., and Zhu, Z. (2023). Mesoporous polydopamine nanoplatforms loaded with calcium ascorbate for amplified oxidation and photothermal combination cancer therapy. *BMEMat* 1, e12041.
42. Hu, D., Li, Y., Li, R., Wang, M., Zhou, K., He, C., Wei, Q., and Qian, Z. (2024). Recent advances in reactive oxygen species (ROS)-responsive drug delivery systems for photodynamic therapy of cancer. *Acta Pharm. Sin. B* 14, 5106–5131. <https://doi.org/10.1016/j.apsb.2024.10.015>.
43. Xie, Y., He, P., Li, X., Zhang, L., Yu, Y., and Chen, F. (2025). Research progress of reactive oxygen species (ROS) scavenging nanoplatforms in the treatment of ocular diseases. *J. Contr. Release* 386, 114078. <https://doi.org/10.1016/j.jconrel.2025.114078>.
44. Checa, J., and Aran, J.M. (2020). Reactive oxygen species: drivers of physiological and pathological processes. *J. Inflamm. Res.* 13, 1057–1073.
45. Dash, U.C., Bhol, N.K., Swain, S.K., Samal, R.R., Nayak, P.K., Raina, V., Panda, S.K., Kerry, R.G., Duttaroy, A.K., and Jena, A.B. (2025). Oxidative stress and inflammation in the pathogenesis of neurological disorders: Mechanisms and implications. *Acta Pharm. Sin. B* 15, 15–34. <https://doi.org/10.1016/j.apsb.2024.10.004>.
46. Pooja, G., Shweta, S., and Patel, P. (2025). Oxidative stress and free radicals in disease pathogenesis: a review. *Discov. Med.* 2, 104. <https://doi.org/10.1007/s44337-025-00303-y>.
47. Endale, H.T., Tesfaye, W., and Mengstie, T.A. (2023). ROS induced lipid peroxidation and their role in ferroptosis. *Front. Cell Dev. Biol.* 11, 1226044.
48. Sies, H., and Jones, D.P. (2020). Reactive oxygen species (ROS) as pleiotropic physiological signalling agents. *Nat. Rev. Mol. Cell Biol.* 21, 363–383.

49. Zhu, H., Cen, J., Hong, C., Wang, H., Wen, Y., He, Q., Yu, Y., Cao, J., and Chen, W. (2023). Targeting Labile Iron-Mediated Ferroptosis Provides a Potential Therapeutic Strategy for Rhabdomyolysis-Induced Acute Kidney Injury. *ACS Chem. Biol.* 18, 1294–1304. <https://doi.org/10.1021/acscchembio.2c00914>.
50. Li, J., Cao, F., Yin, H.L., Huang, Z.J., Lin, Z.T., Mao, N., Sun, B., and Wang, G. (2020). Ferroptosis: past, present and future. *Cell Death Dis.* 11, 88.
51. Su, L.-J., Zhang, J.-H., Gomez, H., Murugan, R., Hong, X., Xu, D., Jiang, F., and Peng, Z.-Y. (2019). Reactive oxygen species-induced lipid peroxidation in apoptosis, autophagy, and ferroptosis. *Oxid. Med. Cell. Longev.* 2019, 1–13.
52. Chen, Y., Dai, Y., Huang, Y., Zhang, L., Zhang, C., Gao, H., and Yan, Q. (2025). Inhibition of tubular epithelial cells ferroptosis alleviates renal interstitial fibrosis by reducing lipid hydroperoxides and TGF- β /Smad signaling. *Cell Commun. Signal.* 23, 81. <https://doi.org/10.1186/s12964-025-02068-4>.
53. Peng, Z., Jia, Q., Mao, J., Jiang, S., Ma, Q., Luo, X., An, Z., Huang, A., Ma, C., and Yi, Q. (2025). The role of ferroptosis and oxidative stress in cognitive deficits among chronic schizophrenia patients: a multicenter investigation. *Schizophrenia* 11, 4.
54. Ciurea, A.V., Mohan, A.G., Covache-Busuioc, R.-A., Costin, H.-P., Glavan, L.-A., Corlatescu, A.-D., and Saceleanu, V.M. (2023). Unraveling molecular and genetic insights into neurodegenerative diseases: advances in understanding Alzheimer's, Parkinson's, and Huntington's diseases and amyotrophic lateral sclerosis. *Int. J. Mol. Sci.* 24, 10809.
55. Domanskyi, A., and Parlato, R. (2022). Oxidative Stress in Neurodegenerative Diseases. *Antioxidants* 11, 504. <https://doi.org/10.3390/antiox11030504>.
56. Xin, Y., Fang, F., Yue, Q., Luo, Y., Tian, S., Cheng, L., Wang, X., Yang, X., Luo, L., and Meng, F. (2024). Microenvironment modulating nanogels by Shiitake-derived lentinan and a reactive oxygen species scavenging conjugated polymer for the treatment of Alzheimer's disease. *Nano Today* 55, 102178. <https://doi.org/10.1016/j.nantod.2024.102178>.
57. Pang, M., Song, X., Miao, Y., Wang, Y., Zhou, C., Geng, Z., Du, J., Mousian, B., Su, Y., Liu, X., and Ming, D. (2023). Aconitum carmichaelii triggers neurotoxicity and Parkinson-like symptoms through initiation of ROS-mitochondrial apoptosis and the Nox5/DJ-1 signaling pathway. *BMEMat* 1, e12033.
58. Chen, Y., Luo, X., Yin, Y., Thomas, E.R., Liu, K., Wang, W., and Li, X. (2025). The interplay of iron, oxidative stress, and α -synuclein in Parkinson's disease progression. *Mol. Med.* 31, 154.
59. Levi, S., Ripamonti, M., Moro, A.S., and Cozzi, A. (2024). Iron imbalance in neurodegeneration. *Mol. Psychiatr.* 29, 1139–1152.
60. Chiurchiù, V., and Maccarrone, M. (2011). Chronic inflammatory disorders and their redox control: from molecular mechanisms to therapeutic opportunities. *Antioxid. Redox Signal.* 15, 2605–2641.
61. Yuan, H., Yi, N., Li, D., Xu, C., Yin, G.-R., Zhuang, C., Wang, Y.-J., and Ni, S. (2024). PPAR γ regulates osteoarthritis chondrocytes apoptosis through caspase-3 dependent mitochondrial pathway. *Sci. Rep.* 14, 11237. <https://doi.org/10.1038/s41598-024-62116-w>.
62. Bolduc, J.A., Collins, J.A., and Loeser, R.F. (2019). Reactive oxygen species, aging and articular cartilage homeostasis. *Free Radic. Biol. Med.* 132, 73–82.
63. Tan, S., Sun, Y., Li, S., Wu, H., and Ding, Y. (2025). The impact of mitochondrial dysfunction on osteoarthritis cartilage: current insights and emerging mitochondria-targeted therapies. *Bone Res.* 13, 77. <https://doi.org/10.1038/s41413-025-00460-x>.
64. Jing, W., Liu, C., Su, C., Liu, L., Chen, P., Li, X., Zhang, X., Yuan, B., Wang, H., and Du, X. (2023). Role of reactive oxygen species and mitochondrial damage in rheumatoid arthritis and targeted drugs. *Front. Immunol.* 14, 1107670.
65. Tang, J., Xia, J., Gao, H., Jiang, R., Xiao, L., Sheng, H., and Lin, J. (2024). IL33-induced neutrophil extracellular traps (NETs) mediate a positive feedback loop for synovial inflammation and NET amplification in rheumatoid arthritis. *Exp. Mol. Med.* 56, 2602–2616. <https://doi.org/10.1038/s12276-024-01351-7>.
66. Chen, T., Zhou, Z., Liu, Y., Xu, J., Zhu, C., Sun, R., Hu, H., Liu, Y., Dai, L., Holmdahl, R., et al. (2024). Neutrophils with low production of reactive oxygen species are activated during immune priming and promote development of arthritis. *Redox Biol.* 78, 103401. <https://doi.org/10.1016/j.redox.2024.103401>.
67. Zhu, X., Lu, H., Jia, H., Wei, X., Xue, J., Li, W., Zhang, J., Wang, Y., Yan, J., Sun, H., et al. (2025). Ferostatin-1 reduces the inflammatory response of rheumatoid arthritis by decreasing the antigen presenting function of fibroblast-like synoviocytes. *J. Transl. Med.* 23, 280. <https://doi.org/10.1186/s12967-025-06300-0>.
68. Zhang, S., Wang, L., Kang, Y., Wu, J., and Zhang, Z. (2023). Nanomaterial-based reactive oxygen species scavengers for osteoarthritis therapy. *Acta Biomater.* 162, 1–19. <https://doi.org/10.1016/j.actbio.2023.03.030>.
69. Olivares-Vicente, M., and Herranz-López, M. (2025). The interplay between oxidative stress and lipid composition in obesity-induced inflammation: Antioxidants as therapeutic agents in metabolic diseases. *Int. J. Mol. Sci.* 26, 8544.
70. Volpe, C.M.O., Villar-Delfino, P.H., dos Anjos, P.M.F., and Nogueira-Machado, J.A. (2018). Cellular death, reactive oxygen species (ROS) and diabetic complications. *Cell Death Dis.* 9, 119. <https://doi.org/10.1038/s41419-017-0135-z>.
71. Hunt, M., Torres, M., Bachar-Wikstrom, E., and Wikstrom, J.D. (2024). Cellular and molecular roles of reactive oxygen species in wound healing. *Commun. Biol.* 7, 1534.
72. Piechota-Polanczyk, A., and Fichna, J. (2014). The role of oxidative stress in pathogenesis and treatment of inflammatory bowel diseases. *Naunyn-Schmiedeberg's Arch Pharmacol* 387, 605–620.
73. Zhang, Y., Hao, M., Yang, X., Zhang, S., Han, J., Wang, Z., and Chen, H.-N. (2024). Reactive oxygen species in colorectal cancer adjuvant therapies. *Biochim. Biophys. Acta. Mol. Basis Dis.* 1870, 166922.
74. Fuloria, S., Subramaniam, V., Karupiah, S., Kumari, U., Sathasivam, K., Meenakshi, D.U., Wu, Y.S., Sekar, M., Chitranshi, N., Malviya, R., et al. (2021). Comprehensive review of methodology to detect reactive oxygen species (ROS) in mammalian species and establish its relationship with antioxidants and cancer. *Antioxidants* 10, 128.
75. Murphy, M.P., Bayir, H., Belousov, V., Chang, C.J., Davies, K.J.A., Davies, M.J., Dick, T.P., Finkel, T., Forman, H.J., Janssen-Heininger, Y., et al. (2022). Guidelines for measuring reactive oxygen species and oxidative damage in cells and in vivo. *Nat. Metab.* 4, 651–662.
76. Akter, S., Khan, M.S., Smith, E.N., and Flashman, E. (2021). Measuring ROS and redox markers in plant cells. *RSC Chem. Biol.* 2, 1384–1401.
77. Aranda, A., Sequedo, L., Tolosa, L., Quintas, G., Burello, E., Castella, J.F., and Gombau, L. (2013). Dichloro-dihydro-fluorescein diacetate (DCFH-DA) assay: a quantitative method for oxidative stress assessment of nanoparticle-treated cells. *Toxicol. Vitro* 27, 954–963.
78. Kauffman, M.E., Kauffman, M.K., Traore, K., Zhu, H., Trush, M.A., Jia, Z., and Li, Y.R. (2016). MitoSOX-based flow cytometry for detecting mitochondrial ROS. *React. Oxyg. Species* 2, 361–370.
79. Shah, P., and Modi, H. (2015). Comparative study of DPPH, ABTS and FRAP assays for determination of antioxidant activity. *Int. J. Res. Appl. Sci. Eng. Technol.* 3, 636–641.
80. Dutta, R.K., Nenavathu, B.P., Gangishetty, M.K., and Reddy, A.V.R. (2012). Studies on antibacterial activity of ZnO nanoparticles by ROS induced lipid peroxidation. *Colloids Surf. B Biointerfaces* 94, 143–150.
81. Samoylenko, A., Hossain, J.A., Mennerich, D., Kellokumpu, S., Hiltunen, J.K., and Kietzmann, T. (2013). Nutritional countermeasures targeting reactive oxygen species in cancer: from mechanisms to biomarkers and clinical evidence. *Antioxid. Redox Signal.* 19, 2157–2196.

82. Yang, B., Chen, Y., and Shi, J. (2019). Reactive oxygen species (ROS)-based nanomedicine. *Chem. Rev.* **119**, 4881–4985.
83. Karpinska, B., and Foyer, C.H. (2024). Superoxide signalling and antioxidant processing in the plant nucleus. *J. Exp. Bot.* **75**, 4599–4610.
84. Chevriaux, J., Zerbetto De Palma, G., Alleva, K., and Zeida, A. (2025). Hydrogen peroxide transport by aquaporins: insights from molecular modeling and simulations. *Biophys. Rev.* **17**, 301–308.
85. Sies, H. (2017). Hydrogen peroxide as a central redox signaling molecule in physiological oxidative stress: Oxidative eustress. *Redox Biol.* **11**, 613–619.
86. Mansour, S.A., Alshehab, A., Fouad, M.K., and Tony, M.A. (2026). From Framework to Radical Pathways: A Critical Review on MOF-Driven Fenton and Fenton-Like Catalysis. *Int. J. Environ. Res.* **20**, 102.
87. Valgimigli, L. (2023). Lipid peroxidation and antioxidant protection. *Biomolecules* **13**, 1291.
88. Sun, J., Cao, X., Lu, W., Wei, Y., Kong, L., Chen, W., Shao, X., and Wang, Y. (2023). Recent advances in fluorescent probes of peroxynitrite: Structural, strategies and biological applications. *Theranostics* **13**, 1716–1744.
89. Li, C., Deng, Z., and Gillies, E.R. (2023). Designing polymers with stimuli-responsive degradation for biomedical applications. *Curr. Opin. Biomed. Eng.* **25**, 100437.
90. de Gracia Lux, C., Joshi-Barr, S., Nguyen, T., Mahmoud, E., Schopf, E., Fomina, N., and Almutairi, A. (2012). Biocompatible polymeric nanoparticles degrade and release cargo in response to biologically relevant levels of hydrogen peroxide. *J. Am. Chem. Soc.* **134**, 15758–15764.
91. Jomova, K., Raptova, R., Alomar, S.Y., Alwasel, S.H., Nepovimova, E., Kuca, K., and Valko, M. (2023). Reactive oxygen species, toxicity, oxidative stress, and antioxidants: chronic diseases and aging. *Arch. Toxicol.* **97**, 2499–2574. <https://doi.org/10.1007/s00204-023-03562-9>.
92. Kumar, S., Saxena, J., Srivastava, V.K., Kaushik, S., Singh, H., Abo-EL-Sooud, K., Abdel-Daim, M.M., Jyoti, A., and Saluja, R. (2022). The Interplay of Oxidative Stress and ROS Scavenging: Antioxidants as a Therapeutic Potential in Sepsis. *The Interplay of Oxidative Stress and ROS Scavenging: Antioxidants as a Therapeutic Potential in Sepsis* **10**, 1575. <https://doi.org/10.3390/vaccines10101575>.
93. Mirończuk-Chodakowska, I., Witkowska, A.M., and Zujko, M.E. (2018). Endogenous non-enzymatic antioxidants in the human body. *Adv. Med. Sci.* **63**, 68–78. <https://doi.org/10.1016/j.advms.2017.05.005>.
94. Chandimali, N., Bak, S.G., Park, E.H., Lim, H.-J., Won, Y.-S., Kim, E.-K., Park, S.-I., and Lee, S.J. (2025). Free radicals and their impact on health and antioxidant defenses: a review. *Cell Death Discov.* **11**, 19. <https://doi.org/10.1038/s41420-024-02278-8>.
95. Lee, D., Huntoon, K., Wang, Y., Kang, M., Lu, Y., Jeong, S.D., Link, T.M., Gallup, T.D., Qie, Y., Li, X., et al. (2024). Synthetic cationic helical polypeptides for the stimulation of antitumor innate immune pathways in antigen-presenting cells. *Nat. Biomed. Eng.* **8**, 593–610.
96. Lacroce, E., Pizzetti, F., Urrego, N.M.B., Nunziata, G., Masi, M., and Rossi, F. (2024). Magnetically Active Bicontinuous Polymer Structures for Multiple Controlled Drug Delivery. *Magnetically Active Bicontinuous Polymer Structures for Multiple Controlled Drug Delivery*, 2400084.
97. Bobbala, S., Allen, S.D., and Scott, E.A. (2018). Flash nanoprecipitation permits versatile assembly and loading of polymeric bicontinuous cubic nanospheres. *Nanoscale* **10**, 5078–5088.
98. He, Y., Rui, W., Yan, Z., Feng, W., Zhao, C., and Yan, H. (2024). Recent Advances of Organic-Inorganic Hybrid Fluorescent Hyperbranched Polymer: Synthesis, Performance Regulation Strategies and Applications. *ChemPlusChem* **89**, e202400302.
99. Lee, K., Kim, T.-H., Jo, S.-H., and Yu, S. (2024). Adsorption effects of electron scavengers and inorganic ions on catalysts for catalytic oxidation of sulfamethoxazole in radiation treatment. *Chemosphere* **354**, 141675.
100. Niella, R.V., Corrêa, J.M.X., Marques, C.S.d.C., Silva, Á.J.C., Santos, L.C., Oliveira, I.S.d., DeFreitas-Silva, G., Rebouças, J.S., Silva, J.F., and de Lavor, M.S.L. (2025). Manganese Porphyrin Treatment Improves Redox Status Caused by Acute Compressive Spinal Cord Trauma. *Antioxidants* **14**, 587. <https://doi.org/10.3390/antiox14050587>.
101. Ma, Y., Pan, J., Ju, C., Yu, X., Wang, Y., Li, R., Hu, H., Wang, X., and Hao, D. (2025). Antioxidant nanozymes: current status and future perspectives in spinal cord injury treatments. *Theranostics* **15**, 6146–6183.
102. Zhang, S., Cao, J., Spasojevic, I., Treggiari, M., and Sheng, H. (2025). Treatment with Manganese Porphyrin, MnTnBuOE-2-PyP5+, Suppressed the Activation of Macrophages in a Mouse Intracerebral Hemorrhage. *Pharmaceuticals* **18**, 547.
103. Kwon, H.-Y., Jung, Y., Jeon, H., and Han, H.-S. (2025). Investigation into recent advanced strategies of reactive oxygen species-mediated therapy based on Prussian blue: Conceptualization and prospect. *Bioact. Mater.* **48**, 71–99. <https://doi.org/10.1016/j.bioactmat.2025.01.023>.
104. Wei, W., Wang, Y., Sun, G., Chen, Z., and Zhang, S. (2025). Prussian blue nanoparticles mitigate inflammatory osteolysis by reducing oxidative stress and enhancing endogenous antioxidant systems. *Colloids Surf. B Biointerfaces* **254**, 114806. <https://doi.org/10.1016/j.colsurfb.2025.114806>.
105. Pansambal, S., Oza, R., Borgave, S., Chauhan, A., Bardapurkar, P., Vyas, S., and Ghotekar, S. (2023). Bioengineered cerium oxide (CeO2) nanoparticles and their diverse applications: a review. *Appl. Nanosci.* **13**, 6067–6092. <https://doi.org/10.1007/s13204-022-02574-8>.
106. Kim, Y.G., Lee, Y., Lee, N., Soh, M., Kim, D., and Hyeon, T. (2024). Ceria-based therapeutic antioxidants for biomedical applications. *Adv. Mater.* **36**, 2210819.
107. Yu, H., Cao, L., Li, F., Wu, Q., Li, Q., Wang, S., and Guo, Y. (2015). The antioxidant mechanism of nitroxide TEMPO: Scavenging with glutathionyl radicals. *RSC Adv.* **5**, 63655–63661. <https://doi.org/10.1039/C5RA06129F>.
108. Sultani, H.N., Morgan, I.M., Hussain, H., Haeri, H.H., Hinderberger, D., Kaluderović, G.N., and Westermann, B. (2025). Synthesis of Rhodamine TEMPO Conjugates via Isonitrile-Based Multicomponent-Reactions for Mitochondria-Targeted ROS-Detection in Cancer Cells. *Adv. Sens. Res.* **4**, 2400180.
109. DeJulius, C.R., Dollinger, B.R., Kavanaugh, T.E., Dailing, E., Yu, F., Gu-lati, S., Miskalis, A., Zhang, C., Uddin, J., Dikalov, S., and Duvall, C.L. (2021). Optimizing an Antioxidant TEMPO Copolymer for Reactive Oxygen Species Scavenging and Anti-Inflammatory Effects in Vivo. *Bioconjug. Chem.* **32**, 928–941. <https://doi.org/10.1021/acs.bioconjchem.1c00081>.
110. Yang, F., Guo, G., and Wang, Y. (2022). Inflammation-triggered dual release of nitroxide radical and growth factor from heparin mimicking hydrogel-tissue composite as cardiovascular implants for anti-coagulation, endothelialization, anti-inflammation, and anti-calcification. *Biomaterials* **289**, 121761. <https://doi.org/10.1016/j.biomaterials.2022.121761>.
111. Liu, J., Shen, K., Chu, D., and Cheng, Y. (2025). Tempo-Functionalized Janus Hydrogel Crosslinked by Tandem Dynamic Covalent Bonds for Internal Organ Injury Repair and Deep Second-Degree Burn Wound Healing. *Adv. Funct. Mater.* **35**, 2505194.
112. Ding, Q., Zhang, Z., Li, M., Zhu, J.-H., Fu, W., He, M., Bai, Y., Zhang, Z., Li, S., Wang, L., et al. (2025). Subcellular targeting strategies: Chemical structure-based design concepts for bioimaging and theranostics. *Cell Biomater.* **7**, 100001.
113. Lin, L., Chen, Z., Huang, C., Wu, Y., Huang, L., Wang, L., Ke, S., and Liu, L. (2023). Mito-TEMPO, a Mitochondria-Targeted Antioxidant, Improves Cognitive Dysfunction due to Hypoglycemia: an Association with Reduced Pericyte Loss and Blood-Brain Barrier Leakage. *Mol. Neurobiol.* **60**, 672–686. <https://doi.org/10.1007/s12035-022-03101-0>.
114. Calabrese, G., Ardizzone, A., Campolo, M., Conoci, S., Esposito, E., and Paterniti, I. (2021). Beneficial Effect of Tempol, a Membrane-Permeable Radical Scavenger, on Inflammation and Osteoarthritis in In Vitro Models. *Biomolecules* **11**, 352. <https://doi.org/10.3390/biom11030352>.

115. Sewelam, A.S., Amin, M.A.S., Abdelmohsen, S.R., Mohammed, O.A., Hashish, A.A., Alfaifi, J., and Aboregela, A.M. (2024). Tempol maintained the cellular integrity of the cerebellar cortex by preserving neuron survival, autophagy, glial cells, and synapses after cisplatin exposure. *Transl. Res. Anat.* 35, 100298. <https://doi.org/10.1016/j.tria.2024.100298>.
116. Nah, Y., Kim, J., Lee, S., Koh, W.-G., and Kim, W.J. (2024). Tailored small molecule for inflammation treatment: Dual scavenger targeting nitric oxide and reactive oxygen species. *J. Contr. Release* 374, 525–537. <https://doi.org/10.1016/j.jconrel.2024.08.026>.
117. Xu, J., Zhang, Y., Yao, X., Wang, S., Lv, K., Luo, G., Wang, J., and Li, G. (2024). Intestinal Targeted Nanogel with Broad-Spectrum Autonomous ROS Scavenging Performance for Enhancing the Bioactivity of trans-R-sveratrol. *Int. J. Nanomed.* 19, 5995–6014. <https://doi.org/10.2147/IJN.S464849>.
118. Biji, C.A., Balde, A., Kim, S.-K., and Nazeer, R.A. (2024). Optimization of alginate/carboxymethyl chitosan microbeads for the sustained release of celecoxib and attenuation of intestinal inflammation in vitro. *Int. J. Biol. Macromol.* 282, 137022. <https://doi.org/10.1016/j.ijbiomac.2024.137022>.
119. Zhang, L., Li, W., Cao, J., Yang, Y., and Gao, H. (2025). Injectable reactive oxygen and nitrogen species-scavenging hydrogels for repairing traumatic brain injury. *Mater. Today Bio* 35, 102336. <https://doi.org/10.1016/j.mtbio.2025.102336>.
120. Qin, Z., Lan, T., Xu, J., Wang, X., Zhang, N., Shi, S., and Guo, Y. (2026). Metal-free carbon dots as a comprehensive and robust antioxidant nanozyme for scavenging reactive oxygen species in cigarette smoke. *Carbon* 247, 120993. <https://doi.org/10.1016/j.carbon.2025.120993>.
121. Fan, L., Yang, F., Jiang, M., Li, Q., Bai, X., Feng, P., and Pan, H. (2025). V-doped CeO₂-loaded biopolymer scaffold for reactive oxygen species scavenging and osteogenic enhancement. *Polym. Test.* 152, 109012. <https://doi.org/10.1016/j.polymertesting.2025.109012>.
122. Kim, Y.E., and Kim, J. (2022). ROS-Scavenging Therapeutic Hydrogels for Modulation of the Inflammatory Response. *ACS Appl. Mater. Interfaces* 14, 23002–23021. <https://doi.org/10.1021/acsami.1c18261>.
123. Dai, Y., Guo, Y., Tang, W., Chen, D., Xue, L., Chen, Y., Guo, Y., Wei, S., Wu, M., Dai, J., and Wang, S. (2024). Reactive oxygen species-scavenging nanomaterials for the prevention and treatment of age-related diseases. *J. Nanobiotechnol.* 22, 252. <https://doi.org/10.1186/s12951-024-02501-9>.
124. Badparvar, F., Marjani, A.P., Salehi, R., and Ramezani, F. (2023). pH/redox responsive size-switchable intelligent nanovehicle for tumor microenvironment targeted DOX release. *Sci. Rep.* 13, 22475. <https://doi.org/10.1038/s41598-023-49446-x>.
125. Nunziata, G., Limiti, E., Aramini, D., Nava, M., Moretti, L., Rainer, A., Sponchioni, M., and Rossi, F. (2025). pH-Thermo Dual-Responsive Polymeric Nanoparticles for Women's Health: Dual Action Against Cervical and Ovarian Cancer Cells. *ACS Appl. Mater. Interfaces* 17, 61888–61904.
126. Cabral, H., Li, J., Miyata, K., and Kataoka, K. (2023). Controlling the bio-distribution and clearance of nanomedicines. *Nat. Rev. Bioeng.* 2, 214–232.
127. Yi, H., Lu, W., Liu, F., Zhang, G., Xie, F., Liu, W., Wang, L., Zhou, W., and Cheng, Z. (2021). ROS-responsive liposomes with NIR light-triggered doxorubicin release for combinatorial therapy of breast cancer. *J. Nanobiotechnol.* 19, 134.
128. Nunziata, G., Sponchioni, M., and Rossi, F. (2026). Programmable shrinking and aggregation of pH-thermo dual-responsive amphiphilic polymeric nanoparticles governed by molecular architecture. *RSC Appl. Interfaces*. <https://doi.org/10.1039/D6LF00061D>.
129. Bodega, G., Alique, M., Puebla, L., Carracedo, J., and Ramirez, R.M. (2019). Microvesicles: ROS scavengers and ROS producers. *J. Extracell. Vesicles* 8, 1626654.
130. Khan, M., Ullah, R., Shah, S.M., Farooq, U., and Li, J. (2025). Manganese-Based nanotherapeutics for targeted treatment of breast cancer. *ACS Appl. Bio Mater.* 8, 3571–3600.
131. Chen, J., Hu, H., Fu, J., He, Y., Zhou, H., Lei, J., Huang, M., Wang, J., Liu, A., and Liu, Z. (2026). NOX4/Keap1/Nrf2/ROS signaling drives ferroptosis in trimethyltin chloride-induced cardiac developmental malformations. *Toxicology* 521, 154394.
132. Sultana, F., Imran-Ul-Haque, M., Arafat, M., and Sharmin, S. (2013). An overview of nanogel drug delivery system. *J. Appl. Pharmaceut. Sci.* 3, S95–S105.
133. Kazakov, S. (2016). Liposome-nanogel structures for future pharmaceutical applications: an updated review. *Curr. Pharm. Des.* 22, 1391–1413.
134. Schifone, M., Nunziata, G., and Rossi, F. (2026). Modelling the impact of temperature on nanocarrier behavior: Thermodynamics, structural transitions, and drug release. *Adv. Colloid Interface Sci.* 355, 103913. <https://doi.org/10.1016/j.cis.2026.103913>.
135. Sun, C., Lin, X., Du, W., and Tian, J. (2026). Research progress in immobilized nanozymes based on hybrid nanoflowers, MOFs and nanogels. *Mikrochim. Acta* 193, 449.
136. Xue, A., Fan, N., Yue, L., Li, X., Wang, X., Liao, H., Xiao, Z., and Wang, Z. (2025). Salicylic acid-functionalized nanogels enhance tobacco mosaic virus resistance and alleviate oxidative stress in crops. *Ind. Crops Prod.* 236, 122009. <https://doi.org/10.1016/j.indcrop.2025.122009>.
137. Ibrahim, R.E., Elshopakey, G.E., AlSaqufi, A.S., Mansour, A.T., Alkhamis, Y., Hassanien, H.A., Abbas, A., Ismail, S.H., Khamis, T., and Abdel Rahman, A.N. (2025). Chitosan nanogel mitigates Shewanella-induced oxidative stress, brain neurotransmitter imbalance, biochemical, hepatic, and renal dysfunction, and histopathological changes in Nile tilapia. *Comp. Biochem. Physiol. B Biochem. Mol. Biol.* 280, 111151. <https://doi.org/10.1016/j.cbpb.2025.111151>.
138. Singh, N., Anand, S.K., Sharma, A., Singh, S., Kakkar, P., and Srivastava, V. (2024). Chitosan/alginate nanogel potentiate berberine uptake and enhance oxidative stress mediated apoptotic cell death in HepG2 cells. *Int. J. Biol. Macromol.* 257, 128717. <https://doi.org/10.1016/j.ijbiomac.2023.128717>.
139. Nunziata, G., Pizzetti, F., Veneruso, V., Rossetti, A., Petillo, E., Frigerio, E., Marabelli, C., Tiboni, M., Casettari, L., Veglianesi, P., et al. (2026). On the comparison between organic batch and aqueous microfluidic PEG-Jeffamine nanogels synthesis for sustained drug release. *Mater. Today Chem.* 53, 103587. <https://doi.org/10.1016/j.mtchem.2026.103587>.
140. Attama, A.A., Nnamani, P.O., Onokala, O.B., Ugwu, A.A., and Onugwu, A.L. (2022). Nanogels as target drug delivery systems in cancer therapy: A review of the last decade. *Front. Pharmacol.* 13, 874510. <https://doi.org/10.3389/fphar.2022.874510>.
141. Duan, Q.-Y., Zhu, Y.-X., Jia, H.-R., Wang, S.-H., and Wu, F.-G. (2023). Nanogels: Synthesis, properties, and recent biomedical applications. *Prog. Mater. Sci.* 139, 101167. <https://doi.org/10.1016/j.pmatsci.2023.101167>.
142. Mauri, E., Giannitelli, S.M., Trombetta, M., and Rainer, A. (2021). Synthesis of Nanogels: Current Trends and Future Outlook. *Gels* 7, 36. <https://doi.org/10.3390/gels7020036>.
143. Giannitelli, S.M., Limiti, E., Mozetic, P., Pinelli, F., Han, X., Abbruzzese, F., Basoli, F., Del Rio, D., Scialla, S., Rossi, F., et al. (2022). Droplet-based microfluidic synthesis of nanogels for controlled drug delivery: tailoring nanomaterial properties via pneumatically actuated flow-focusing junction. *Nanoscale* 14, 11415–11428. <https://doi.org/10.1039/D2NR00827K>.
144. Li, H., Yang, X., and Sun, M. (2025). Self-strengthened cascade-explosive nanogel using host-guest interaction strategy for synergistic tumor treatment. *Chin. Chem. Lett.* 36, 110651. <https://doi.org/10.1016/j.ccl.2024.110651>.
145. Abedi, F., Ghandforoushan, P., Adeli, F., Yousefnezhad, M., Mohammadi, A., Moghaddam, S.V., and Davaran, S. (2023). Development of

- stimuli-responsive nanogels as drug carriers and their biomedical application in 3D printing. *Mater. Today Chem.* 29, 101372. <https://doi.org/10.1016/j.mtchem.2022.101372>.
146. Qi, L., Huang, D., Kou, H., Chernatynskaya, A., Ercal, N., and Yang, H. (2025). Synthesis and Characterization of Free Radical Scavenging Dendrimer Nanogels via Cross-Linking Reaction-Enabled Flash Nanoprecipitation. *Biomacromolecules* 26, 2986–2995. <https://doi.org/10.1021/acs.biomac.5c00050>.
147. Basak, S., Mukherjee, I., and Das, T.K. (2024). Injectable biocompatible RAFT mediated nitroxide nanogels: A robust ROS-reduction antioxidant approach. *Colloids Surf. B Biointerfaces* 236, 113790. <https://doi.org/10.1016/j.colsurf.2024.113790>.
148. Rajabi, M., and Feyzbakhsh, H. (2025). Polymeric nanocarriers in drug delivery, bioimaging, and biosensing: advances and perspectives on nanomicelles, nanogels, and dendrimers. *RSC Adv.* 15, 40400–40438. <https://doi.org/10.1039/d5ra05399d>.
149. Pareek, A., Kumar, S., Kapoor, D.U., and Prajapati, B.G. (2025). Advancements in superparamagnetic nanogels: A dual-role platform for diagnosis and targeted drug delivery. *Int. J. Pharm.* 677, 125683. <https://doi.org/10.1016/j.ijpharm.2025.125683>.
150. Xu, D., Qiao, T., Zhou, Y.-M., Wu, X.-Y., and Cui, Y.-L. (2025). A brain-targeted and ROS-responsive natural polysaccharide nanogel for enhancing antidepressant therapy. *Chem. Eng. J.* 507, 160719. <https://doi.org/10.1016/j.cej.2025.160719>.
151. Nunziata, G., Pizzetti, F., Veglianesi, P., Forloni, G., Balducci, C., and Rossi, F. (2025). Can nanoparticle-based intranasal delivery systems revolutionize treatment of central nervous system diseases? *Expert Opin. Drug Deliv.* 23, 7–11.
152. Li, X., Huang, J., Zhou, Y.-M., Chen, X., Dong, Q.-W., Cui, Y.-L., and Chen, Y.-B. (2026). ROS-responsive nanogels for brain targeted delivery of icaritin in the treatment of Parkinson's disease. *Int. J. Pharm.* 687, 126361. <https://doi.org/10.1016/j.ijpharm.2025.126361>.
153. Ismail, M., Wang, Y., Li, Y., Liu, J., Zheng, M., and Zou, Y. (2024). Stimuli-responsive polymeric nanocarriers accelerate on-demand drug release to combat glioblastoma. *Biomacromolecules* 25, 6250–6282.
154. Birhan, Y.S., and Tsai, H.-C. (2021). Recent developments in selenium-containing polymeric micelles: prospective stimuli, drug-release behaviors, and intrinsic anticancer activity. *J. Mater. Chem. B* 9, 6770–6801.
155. Gao, J., Zhai, Y., Lu, W., Jiang, X., Zhou, J., Wu, L., Du, L., Ou, C., Zhang, X., He, H., et al. (2024). ROS-sensitive PD-L1 siRNA cationic selenide nanogels for self-inhibition of autophagy and prevention of immune escape. *Bioact. Mater.* 41, 597–610. <https://doi.org/10.1016/j.bioactmat.2024.08.013>.
156. Lu, J., Luo, J., Li, J., Fu, S., Ran, Y., Li, J., Zhao, Y., and Hao, Y. (2023). Fluorescent Pirfenidone-Cerium(III) nanocomplexes protect against radiation-induced pulmonary fibrosis and inhibit tumor cell growth. *J. Drug Deliv. Sci. Technol.* 86, 104651. <https://doi.org/10.1016/j.jddst.2023.104651>.
157. Machhi, J., Yeapuri, P., Markovic, M., Patel, M., Yan, W., Lu, Y., Cohen, J.D., Hasan, M., Abdelmoaty, M.M., Zhou, Y., et al. (2022). Europium-Doped Cerium Oxide Nanoparticles for Microglial Amyloid Beta Clearance and Homeostasis. *ACS Chem. Neurosci.* 13, 1232–1244. <https://doi.org/10.1021/acschemneuro.1c00847>.
158. Dai, Y., Gao, S., Zhang, Z., Du, W.-J., Bao, X., and Xu, Y. (2025). CART and Ceria nanozyme co-loaded ROS-responsive nanogel for synergistic treatment of Alzheimer's disease. *Chem. Eng. J.* 522, 168038. <https://doi.org/10.1016/j.cej.2025.168038>.
159. Wang, C., Song, X., Li, P., Sun, S., Su, J., Liu, S., and Wei, W. (2024). Multifunctional Nanocarrier for Synergistic Treatment of Alzheimer's Disease by Inhibiting β -Amyloid Aggregation and Scavenging Reactive Oxygen Species. *ACS Appl. Mater. Interfaces* 16, 27127–27138. <https://doi.org/10.1021/acsami.4c02825>.
160. Huang, Q., Liao, J., Li, J., Gu, Z., Zhang, X., Sun, M., Meng, W., Mao, G., Pei, Z., Zhang, S., et al. (2025). Curcumin-loaded ceria nanoenzymes for dual-action suppression of inflammation and alleviation of oxidative damage in the treatment of acute lung injury. *Chin. Chem. Lett.* 36, 109914. <https://doi.org/10.1016/j.ccl.2024.109914>.
161. Sharif, N., Murtaza, I., Shahnaz, G., Saeed, A., Ishtiaq, A., Tabassum, S., Rahdar, A., Fathi-karkan, S., and Pandey, S. (2025). Precision nanophytomedicine: Alginate-chitosan nanogels encapsulating Olea ferruginea ethyl acetate fraction for enhanced cardiovascular protection. *Int. J. Biol. Macromol.* 318, 144656. <https://doi.org/10.1016/j.ijbiomac.2025.144656>.
162. Nunziata, G., Borroni, A., and Rossi, F. (2025). Advanced microfluidic strategies for core-shell nanoparticles: the next-generation of polymeric and lipid-based drug nanocarriers. *Chem. Eng. J. Adv.* 22, 100759. <https://doi.org/10.1016/j.cej.2025.100759>.
163. Wang, D., Li, D., Liu, X., Wang, S., Fan, Y., Lu, L., Shen, C., and Li, C. (2025). Reactive oxygen species-responsive sulfated polysaccharide-based nanogels for ischemic stroke: A precision therapy through pathological microenvironment modulation. *Acta Biomater.* 206, 403–419. <https://doi.org/10.1016/j.actbio.2025.09.031>.
164. Yeo, J., Lee, J., Yoon, S., and Kim, W.J. (2020). Tannic acid-based nanogel as an efficient anti-inflammatory agent. *Biomater. Sci.* 8, 1148–1159. <https://doi.org/10.1039/C9BM01384A>.
165. Xiang, Y., Qiu, Z., Ding, Y., Du, M., Gao, N., Cao, H., Zuo, H., Cheng, H., Gao, X., Zheng, S., et al. (2024). Dexamethasone-loaded ROS stimuli-responsive nanogels for topical ocular therapy of corneal neovascularization. *J. Contr. Release* 372, 874–884. <https://doi.org/10.1016/j.jconrel.2024.07.012>.
166. Cai, J., Zhong, H., Luo, J., Huang, X., Xu, Q., and Li, P. (2024). Inhalable multi-stimulus sensitive curcumin-alginate nanogels for scavenging reactive oxygen species and anti-inflammatory co-ordination to alleviate acute lung injury. *Int. J. Biol. Macromol.* 283, 137816. <https://doi.org/10.1016/j.ijbiomac.2024.137816>.
167. Zhang, J., Xu, Y., Li, M., Cai, W., He, G., Zhang, J., Li, M., Li, J., Yu, H., Wang, J., et al. (2025). Construction of ROS responsive photo-crosslinking nanogels based on 4-azidobenzoic acid-modified carboxymethyl chitosan for enhanced photostability and antimicrobial effect of toluidine blue O. *Mater. Today Commun.* 46, 112510. <https://doi.org/10.1016/j.mtcomm.2025.112510>.
168. Altinbasak, I., Alp, Y., Sanyal, R., and Sanyal, A. (2024). Theranostic nanogels: multifunctional agents for simultaneous therapeutic delivery and diagnostic imaging. *Nanoscale* 16, 14033–14056.
169. Arkaban, H., Barani, M., Akbarzadeh, M.R., Pal Singh Chauhan, N., Jadoun, S., Dehghani Soltani, M., and Zarrintaj, P. (2022). Polyacrylic Acid Nanoplateforms: Antimicrobial, Tissue Engineering, and Cancer Theranostic Applications. *Polymers* 14, 1259. <https://doi.org/10.3390/polym14061259>.
170. Mansur, A.A.P., Carvalho, S.M., Lobato, Z.I.P., Leite, M.F., Krambrock, K., and Mansur, H.S. (2025). Bioengineering stimuli-responsive organic-inorganic nanoarchitectures based on carboxymethylcellulose-poly-L-lysine nanoplexes: Unlocking the potential for bioimaging and multimodal chemodynamic-magnetothermal therapy of brain cancer cells. *Int. J. Biol. Macromol.* 290, 138985. <https://doi.org/10.1016/j.ijbiomac.2024.138985>.
171. Brandi, C., De Nino, A., Ruggiero, F., Limiti, E., Abbruzzese, F., Trombetta, M., Rainer, A., Bisegna, P., and Caselli, F. (2024). On the compatibility of single-cell microcarriers (nanovials) with microfluidic impedance cytometry. *Lab Chip* 24, 2883–2892. <https://doi.org/10.1039/D4LC00002A>.
172. Zhao, J., Yang, Y., Han, X., Liang, C., Liu, J., Song, X., Ge, Z., and Liu, Z. (2017). Redox-Sensitive Nanoscale Coordination Polymers for Drug Delivery and Cancer Theranostics. *ACS Appl. Mater. Interfaces* 9, 23555–23563. <https://doi.org/10.1021/acsami.7b07535>.
173. He, S., Zhang, Z., Li, H., Jiang, S., Han, G., and Shao, B. (2026). The biphasic interactions between ferroptosis and oxidative stress: from molecular mechanisms to disease interventions. *Mol. Biol. Rep.* 53, 903.

174. Srivastava, S., Saha, S., and Jakhmola, V. (2023). Nanogel: Types, Methods of Preparation, Limitation, Evaluation and Application—A Systematic Review. *Int. J. Drug Deliv. Technol* **13**, 1631–1639.
175. Pinelli, F., Saadati, M., Zare, E.N., Makvandi, P., Masi, M., Sacchetti, A., and Rossi, F. (2023). A perspective on the applications of functionalized nanogels: promises and challenges. *Int. Mater. Rev.* **68**, 1–25.
176. Delgado-Pujol, E.J., Martínez, G., Casado-Jurado, D., Vázquez, J., León-Barberena, J., Rodríguez-Lucena, D., Torres, Y., Alcudia, A., and Begines, B. (2025). Hydrogels and Nanogels: Pioneering the Future of Advanced Drug Delivery Systems. *Pharmaceutics* **17**, 215.
177. Nunziata, G., Pollonio, D., Lacroce, E., and Rossi, F. (2025). Smart pH-Responsive polymers in biomedical Applications: Nanoparticles, hydrogels, and emerging hybrid platforms. *Mater. Today Chem.* **49**, 103063.
178. Aswathy, S.H., NarendraKumar, U., and Manjubala, I. (2023). The influence of molecular weight of cellulose on the properties of carboxylic acid crosslinked cellulose hydrogels for biomedical and environmental applications. *Int. J. Biol. Macromol.* **239**, 124282. <https://doi.org/10.1016/j.ijbiomac.2023.124282>.
179. Li, H., Xiang, D., Gong, C., Wang, X., and Liu, L. (2022). Naturally derived injectable hydrogels with ROS-scavenging property to protect transplanted stem cell bioactivity for osteoarthritic cartilage repair. *Front. Bioeng. Biotechnol.* **10**, 1109074. <https://doi.org/10.3389/fbioe.2022.1109074>.
180. Rouco, H., Permuy, M., Muñoz, F., Vázquez, J.A., Caeiro, J.R., Landin, M., and Diaz-Rodriguez, P. (2025). Micelle-to-Gel: Thermosensitive intra-articular hydrogels for osteoarthritis management. *J. Contr. Release* **381**, 113639. <https://doi.org/10.1016/j.jconrel.2025.113639>.
181. Zewail, M., Abbas, H., El Sayed, N., El Newehy, N.M., and Abd-El-Azim, H. (2025). Intradermal delivery of nanohybrid thermosensitive hydrogels loaded with chondroitin sulphate coated pomegranate lipid carriers via hollow microneedles for effective rheumatoid arthritis management. *J. Drug Deliv. Sci. Technol.* **112**, 107234. <https://doi.org/10.1016/j.jddst.2025.107234>.
182. He, H., Zhang, Q., Zhang, Y., Qu, S., Li, B., Lin, J., Lu, X., and Xie, C. (2025). Injectable bioadhesive and lubricating hydrogel with polyphenol mediated single atom nanozyme for rheumatoid arthritis therapy. *Nat. Commun.* **16**, 2768. <https://doi.org/10.1038/s41467-025-58059-z>.
183. Sabarees, G., Jebaraj, Y.S., Ezhilarasan, E., and Ragul, Y.D. (2025). Next-Generation Injectable Hydrogels: Advanced Crosslinking Strategies, Multi-Stimuli Responsiveness, and Translational Advances for Precision Regenerative Medicine. *Nano TransMed*, 100109. <https://doi.org/10.1016/j.ntm.2025.100109>.
184. Zhou, K., He, X., Shi, K., Hu, D., Yang, C., Gao, M., Wang, Y., Liu, Y., Yang, Q., Chu, B., et al. (2026). A dual responsive nanohydrogel system for sustained drug delivery and cartilage penetration in osteoarthritis therapy. *J. Contr. Release* **389**, 114494. <https://doi.org/10.1016/j.jconrel.2025.114494>.
185. Zhao, H., Huang, J., Li, Y., Lv, X., Zhou, H., Wang, H., Xu, Y., Wang, C., Wang, J., and Liu, Z. (2020). ROS-scavenging hydrogel to promote healing of bacteria infected diabetic wounds. *Biomaterials* **258**, 120286. <https://doi.org/10.1016/j.biomaterials.2020.120286>.
186. Yang, C., Chen, Y., Huang, H., Fan, S., Yang, C., Wang, L., Li, W., Niu, W., and Liao, J. (2021). ROS-Eliminating Carboxymethyl Chitosan Hydrogel to Enhance Burn Wound-Healing Efficacy. *Front. Pharmacol.* **12**, 679580. <https://doi.org/10.3389/fphar.2021.679580>.
187. Zhu, J., Li, F., Wang, X., Yu, J., and Wu, D. (2018). Hyaluronic Acid and Polyethylene Glycol Hybrid Hydrogel Encapsulating Nanogel with Hemostasis and Sustainable Antibacterial Property for Wound Healing. *ACS Appl. Mater. Interfaces* **10**, 13304–13316. <https://doi.org/10.1021/acsami.7b18927>.
188. Lei, H., Yu, X., and Fan, D. (2024). Nanocomposite Hydrogel for Real-Time Wound Status Monitoring and Comprehensive Treatment. *Adv. Sci.* **11**, 2405924. <https://doi.org/10.1002/advs.202405924>.
189. Mao, Y., Zhao, B., Xu, L., Wang, Y., Qiu, X., Sun, Y., and Yang, C. (2025). Protein-polysaccharide based nanogel/hydrogel composite with controlled continuous delivery of drug for enhanced wound healing. *Carbohydr. Polym.* **356**, 123407. <https://doi.org/10.1016/j.carbpol.2025.123407>.
190. Benedetto Mas, A., Barberi, J., Montalbano, G., Fraterrigo Garofalo, S., Marta, E., Silva, E.D., Gomes, R.N., Nascimento, D.S., Sirolli, S., Cafarelli, A., et al. (2025). Piezoelectric electrospun scaffold incorporating ibuprofen loaded ultrasound-responsive mesoporous silica nanoparticles for tissue regeneration. *RSC Adv.* **15**, 33868–33883. <https://doi.org/10.1039/d5ra05217c>.
191. Zhang, M., Xiao, X., Li, X., Shi, X., and Li, Z. (2025). pH-sensitive fibronectin nanogels combined with UTMD for anti-atherosclerosis treatment through anti-inflammatory and antioxidant effects. *Mater. Today Bio* **33**, 102044. <https://doi.org/10.1016/j.mtbio.2025.102044>.
192. Zhang, S., Li, Y., Qiu, X., Jiao, A., Luo, W., Lin, X., Zhang, X., Zhang, Z., Hong, J., Cai, P., et al. (2021). Incorporating redox-sensitive nanogels into bioabsorbable nanofibrous membrane to acquire ROS-balance capacity for skin regeneration. *Bioact. Mater.* **6**, 3461–3472. <https://doi.org/10.1016/j.bioactmat.2021.03.009>.
193. Wang, L., Zhu, B., Deng, Y., Li, T., Tian, Q., Yuan, Z., Ma, L., Cheng, C., Guo, Q., and Qiu, L. (2021). Biocatalytic and antioxidant nanostructures for ROS scavenging and biotherapeutics. *Adv. Funct. Mater.* **31**, 2101804.
194. Deng, X., Zhang, Z., Ren, T., and Chen, L. (2025). Regulation of oxidative stress and inflammation caused by drug accumulation in the TME based on EPR-passive strategy and active targeting. *Cancer Nanotechnol.* **16**, 40.
195. Chu, Y., Luo, E., Wei, Y., Zhu, S., Wang, X., Yang, L., Gao, N., Wang, Y., Jiang, Z., Liu, C., et al. (2023). Dual single-atom catalyst design to build robust oxygen reduction electrode via free radical scavenging. *Chem Catal.* **3**, 100532.
196. Cho, E., Yun, S., Lee, S., Kim, M., Choi, J., Choi, S.E., Lim, K.S., Ha, S.-J., Yun, J.-H., and Kim, H.-O. (2025). Polymer-and Lipid-Based Nanostructures for Wound Healing with Barrier-Resolved Design. *Pharmaceutics* **17**, 1501.