

Activation of the Aromatic Core of 3,3'-(Pyridine-2,6-diylbis(1*H*-1,2,3-triazole-4,1-diyl))bis(propan-1-ol)—Effects on Extraction Performance, Stability Constants, and Basicity

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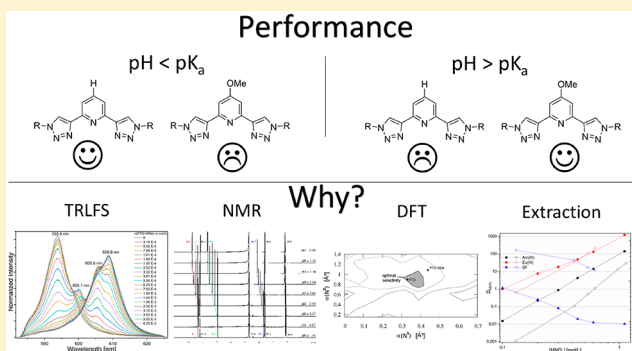
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Supporting Information

ABSTRACT: The “CHON” compatible water-soluble ligand 3,3'-(pyridine-2,6-diylbis(1*H*-1,2,3-triazole-4,1-diyl))bis-(propan-1-ol) (PTD) has shown promise for selectively stripping actinide ions from an organic phase containing both actinide and lanthanide ions, by preferential complexation of the former. Aiming at improving its complexation properties, PTD-OMe was synthesized, bearing a methoxy group on the central pyridine ring, thus increasing its basicity and hence complexation strength. Unfortunately, solvent extraction experiments in the range of 0.1–1 mol/L nitric acid proved PTD-OMe to be less efficient than PTD. This behavior is explained by its greater pK_a value ($pK_a = 2.54$) compared to PTD ($pK_a = 2.1$). This counteracts its improved complexation properties for Cm(III) ($\log \beta_3(\text{PTD-OMe}) = 10.8 \pm 0.4$ versus $\log \beta_3(\text{PTD}) = 9.9 \pm 0.5$).

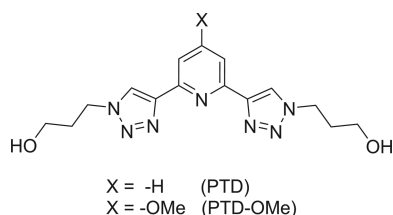


INTRODUCTION

Hydrophilic N-donor complexing agents are used in several solvent extraction processes developed to separate actinides from lanthanides.^{1–3} Such separations may play a role in future advanced nuclear fuel cycles. On the basis of the chemistry of the TALSPEAK process,^{4,5} actinide and lanthanide ions are co-extracted, followed by selective stripping of actinides using N-donor complexing agents such as aminopolycarboxylates or sulfonated N-heterocyclic compounds.^{6–13}

For this purpose, 3,3'-(pyridine-2,6-diylbis(1*H*-1,2,3-triazole-4,1-diyl))bis(propan-1-ol) (PTD, Scheme 1)¹⁴ is currently studied in the European research program GENIORS.¹⁵

Scheme 1. Molecular Structure of PTD and PTD-OMe



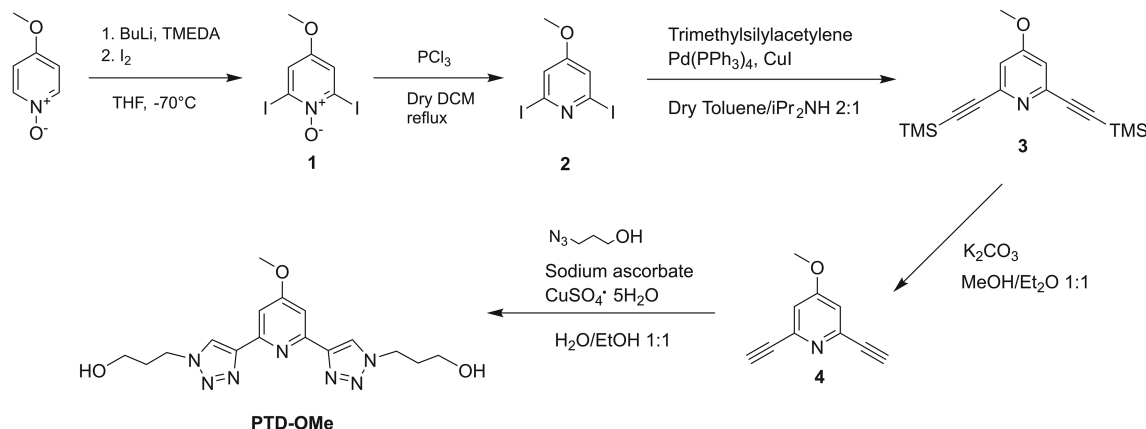
In contrast to aminopolycarboxylates, PTD remains efficient in up to 0.5 mol/L nitric acid, which is advantageous with respect to process applications. Furthermore, PTD (other than sulfonated N-heterocyclic compounds) is a “CHON” compound (i.e., containing only carbon, hydrogen, oxygen, and nitrogen, making it fully combustible to gaseous products without generating solid wastes).¹⁶ The favorable properties of PTD such as pronounced selectivity for actinides(III) over lanthanides(III), fast kinetics, and good stability^{14,17} make it a promising candidate for process development.¹⁸

To further improve its complexation properties, a modified PTD was synthesized, PTD-OMe (Scheme 1). Adding a methoxy group in the para position of the pyridine ring to activate the aromatic core, metal ion complexation should be improved due to the higher electron density in the aromatic ring. Such modifications had successfully been employed to tune the properties of lipophilic 2,6-bis-triazinyl-pyridine (BTP)¹⁹ and 2,9-bis-triazinyl-1,10-phenanthroline (BTPPhen)^{20,21} compounds.

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Scheme 2. Synthesis of PTD-OMe



PTD-OMe was studied using solvent extraction, time-resolved laser fluorescence spectroscopy (TRLFS), and NMR. The pK_a value, stability constants for the complexation of Cm(III), and the Cm(III) speciation in solution were determined. Computational calculations based on density functional theory (DFT) of actinide and lanthanide PTD-OMe complexes were performed to support experimental findings.

EXPERIMENTAL SECTION

Chemicals. PTD-OMe was synthesized as described below (Scheme 2). D₂O was purchased from Deutero GmbH. All commercially available chemicals (Sigma-Aldrich) used in this study were analytical reagent grade and used without further purification.

Synthesis. Melting points were determined on an Electrothermal apparatus in capillaries sealed under nitrogen. ¹H and ¹³C NMR spectra were recorded on Bruker AV300 and AV400 spectrometers. *J* coupling constants are given in hertz. Partially deuterated solvents were used as internal standards. Electrospray ionization mass spectrometry (ESI-MS) spectra were recorded on a Waters single quadrupole instrument SQ Detector, while high-resolution mass spectrometry (HR-MS) spectra were obtained with a Thermo Scientific Orbitrap LTQ-XL. Thin-layer chromatography (TLC) was performed on Merck 60 F254 silica gel, and flash column chromatography was performed on 230–400 mesh Merck 60 silica gel.

Synthesis of 2,6-Diiodo-4-methoxypyridine 1-oxide (1). Prepared according to literature procedure¹⁴ from 4-methoxypyridine 1-oxide in 30% yield. ¹H NMR (400 MHz, CD₃OD): δ 7.75 (2H, s, PyH3), 3.90 (3H, s, Py-OCH₃).

Synthesis of 2,6-Diiodo-4-methoxypyridine (2). Prepared according to literature procedure¹⁴ from 4-methoxy-pyridine 1-oxide in 90% yield. ¹H NMR (300 MHz, CD₃OD): δ 7.42 (2H, s, PyH3), 3.86 (3H, s, Py-OCH₃).

Synthesis of 4-Methoxy-2,6-bis(trimethylsilyl)ethynyl pyridine (3). 2,6-Diiodo-4-methoxypyridine (1.30 g, 3.60 mmol) is dissolved in a dry mixture of toluene and diisopropylamine 2:1 (150 mL) under inert conditions. Then CuI (0.03 g, 0.15 mmol), Pd(PPh₃)₄ (0.08 g, 0.07 mmol), and trimethylsilylacetylene (0.82 g, 8.32 mmol) are added, and the reaction mixture is stirred at room temperature. After 24 h the reaction is quenched with water, and the aqueous phase is extracted with ethyl acetate (AcOEt). The organic phases collected are dried over anhydrous Na₂SO₄, and the solvents are evaporated under reduced pressure. The crude is purified by flash column chromatography using hexane/AcOEt 8.5:1.5 as eluent. Yield: 92% ¹H NMR (400 MHz, CDCl₃): δ 6.95 (2H, s, PyH3), 3.87 (3H, s, Py-OCH₃), 0.27 (18H, s, Si(CH₃)₃); ¹³C NMR (100 MHz, CDCl₃): δ 165.7, 144.4, 113.2, 103.2, 94.8, 55.5, 0.3. mp 64–66 °C.

Synthesis of 2,6-Diethynyl-4-methoxypyridine (4). One gram (3.32 mmol) of 4-methoxy-2,6-bis(trimethylsilyl)ethynyl pyridine is dissolved in 70 mL of a mixture of MeOH/Et₂O 2:1, and then 2.3 g (16.6 mmol) of K₂CO₃ is added. The reaction mixtures are stirred for 1 h and then quenched with water. The aqueous layer is extracted three times with AcOEt. The organic phases collected are dried over anhydrous Na₂SO₄, and then the solvent is removed under reduced pressure. The product is obtained as a brownish solid in 88% yield. ¹H NMR (400 MHz, CDCl₃): δ 7.01 (2H, s, PyH3), 3.88 (3H, s, Py-OCH₃), 3.13 (2H, s, CCH); ¹³C NMR (100 MHz, CDCl₃): δ 165.8, 143.7, 113.7, 82.1, 77.3, 55.6; HR-MS (ESI+) *m/z*: [M + H]⁺ Calcd for C₁₀H₈NO 158.0600; Found 158.0601; mp 140 °C dec.

Synthesis of 3-Azidopropan-1-ol. Prepared according to literature procedure.²² The 9.1 g (140 mmol) of NaN₃ are added to 50 mL of water. When the azide is completely dissolved, 6.87 g (68 mmol) of 3-chloropropan-1-ol are added dropwise. The reaction mixture is stirred at 80 °C for 24 h. The aqueous solution is extracted three times with dichloromethane, and then the organic phases collected are dried over anhydrous Na₂SO₄. The solvent is evaporated in vacuo to obtain the product as a yellowish liquid. Yield: 85% ¹H NMR (400 MHz, CDCl₃): δ 3.76 (2H, t, *J* = 6.0 Hz, CH₂N₃), 3.46 (2H, t, *J* = 6.6 Hz, CH₂OH), 1.84 (2H, quint, *J* = 6.3 Hz, CH₂CH₂CH₂). HR-MS (ESI+) *m/z*: [M + H]⁺ Calcd for C₁₆H₂₄NOSi₂ 302.1391; Found 302.1392.

Synthesis of PTD-OMe. The 0.25 g (1.59 mmol) of 2,6-diethynyl-4-methoxypyridine and 0.52 g (5.2 mmol) of 3-azidopropan-1-ol are dissolved in 30 mL of a 1:1 mixture of water and ethanol. Then 7.5 mg (0.03 mmol) of CuSO₄·5H₂O and 63.4 mg (0.32 mmol) of sodium ascorbate are added. The reaction mixture is stirred for 3 d and then quenched by removal of the solvents under reduced pressure. The crude is purified by flash column chromatography using dichloromethane/methanol 92/8 as eluent. The product was obtained as a white solid in 57% yield. ¹H NMR (400 MHz, CD₃OD): δ 8.57 (2H, s, Triaz-H), 7.53 (2H, s, PyH3), 4.62 (4H, t, *J* = 7.2 Hz, CH₂N), 4.00 (3H, s, Py-OCH₃), 3.64 (4H, t, *J* = 6.0 Hz, CH₂OH), 2.20 (4H, quint, *J* = 6.4 Hz, CH₂CH₂N). ¹³C NMR (100 MHz, CD₃OD): δ 167.6, 151.3, 147.6, 123.6, 104.7, 57.9, 54.8, 32.6. ESI-MS (+): 382.3 [M + Na]⁺, 398.2 [M + K]⁺, 558.9 [M + Na ascorbate]⁺, 741.5 [2M + Na]⁺. HR-MS (ESI+) *m/z*: [M + H]⁺ Calcd for C₁₆H₂₂N₇O₃ 360.1779; Found 360.1786. mp: 118–120 °C.

Solvent Extraction. Organic phase was 0.2 mol/L N,N,N',N'-tetra-*n*-octyl-3-oxapentanediamide (TODGA)^{23–27} + 5 vol % 1-octanol in kerosene. Aqueous phase was 80 mmol/L PTD or PTD-OMe in HNO₃ (varied concentration) spiked with each 2.5 kBq/mL ²⁴¹Am(III) and ¹⁵⁴Eu(III).

Each 300 μ L of aqueous and organic phases were placed in 2 mL Eppendorf tubes and shaken on an orbital shaker for 60 min at 1100 rpm and 295 K. Following centrifugation for 10 min at 1000 rpm, 200 μ L aliquots of both phases were analyzed on a gamma counter (Packard Cobra Auto-Gamma 5003).

TRLFS Sample Preparation. Stock solutions containing 0.5 mol/L PTD-OMe were prepared by dissolving 50.3 mg of PTD-OMe in 280 μL of 1×10^{-3} mol/L HClO_4 or 0.44 mol/L HNO_3 . Solutions with lower PTD-OMe concentrations were prepared by dilution with 1×10^{-3} mol/L HClO_4 or 0.44 mol/L HNO_3 , respectively.

TRLFS samples were prepared by adding 4.7 μL of a Cm(III) stock solution (2.12×10^{-5} mol/L $\text{Cm}(\text{ClO}_4)_3$ in 0.1 mol/L HClO_4 ; ^{248}Cm : 89.7%, ^{246}Cm : 9.4%, ^{243}Cm : 0.4%, ^{244}Cm : 0.3%, ^{245}Cm : 0.1%, ^{247}Cm : 0.1%) to 995.3 μL of 1×10^{-3} mol/L HClO_4 or 0.44 mol/L HNO_3 , resulting in an initial Cm(III) concentration of 1×10^{-7} mol/L. Ligand concentration was adjusted by adding appropriate volumes of the PTD-OMe solutions. TRLFS spectra were recorded following an equilibration time of 10 min. Preliminary tests showed this to be sufficient to attain equilibrium.

Solvent extraction samples for TRLFS measurements were prepared as described in Solvent Extraction, with the exception that samples were 500 μL per phase each spiked with 4.7 μL of the Cm(III) stock solution instead of ^{241}Am and ^{154}Eu .

TRLFS Measurements. TRLFS measurements were performed at 298 K using a Nd:YAG (Surelite II laser, Continuum) pumped dye laser system (NarrowScan D-R; Radiant Dyes Laser Accessories GmbH). A wavelength of 396.6 nm was chosen to excite Cm(III). A spectrograph (Shamrock 303i, ANDOR) with 300, 1199, and 2400 lines per millimeter gratings was used for spectral decomposition. The fluorescence emission was detected by an ICCD camera (iStar Gen III, ANDOR) after a delay time of 1 μs to discriminate short-lived, organic fluorescence, and light scattering.

NMR Sample Preparation. NMR samples for pK_a determination contained initially 9×10^{-3} mol/L PTD-OMe in an aqueous formic acid/formate buffer containing 10 vol % of D_2O . pH was measured with a microelectrode (Orion PerpHecT ROSS, Thermo Fisher Scientific) and a pH meter (Orion Star, Thermo Fisher Scientific) before and after NMR measurement. The pH was adjusted with 1, 0.1, or 0.01 mol/L HCl or NaOH solutions.

NMR Measurements. NMR spectra were recorded at $T = 300$ K on a Bruker Avance III 400 spectrometer operating at a resonance frequency of 400.18 MHz for ^1H nuclei. The spectrometer was equipped with a z-gradient observe room-temperature probe. Chemical shifts were referenced internally to tetramethylsilane (TMS) ($\delta(\text{TMS}) = 0$ ppm) by the deuterium lock signal of D_2O . For single scan ^1H spectra, standard 90° pulse sequences were used. Water suppression was achieved by the WATERGATE^{28,29} pulse sequence. All spectra were recorded with 32k data points and were zero filled to 64k points. For WATERGATE spectra, eight scans were acquired per spectrum with a relaxation delay of 2 s. Exponential window functions with a line broadening factor of 0.05 Hz were applied for processing.

Theoretical Model. Protonation and the complex formation of PTD-OMe were investigated. Structures were optimized employing density functional theory at the B3-LYP³⁰/def2-TZVP³¹ level of theory as implemented in the TURBOMOLE³² program package. To avoid the known problem of spin contamination in Gd(III) complexes computed by the B3-LYP functional, the BH-LYP³³ functional was used instead on all metal ion complexes. NMR shielding constants were calculated for all atoms of unprotonated and protonated PTD-OMe using the MPSHIFT³⁴ routine in TURBOMOLE. Binding energies corrected for basis set superposition error (BSSE) of $[\text{M}(\text{PTD-OMe})_3]^{3+}$ and $[\text{M}(\text{PTD})_3]^{3+}$ were determined on the MP2/def2-TZVP level, which was shown to yield good results when comparing to experimental separation factors (SF).³⁵ The Cm(III) ion was described by a ECP60MWB³⁶ small-core pseudopotential, and the Gd(III) was described by an ECP28MWB³⁶ small-core pseudopotential. Differences in selectivity have recently been connected to atomic polarizabilities of the coordinating atoms.³⁷ To investigate the ligands at hand within this approach, atomic charges and dipole polarizabilities were calculated on the B3LYP/aug-cc-pVTZ³⁸ level using the Hirshfeld method.^{37,39}

RESULTS AND DISCUSSION

Solvent Extraction. The solvent extraction behavior of PTD-OMe was studied using Am(III) and Eu(III) as representatives for trivalent actinides and lanthanides. Figure 1 compares distribution ratios ($D_{\text{M(III)}} = [\text{M(III)}]_{\text{org}}/[\text{M-}$

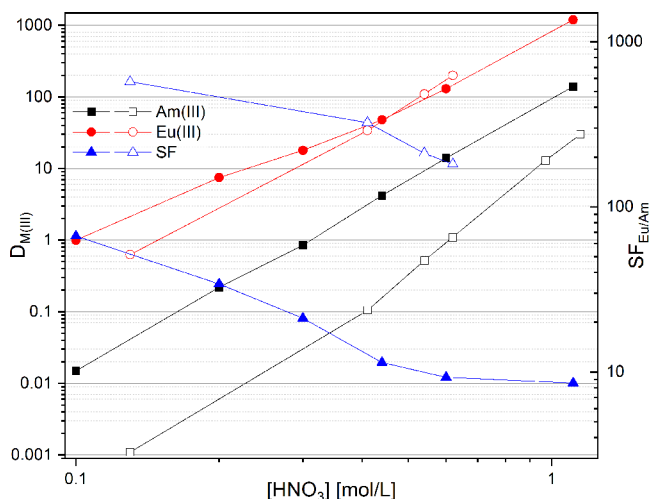


Figure 1. Distribution ratios for the extraction of Am(III) and Eu(III) with TODGA/PTD-OMe (solid symbols) and TODGA/PTD (open symbols). Organic phase, 0.2 mol/L TODGA in TPH/1-octanol (5 vol %). Aqueous phase, 0.08 mol/L PTD-OMe or PTD in nitric acid. A/O = 1, $T = 295$ K.

$(\text{III})]_{\text{aq}}$) and separation factors ($\text{SF} = D_{\text{Eu(III)}}/D_{\text{Am(III)}}$) for the systems TODGA/PTD and TODGA/PTD-OMe as a function of nitric acid concentration.

Am(III)/Eu(III) separation is achieved for $D_{\text{Am(III)}} < 1$ and $D_{\text{Eu(III)}} > 1$. With PTD, this is the case at 0.15–0.6 mol/L nitric acid, with a selectivity in the range of $500 > \text{SF} > 180$. With PTD-OMe, separation is achieved at 0.1–0.3 mol/L nitric acid, with a selectivity in the range of $60 > \text{SF} > 20$. Contrary to expectation, PTD-OMe is a less efficient ligand compared to PTD.

Determination of the pK_a Value by NMR. The insertion of an activating –OMe group on the pyridine nucleus of the ligand will impact the pK_a value. Therefore, the pK_a value of PTD-OMe was determined by evaluating the shifts of its ^1H NMR signals as a function of the measured pH. Note that the ionic strength is not constant during titration. Because of the rather low ionic strength (10^{-2} to 10^{-1} mol/L), however, no corrections for the pH were performed. Corresponding ^1H NMR spectra at different pH values are shown in Figure 2.

The ^1H NMR signals of PTD-OMe show a downfield shift with decreasing pH due to protonation of the ligand. The ^1H NMR signals shift in a pH range of 4.02–1.25. No shifts are observed beyond this range, indicating the existence of only the protonated or unprotonated ligand. Both species are characterized as follows (for proton assignment see Figure 3):

- Unprotonated PTD-OMe (pH = 4.39): ^1H NMR (400.18 MHz in formic acid/formate buffer +10 vol % D_2O): δ 8.20 (s, 2H, H-1); 6.93 (s, 2H, H-3); 3.65 (s, 3H, H-2); 3.58 (t, $^3J_{\text{H-H}} = 6.30$ Hz, 4H, H-6); 2.09 (quint, $^3J_{\text{H-H}} = 6.68$ Hz, 4H, H-5).
- Protonated PTD-OMe (pH = 0.96): ^1H NMR (400.18 MHz in formic acid/formate buffer +10 vol % D_2O): δ 8.75 (s, 2H, H-1); 7.65 (s, 2H, H-3); 4.12 (s, 3H, H-2);

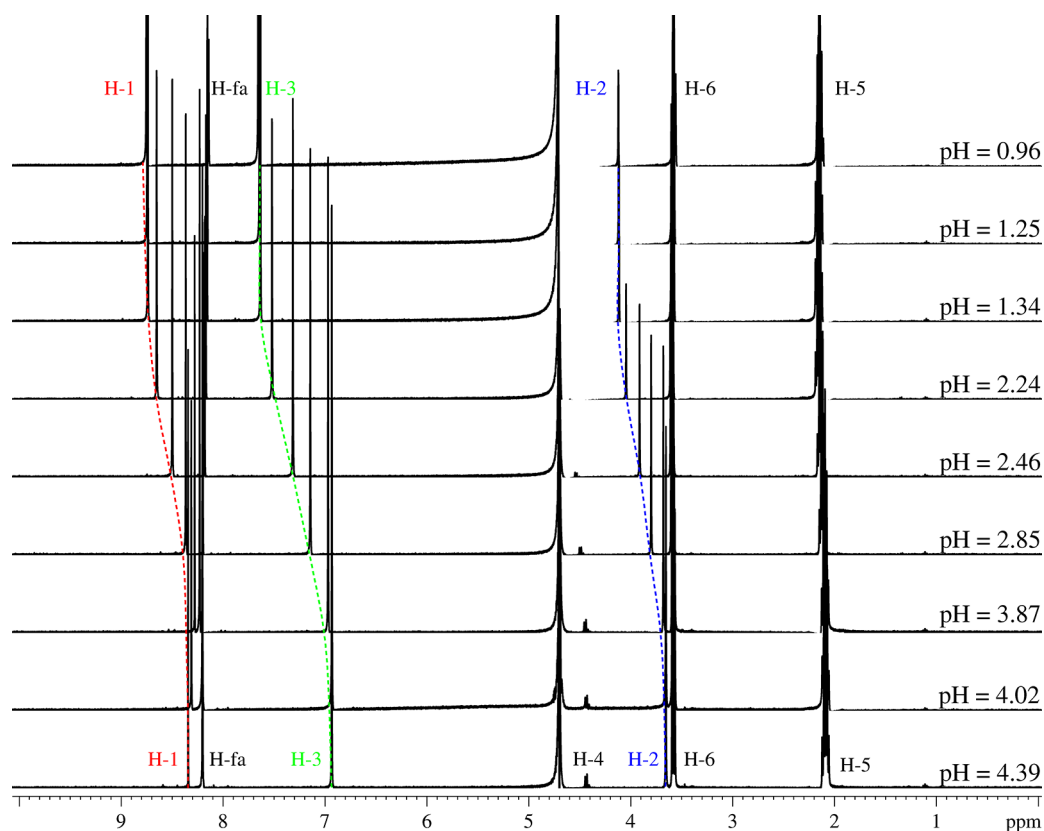


Figure 2. ^1H NMR spectra of PTD-OME in formic acid/formate (H-fa) buffer with 10 vol % D_2O at varied pH. The water signal at $\delta = 4.702$ ppm was suppressed using the WATERGATE technique.

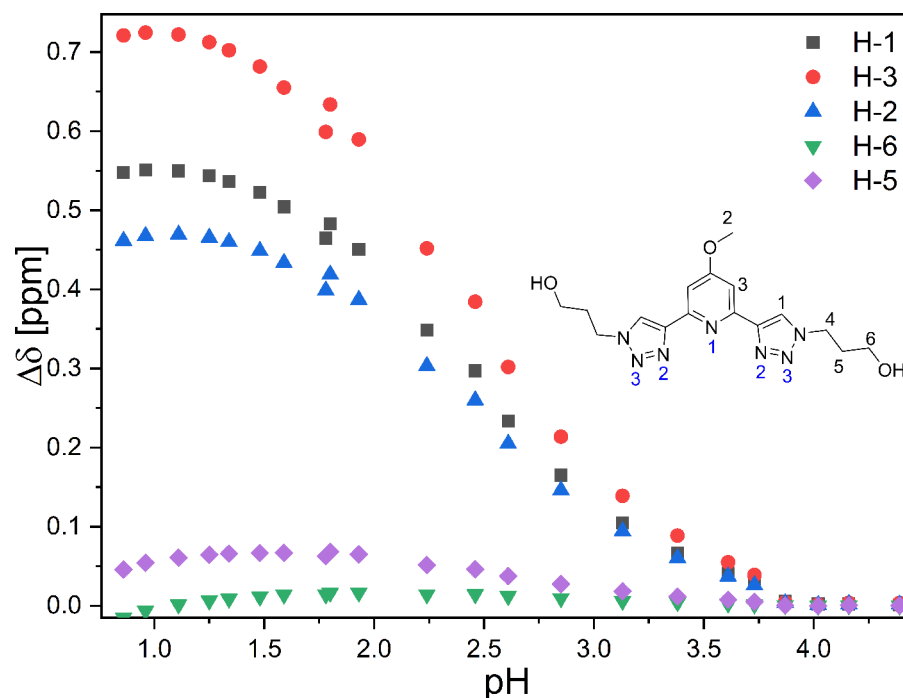


Figure 3. Shift of the ^1H signals in respect to the ^1H shift of the unprotonated PTD-OME as a function of pH.

3.57 (t, $^3J_{\text{H-H}} = 6.21$ Hz, 4H, H-6); 2.14 (quint, $^3J_{\text{H-H}} = 6.55$ Hz, 4H, H-5).

Spectra in the pH range of 1.25–3.87 contain both protonated and unprotonated species. The presence of only one set of ^1H signals indicates fast proton exchange, and,

therefore, only an average of the signals of the two species is detected. Consequently, the peak shift ($\Delta\delta$, eq 1) at a given pH relative to the shift of the unprotonated species ($\delta_0 = \delta_{\text{pH}=4.39}$) is used for pK_a determination. The relative peak shifts for all protons of PTD-OME are shown in Figure 3.

$$\Delta\delta_i = \delta_i - \delta_0 \quad (1)$$

With decreasing pH, aromatic protons shift more strongly downfield than protons farther away from the aromatic center. The strongest shifts are observed for the pyridine protons (H-3; $\Delta\delta_{\max} = 0.725$ ppm), followed by the triazole protons (H-1; $\Delta\delta_{\max} = 0.552$ ppm) and the methoxy protons (H-2; $\Delta\delta_{\max} = 0.470$ ppm). Shifts of the protons H-5 and H-6 of the propanol units are negligible. The shift of the proton H-4 cannot be evaluated due to the WATERGATE method. The strong shifts of the protons in the aromatic region indicate protonation in the aromatic region and most probably on the pyridine nucleus.

To determine the pK_a value, proton signals H1, H2, and H3 were evaluated individually. With eq 2, relative concentrations were calculated. The species distribution is given in the Supporting Information, Figure S1.

$$\chi_i = \frac{\Delta\delta_i}{\Delta\delta_{\max}} \quad (2)$$

The Henderson–Hasselbalch eq 3 was used to determine the pK_a value of PTD-OMe.

$$\log\left(\frac{[LH_n^{n+}]}{[L]}\right) = -n \times \text{pH} + pK_a \quad (3)$$

Slope analyses of H-1, H-2, and H-3 shifts as a function of pH are shown in the Supporting Information, Figure S2. Slopes (see Table 1) of -1 confirm that one proton is transferred. pK_a

Table 1. Slopes and pK_a Values for the Protonation of PTD-OMe

proton	slope	pK_a
H-1	-1.06 ± 0.03	2.55 ± 0.05
H-2	-1.06 ± 0.03	2.51 ± 0.03
H-3	-1.03 ± 0.03	2.57 ± 0.06

values derived from the shifts of H-1, H-2, and H-3 are reported in Table 1. The average pK_a of PTD-OMe has a value of 2.54 ± 0.08 . The attachment of a methoxy group, as expected for an electron-donating moiety, to the pyridine core results in a slightly increased basicity compared to PTD ($pK_a = 2.1$).¹⁴

Complexation of Cm(III) with PTD-OMe at pH = 3.

TRLFS was utilized to study the complexation of Cm(III) with PTD-OMe. Stability constants and the speciation in solution were determined. Because of its favorable spectroscopic properties, Cm(III) was used to represent Am(III). This is valid due to the similar chemical properties of both elements.

The evolution of the Cm(III) fluorescence spectra resulting from the ${}^6D_{7/2} \rightarrow {}^8S_{7/2}$ transition is shown in Figure 4 as a function of the PTD-OMe concentration.

Without addition of PTD-OMe the emission band of the Cm(III) aqua ion is observed at 593.8 nm.⁴⁰ With addition of PTD-OMe new emission bands evolve at 600.1, 605.6, and 608.8 nm. The bathochromic shift is explained by the increased splitting of the ${}^6D_{7/2}$ state due to the complexation of Cm(III) with PTD-OMe. The emission bands are in excellent agreement with those of the Cm(PTD)_n complexes ($n = 1-3$) in 1×10^{-3} mol/L HClO_4 ,¹⁷ indicating that the observed emission bands correspond to the Cm(PTD-OMe)_n complexes ($n = 1-3$).

With single-component spectra of the Cm(PTD-OMe)_n complexes obtained by peak deconvolution (Figure 5) and the respective fluorescence intensity factors¹³ ($FI_1 = 1$, $FI_2 = 1.1$, and $FI_3 = 1.1$), the species distribution of the $[\text{Cm(PTD-OMe)}_n]^{3+}$ complexes ($n = 1-3$) as a function of the free (i.e., unprotonated and uncomplexed) PTD-OMe concentration was derived (see Figure 6). The free ligand concentration was calculated using eq 4 with $[L]_0$ being the initial ligand concentration, $[H^+]_0$ the initial proton concentration, K_a the ligand protonation constant, and χ_i the relative fraction of the Cm(III) complex species present at a given ligand concentration.

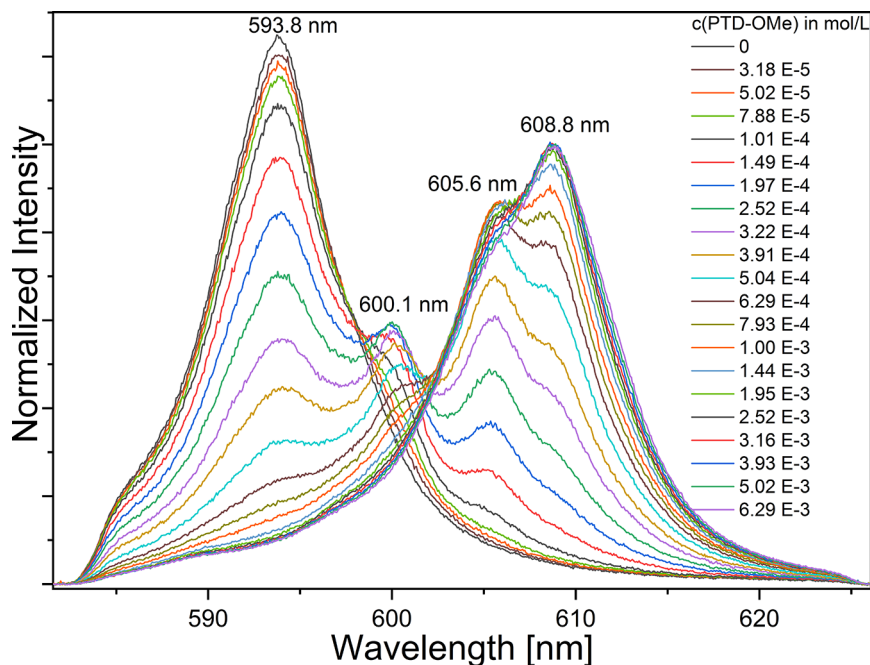


Figure 4. Normalized Cm(III) emission spectra in 1×10^{-3} mol/L HClO_4 with increasing PTD-OMe concentrations. $[\text{Cm}]_{\text{ini}} = 1 \times 10^{-7}$ mol/L.

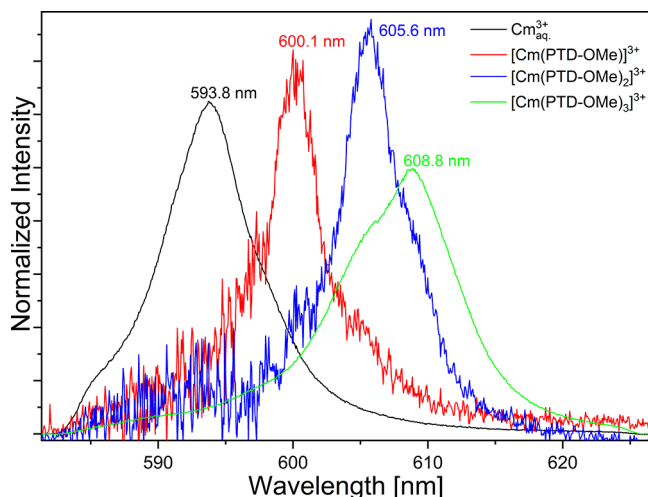
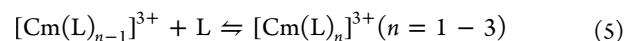


Figure 5. Normalized single-component spectra of the $[\text{Cm}(\text{PTD-OMe})_n]^{3+}$ ($n = 0-3$) complexes in 1×10^{-3} mol/L HClO_4 .

$$[\text{L}]_{\text{free}} = 0.5 \times ((4 \times [\text{L}]_0 \times K_a + [\text{H}^+]_0^2 + 2 \times [\text{H}^+]_0 \times K_a + K_a^2)^{0.5} - [\text{H}^+]_0 - K_a) - ([\text{Cm(III)}]) \times (\chi_{1:1} + 2 \times \chi_{1:2} + 3 \times \chi_{1:3}) \quad (4)$$

The 1:1 complex starts forming at a PTD-OMe concentration of $\approx 10^{-5}$ mol/L, with a maximum of 23% at 2.5×10^{-4} mol/L. The 1:2 complex has a maximum fraction of 26% at 5.3×10^{-4} mol/L PTD-OMe. For PTD-OMe concentrations greater than 3.5×10^{-4} mol/L the 1:3 complex is the dominating species. The fluorescence lifetime ($\tau = 495 \pm 20$ μs) determined at a ligand concentration of 6.29×10^{-3} mol/L (see Figure S3) indicates the absence of water molecules in the first coordination sphere ($n(\text{H}_2\text{O}) = 0.4 \pm 0.5$),⁴⁰ in agreement with full coordination by three PTD-OMe molecules.

To verify the stepwise complexation according to eq 5 ($\text{L} = \text{PTD-OMe}$)



slope analyses were performed using eq 6, with the results shown in the Supporting Information, Figure S4.

$$\log \left(\frac{[\text{Cm}(\text{L})_n]^{3+}}{[\text{Cm}(\text{L})_{n-1}]^{3+}} \right) = 1 \times \log([\text{L}]_{\text{free}}) + \log K'_n \quad (6)$$

Slopes of $m_1 = 1.13 \pm 0.06$, $m_2 = 0.98 \pm 0.15$, and $m_3 = 1.19 \pm 0.07$ confirm the stepwise complexation according to eq 5.

With eq 7, conditional stability constants were determined: $\log \beta'_1 = 3.4 \pm 0.3$, $\log \beta'_2 = 7.0 \pm 0.4$, and $\log \beta'_3 = 10.8 \pm 0.4$. Calculated and experimental relative fractions deviate slightly in the range from 1×10^{-4} to 4×10^{-4} mol/L. This is due to the lower signal-to-noise ratio of the single-component spectra of the 1:1 and 1:2 complexes used for peak deconvolution.

$$\log \beta'_n = \log \left(\frac{[\text{Cm}(\text{L})_n]^{3+}}{[\text{Cm}_{\text{aq}}]^{3+} \times [\text{L}]_{\text{free}}^n} \right) \quad (7)$$

TRLFS on Solvent Extraction Samples. To study the speciation under extraction conditions (0.44 mol/L HNO_3), TRLFS was performed on both the aqueous and organic phases of a solvent extraction experiment.

The organic phase (Figure 7, top) shows the emission spectrum of the $[\text{Cm}(\text{TODGA})_3]^{3+}$ complex¹⁷ with its characteristic emission band at 608.8 nm and a hot band at 595 nm. The species is also verified by its fluorescence lifetime (see Supporting Information, Figure S5; $\tau = 403 \pm 12$ μs ; $n(\text{H}_2\text{O}) = 0.7 \pm 0.5$), confirming coordination of three TODGA ligands.

In case of PTD, 80% of the 1:3 complex and 20% of the 1:2 complex are found under the extraction conditions (0.44 mol/L HNO_3).¹⁷ To observe a high selectivity in an extraction

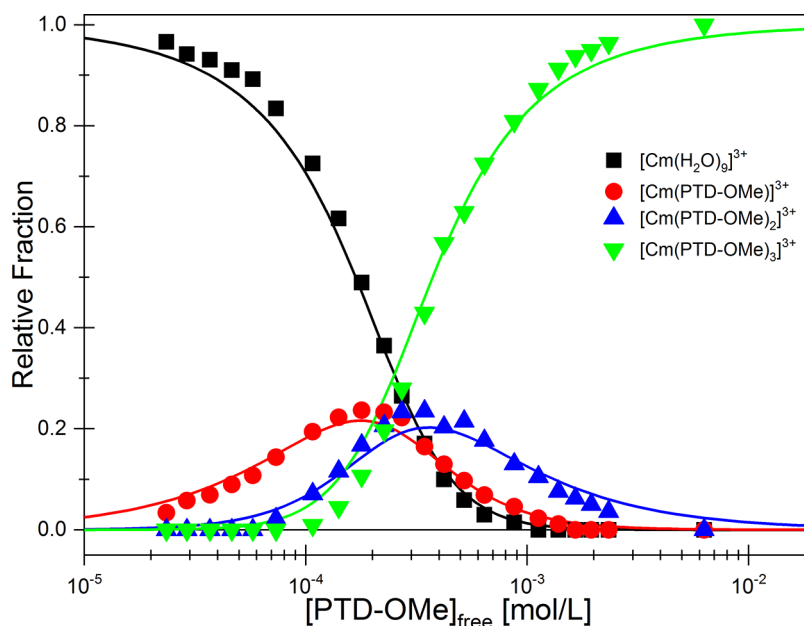


Figure 6. Cm(III) species distribution in 1×10^{-3} mol/L HClO_4 as a function of the free PTD-OMe concentration. Symbols, experimental data. Lines, calculated with $\log \beta'_1 = 3.4$, $\log \beta'_2 = 7.0$, and $\log \beta'_3 = 10.8$.

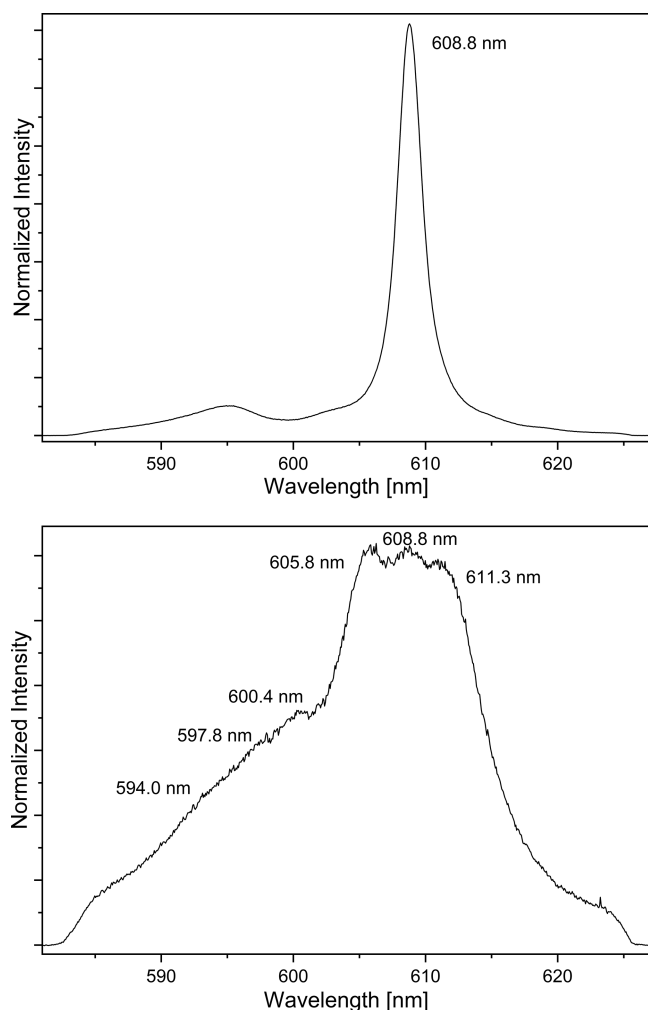


Figure 7. Cm(III) emission spectra of the organic (top) and aqueous (bottom) phases of a solvent extraction sample. Organic phase, 0.2 mol/L TODGA in TPH/1-octanol (5 vol %). Aqueous phase, 1×10^{-7} mol/L Cm(III) and 0.08 mol/L PTD-OMe in 0.44 mol/L nitric acid.

experiment, however, the presence of only the 1:3 complex is mandatory. To improve the speciation in 0.44 mol/L HNO_3 the methoxy group was introduced in PTD-OMe. Yet, multiple species are present in 0.44 mol/L HNO_3 in case of PTD-OMe as shown by the emission spectrum of the aqueous phase (Figure 7, bottom) explaining the observed diminished selectivity of PTD-OMe.

Comparison of PTD-OMe and PTD. An overview of the conditional stability constants and $\text{p}K_a$ values of PTD-OMe and PTD is given in Table 2. The $\text{p}K_a$ value of PTD-OMe is ≈ 0.5 higher than the $\text{p}K_a$ of PTD. This higher basicity is in

Table 2. Conditional Stability Constants of the $[\text{Cm}(\text{PTD-OMe})_n]^{3+}$ and $[\text{Cm}(\text{PTD})_n]^{3+}$ Complexes

PTD-OMe		PTD ¹⁷
$\text{p}K_a = 2.54$		$\text{p}K_a = 2.1$
n	$\log \beta'_n$	$\log \beta'_n$
1	3.4 ± 0.3	3.2 ± 0.2
2	7.0 ± 0.4	6.8 ± 0.2
3	10.8 ± 0.4	9.9 ± 0.5

good agreement with the higher conditional stability constants observed for PTD-OMe compared to PTD. Stability constants for the lower complex species are less influenced by the activation of the aromatic core, whereas the conditional stability constant of the 1:3 complex is almost 1 order of magnitude higher for PTD-OMe. A linear correlation between stability constants and $\text{p}K_a$ values has been observed for many other systems.^{41,42} The results for PTD-OMe are in good agreement with this observed trend.

The higher basicity of PTD-OMe explains its inferior performance in extraction experiments compared to PTD. The free ligand concentration decreases due to the acidic conditions used in the extraction experiments (0.44 mol/L HNO_3). Because of the higher $\text{p}K_a$ value of PTD-OMe, the concentration of unprotonated ligand is lower compared to that of PTD. Consequently, Am(III) distribution ratios are less favorable, that is, higher. Since N-donor ligands such as PTD(-OMe) have lower affinity for Eu(III) compared to Am(III), Eu(III) distribution ratios are less affected, and hence selectivity is lower for PTD-OMe (cf. Figure 1).

As expected, the activation of the aromatic core of PTD made PTD-OMe a stronger ligand. However, in systems in which the pH is smaller than the $\text{p}K_a$ value the ligand with the lower $\text{p}K_a$ shows the better extraction performance, as the free ligand concentration is higher.

Quantum-Chemical Calculations. To support the experimental findings, various computational calculations were performed using DFT. The effect of the protonation on NMR shielding was investigated. The NMR spectra (see Figure 3) reveal the strongest downfield shift for the pyridine protons (H-3), followed by the triazole (H-1) and methoxy (H-2) protons. For the latter, a static model representation is not able to capture the equivalency of the three methoxy protons. Hence, average values are reported in the following. To elucidate the site of protonation, various protonated structures of PTD-OMe were calculated and compared (Table 3).

Table 3. Energy Difference of the Different Optimized Protonated PTD-OMe Structures Relative to the Most Stable One^a

site of protonation	ΔE [kJ/mol]
H-N ¹	0
H-N ²	55.8
H-N ³	150.5
H-OMe	311.8
H-OH	311.2

^aCalculated at B3-LYP/def2-TZVP level of theory. For atom assignment, see Figure 3.

The most favorable protonation energy is found for the pyridine nitrogen (N¹). The protonation of the coordinating nitrogen atoms of the triazole rings (N²) is disfavored by 55.8 kJ/mol. The protonation of the other nitrogen atoms (N³) or the oxygen donors is disfavored even more. Similar trends are reported for BTP-type ligands, where protonation also takes place at the pyridine nitrogen, but the protonation of the coordinating triazine nitrogen atoms is disfavored by only 25.1 kJ/mol.^{19,43,44}

¹H NMR shifts were calculated to further support the site of protonation. $\Delta\delta_i$ values (cf. equation 1) between the unprotonated PTD-OMe and all protonated PTD-OMe

structures were calculated. Table 4 reports $\Delta\delta_i$ values for the protons of the pyridine (H-3) and the triazole (H-1) ring and the methoxy group (H-2).

Table 4. Differences of the ^1H -NMR Shifts between Different Protonated PTD-OMe and the Unprotonated PTD-OMe^a

	site of protonation				
	H–N ¹	H–N ²	H–N ³	H–OMe	H–OH
H-1 (Tri)	0.130	0.281	0.190	0.004	–0.021
H-2 (OMe)	0.326	0.157	0.166	1.106	–0.015
H-3 (Py)	0.267	0.176	0.172	0.120	–0.221

^a $\Delta\delta_i$ in analogy to equation 1, calculated with the MPSHIFT package at B3-LYP/def2-TZVP level of theory.

Downfield shifts observed experimentally decrease in the order of $\Delta\delta_{\text{H-3}} > \Delta\delta_{\text{H-1}} > \Delta\delta_{\text{H-2}}$. Note that, for the evaluation of the calculations, the shifts of H-2 are not considered, as the methoxy groups rotate very quickly in solution, which leads to larger deviations of the calculations. Therefore, the trend $\Delta\delta_{\text{H-3}} > \Delta\delta_{\text{H-1}}$ was evaluated. It was found only for the protonation at the pyridine nitrogen or the methoxy oxygen. Since the protonation of the methoxy oxygen is disfavored by 311.3 kJ/mol in all calculations, both protonation energies and ^1H shifts suggest the protonation of PTD-OMe to occur at the pyridine nitrogen.

Furthermore, the structures of the 1:3 complexes of Cm(III) and Gd(III) with PTD and PTD-OMe were optimized, and binding energies (BE) of the complexes and atomic charges (q) and dipole polarizabilities (α) of the coordinating nitrogen atoms (N¹ and N²) were calculated. It had been shown that dipole polarizabilities of ligands influence their selectivity.³⁷ The calculated values are listed in Table 5.

Introduction of the methoxy group does not affect the charge density (q) within the pyridine ring in PTD-OMe compared to PTD but has a significant effect on the dipole polarizabilities (α) of all nitrogen atoms, both in the pyridine as well as the triazole ring. This effect is also reflected in the calculated bond distances between the metal ions and N₁ in the $[\text{M}(\text{PTD-OMe})_3]^{3+}$ (M = Cm, Gd) complexes, in which the bond length decreases by 2 pm in both complexes compared to PTD (cf. Supporting Information, Table S1).

According to theory,³⁷ An(III)/Ln(III) selectivity with respect to the dipole polarizabilities of N¹ versus N² for both PTD and PTD-OMe is plotted in Figure 8 with the optimal zone for high selectivity highlighted.

The dipole polarizabilities of PTD are within the optimal zone for high selectivity. On the basis of the atomic properties, a significant decrease in selectivity is expected for PTD-OMe. Accordingly, lower separation factors $\text{SF}_{\text{Cm/Gd}}$ are expected to be obtained. To calculate $\text{SF}_{\text{Cm/Gd}}$ values, an exchange reaction according to equation 8 was considered.

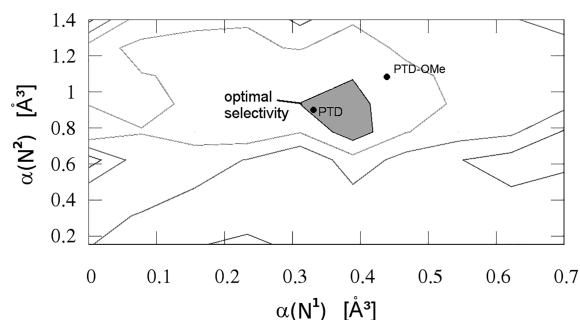
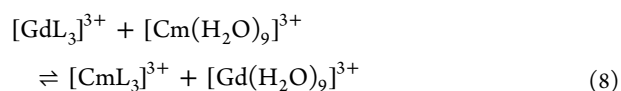


Figure 8. An(III)/Ln(III) selectivity for different N¹/N² polarizabilities according to ref 37.



$$\Delta E_g = \text{BE}([\text{CmL}_3]^{3+}) - \text{BE}([\text{GdL}_3]^{3+}) \quad (9)$$

$$\text{SF}_{\text{Cm/Gd}} = \exp\left(-\frac{\Delta E_g - 74.1 \frac{\text{kJ}}{\text{mol}}}{RT}\right) \quad (10)$$

First, BE of the 1:3 complexes for Gd(III) and Cm(III) were calculated (Table 5). Although the BE for PTD-OMe complexes are higher in general, smaller differences (ΔE_g , eq 9) in the BE of the 1:3 PTD-OMe complexes of Cm(III) and Gd(III) were found compared to the corresponding PTD complexes. Separation factors were calculated using equation 10 taking into account a difference of 74.1 kJ/mol between the corresponding aqua ions.³⁵

The smaller difference in binding energy (ΔE_g) of PTD-OMe leads to a decrease in selectivity by a factor of 5 from a calculated separation factor ($\text{SF}_{\text{Cm/Gd}}$) of 31 for PTD to 6 for PTD-OMe. This is in good agreement with experimental findings ($\text{SF}_{\text{Am/Eu}}(\text{PTD})_{\text{exp.}} = 100\text{--}36$; $\text{SF}_{\text{Am/Eu}}(\text{PTD-OMe})_{\text{exp.}} = 12\text{--}4$). Note that experimentally determined separation factors reflect the selectivity of both PTD and TODGA, with the Gd(III)/Cm(III) selectivity of TODGA being ~ 5 .⁶

CONCLUSION

Hoping to improve the extraction and complexation properties of PTD, a water-soluble complexing agent, PTD-OMe was synthesized. By placing a methoxy moiety at the 4-position of the central pyridine the aromatic core was activated. To study the impact of this activation the pK_a and the speciation in aqueous and acidic solutions were investigated.

The pK_a of PTD-OMe was determined using NMR. PTD-OMe ($\text{pK}_a = 2.54 \pm 0.08$) is more prone to protonation than PTD ($\text{pK}_a = 2.1$). Consequently, greater stability constants were expected^{41,42} for the metal ion complexes with PTD-OMe. TRLFS confirmed the conditional stability constant of the Cm(III) 1:3 complex to be almost 1 order of magnitude

Table 5. Calculated Atomic Charges (q), Dipole Polarizabilities (α), Binding Energies (BE), and Separation Factors (SF) of Cm(III) and Gd(III) 1:3 Complexes

	N ¹		N ²		BE(Cm) [kJ/mol]	BE(Gd) [kJ/mol]	$\text{SF}_{\text{Cm/Gd}}$
	q [e]	α [\AA^3]	q [e]	α [\AA^3]			
PTD	–0.13	0.32	–0.12	0.93	–3292.3	–3357.9	33
PTD-OMe	–0.14	0.42	–0.12	1.10	–3404.7	–3474.3	6

greater for PTD-OMe ($\log \beta_3(\text{PTD-OMe}) = 10.8 \pm 0.4$) than for PTD ($\log \beta_3(\text{PTD}) = 9.9 \pm 0.5$).

Unfortunately, in solvent extraction experiments (involving 0.44 mol/L nitric acid in the aqueous phase) PTD-OMe performed inferior to PTD. This was due to the smaller amount of free ligand present under the used solvent extraction conditions, although it was the stronger ligand. Therefore, protonation outcompeted complexation under solvent extraction conditions.

NMR experimental data and DFT calculations confirmed protonation of PTD-OMe to occur at the pyridine nitrogen atom. The lower selectivity of PTD-OMe compared to PTD was explained by an increased polarizability of the coordinating nitrogen atoms, actually leaving the small zone of optimum polarizability.

Clearly, the positive effect of methoxy substitution observed for some lipophilic N-heterocyclic extracting agents^{19–21} is overcompensated by increased susceptibility to protonation in the case of the water-soluble PTD complexing agents.

■ ASSOCIATED CONTENT

● Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: [10.1021/acs.inorgchem.9b02325](https://doi.org/10.1021/acs.inorgchem.9b02325).

Slope analyses, species distribution for the pK_a determination, and Cm(III) fluorescence lifetime measurements not shown in the manuscript (PDF)

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Notes

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