Exploiting Ladder Networks for Gene Expression Classification

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Abstract. The application of deep learning to biology is of increasing relevance, but it is difficult; one of the main difficulties is the lack of massive amounts of training data. However, some recent applications of deep learning to the classification of labeled cancer datasets have been successful. Along this direction, in this paper, we apply Ladder networks, a recent and interesting network model, to the binary cancer classification problem; our results improve over the state of the art in deep learning and over the conventional state of the art in machine learning; achieving such results required a careful adaptation of the available datasets and tuning of the network.

Keywords: Deep learning · Ladder network · Cancer detection
RNA-seq expression · Classification

1 Introduction

Gene expression measures the transcriptional activity of genes; the analysis of gene expression has a great potential to lead to biological discoveries; in particular, it can be used to explain the role of genes in causing tumors. Different forms of gene expression data (produced by micro-arrays or next generation sequencing through RNA-seq experiments) have been used for classification and clustering studies, using different approaches. In particular, Danaee et al. [1] applied deep learning for analyzing the binary classification problem for breast cancer using TCGA public dataset.

Deep learning is a branch of machine learning; it has achieved tremendous performance in several fields such as image classification, semantic segmentation and speech recognition [2–4]. Recently, deep learning methods have also achieved success in computational biology [5].

The problem considered in [1] consists of using classified gene expression vectors representing samples which are taken from normal and tumor cells (hence carrying a label) and then training a classifier to learn the label; this is an interesting preliminary problem for testing the usability of classifiers in medical studies. The problem is difficult in the context of deep learning, due to the high

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number of genes and the small number of samples ("small n large p" problem) [6]. In [1], the Stacked Denoising Autoencoder (SDAE) approach was compared to conventional machine learning methodologies. The comparison table of different feature selections and classifications is available in Table 3.

Deep learning can be performed in three ways: supervised, unsupervised and semi-supervised learning. Semi-supervised learning [7] uses supervised learning tasks and techniques to make use of unlabeled data for training. This method is recommended when the amount of labeled data is very small, while the unlabeled data is much larger. In this work, we use Ladder network [8] approach, which is a semi-supervised deep learning method, to classify tumorous or healthy samples of the gene expression data for breast cancer and we evaluated the Ladder network against the state-of-the-art machine learning and dimensionality reduction methods; therefore, our work directly compares to [1]. In comparison to the state-of-the-art, the Ladder structure yielded stronger results than both the machine learning algorithms and the SDAE approach of [1], thanks to its improved applicability to datasets with small sample sizes and high dimensions.

We considered the datasets extracted from the GMQL [9] project's public repository. They were originally published by TCGA [10] and enriched by TCGA2BED [11] project. Figure 1 illustrates the number of patients for each cancer type and also shows that there are fewer normal cells compared to the cancerous cells; Breast Invasive Carcinoma (BRCA) has the highest number of cases. We used TCGA RNA-seq V2 Rsem [12] gene normalized BRCA dataset with 1104 tumorous samples and 114 normal samples available.

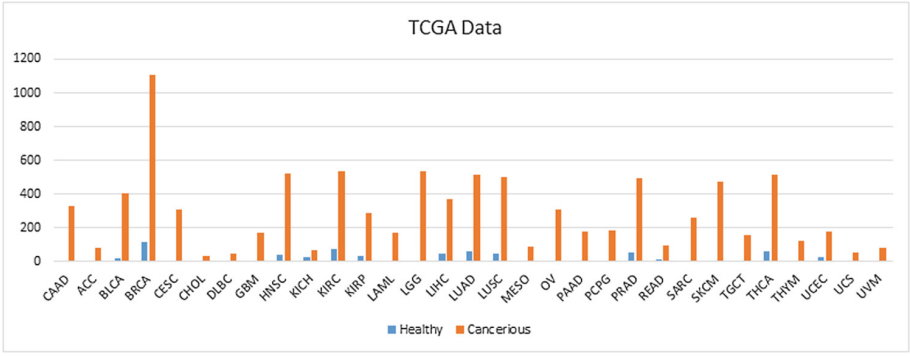

Fig. 1. The number of patients for each tumor type. Tumor type abbreviations are available at: http://gdc.cancer.gov/resources-tcga-users/tcga-code-tables/tcga-study-abbreviations

2 Dimensionality Reduction and Machine Learning Techniques

One of the main characteristics of the gene expression datasets is the high-dimensionality. Therefore, a feature selection or a feature extraction step is often required prior to the classification. Feature selection methods attempt to identify

the most informative subset of features. A common way of performing feature selection is to first compute the chi-squared statistic between each feature and the class labels, then to select the features having the highest chi-squared statistic scores [13]. Feature extraction methods, on the other hand, derive new features by combining the initial features of the dataset.

- Principal Component Analysis (PCA): is a well-established method for feature extraction that uses orthogonal transformations to derive uncorrelated features and increase the amount of variance explained [14].
- Kernel Principal Component Analysis (KPCA): is an extension of the PCA that uses kernel methods. With the help of the kernel methods, the principal components can be computed in the high-dimensional spaces [15].
- Non-negative matrix factorization (NMF): is a technique to reduce the dimensions of a non-negative matrix by finding two non-negative matrices, whose multiplication reconstructs an approximation of the initial matrix [16].

Support Vector Machines (SVM) is proposed by Vapnik and Cortes [17] and it has been extensively used on the classification of gene expression datasets [18–21]. Support vector machines can also be adopted to fit non-linear data by using kernel functions. Single layer and multi-layer perceptron architectures have also been widely used in predicting the gene expression profiles of the samples in various works [22–24].

3 Ladder Networks

Ladder networks are deep neural networks using both supervised and unsupervised learning; training of both supervised and unsupervised learning simultaneous, without using layer-wise pre-training (as in the Danaee et al. [1]).

We next provide a simplified description of implementation of the ladder network introduced in Rasmus et al. [8]:

1. A Ladder network has a feed-forward model that is used as a supervised learning encoder. The complete system has 2 encoder paths, one is *clean* the other is *corrupted*. The difference between them is the gaussian noises which are added to all layers of the corrupted one.
2. A decoder is utilized to acquire the inverse of the output at each layer. This decoder gets the benefit of using denoising function which reconstructs the activation of each layer in corrupted encoder to approximate the activation of the clean encoder. The term denoising cost is defined as the difference between reconstructed and the clean version of that layer.
3. Since it uses both supervised and unsupervised learning, it has corresponding costs for them. *Supervised cost* is the difference between the output of corrupted encoder and the desired output. *Unsupervised cost* is the sum of denoising cost of all layers scaled by the *significance parameter*. The entire cost of training the system is the summation of supervised and unsupervised cost.
4. Fully labeled and semi-supervised structures are trained to minimize the costs by using an optimization technique.

Figure 2 illustrates the structure of 2 layered ($l = 2$) ladder network example in Rasmus et al. [8]. The clean path at the right ($x \rightarrow z^{(1)} \rightarrow z^{(2)} \rightarrow y$) shares the mapping $f^{(l)}$ with the corrupted path on the left ($x \rightarrow \tilde{z}^{(1)} \rightarrow \tilde{z}^{(2)} \rightarrow y$). On each layer, the decoder in the middle ($\tilde{z}^{(l)} \rightarrow \hat{z}^{(l)} \rightarrow \hat{x}$) consists of denoising functions $g^{(l)}$ and cost functions $C_d^{(l)}$ try to minimize the difference between $\hat{z}^{(l)}$ and $z^{(l)}$.

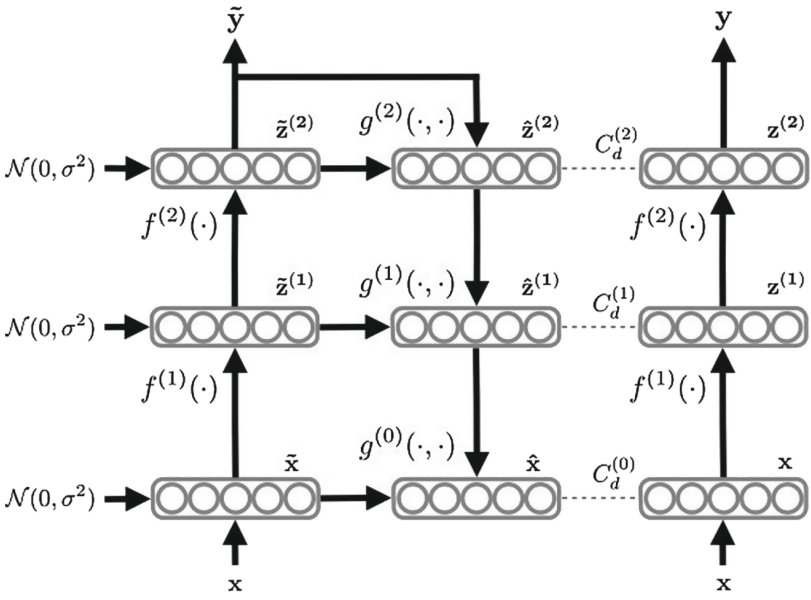

Fig. 2. Structure of 2 layered Ladder network. On the right there is clean path, which is work as supervised learning, in the middle and the left one is part of unsupervised learning with encoder (leftmost) and the decoder (middle).

The ability of ladder network reaching high accuracy with very small amount of labeled data on MNIST dataset [25] suggested us that it could be conveniently applied to our problem. To the best of our knowledge, this work is the first to apply the ladder network structure on the gene expression datasets.

Before analyzing the gene expression data, we applied preprocessing techniques to fill the missing data and also normalize all the expression data in order to get same expression level for each gene type. For this purpose, min-max normalization was applied on the data. In order to test properly, all samples are divided into three mutually disjoint subset: training, validation and test with 60%, 20% and 20%, respectively.

The configured Ladder Network is freely available as a python-based software implementation and source code online via an MIT License: http://github.com/acanakoglu/genomics-ladder.

4 Tuning of the Ladder Network

In order to optimize the network configuration, different hyper parameters of the network were analyzed. First of all, the number of layers and structure (number of nodes) of each layer were detected. Then, the batch size for a given network were analyzed, for the purpose of optimizing the execution time and the accuracy of the network.

Table 1. Ladder network performance with different number of levels

Layers	Accuracy	Sensitivity	Specificity	Precision	F_1 score
1 hidden layer[a]	55.33	57.23	39.13	90.36	0.700
2 hidden layers[b]	97.38	98.55	86.09	98.55	0.986
3 hidden layers[c]	96.64	97.28	90.43	98.99	0.981
5 hidden layers[d]	98.69	98.64	99.13	99.91	0.993
7 hidden layers[e]	97.30	99.17	81.54	97.83	0.985
10 hidden layers[f]	97.56	98.64	87.75	98.64	0.986

The number of the nodes:
[a] 1 layer → 2000
[b] 2 layers → 2000 - 200
[c] 3 layers → 2000 - 200 - 20
[d] 5 layers → 2000 - 1000 - 500 - 250 - 10
[e] 7 layers → 2048 - 1024 - 512 - 256 - 128 - 64 - 32
[f] 10 layers → 2048 - 1024 - 512 - 256 - 128 - 64 - 32 - 16 - 8 - 4

We tuned the network by using different parameters, the most relevant ones are the number of layers (single layer or 2, 3, 5, 7 and 10 hidden layers) as shown in Table 1 and the training feed size (10, 20, 30, 40, 60, 80 and 120 labeled data) as shown in Table 2. All of the evaluations were performed by using the 5-fold cross validation technique.

In the Table 1, we analyze the effect of the number of hidden ladders. As shown in the table, having 5 hidden layers produces the top performance. Having less than 5 hidden layers result in lower performance, yet, having more causes

Table 2. Ladder network performance with different batch sizes

Labeled data	Accuracy	Sensitivity	Specificity	Precision	F_1 score
10 label	85.08	85.06	85.22	98.22	0.912
20 label	89.76	98.80	50.22	89.66	0.940
30 label	95.82	98.43	74.24	96.92	0.977
40 label	97.64	98.64	85.87	98.53	0.987
60 label	98.69	98.64	99.13	99.91	0.993
80 label	97.62	98.46	89.09	98.91	0.987
120 label	98.36	98.64	95.65	99.54	0.991

overfitting of the data. The structure with 5 hidden layers has 2000, 1000, 500, 250 and 10 nodes for each layer and two output nodes, one for healthy, the other one for cancerous case. Significance number, which is mentioned in step 3 of the method, is selected as $[1000, 10, 0.1, 0.1, 0.1, 0.1, 0.1]$ respectively to indicate the importance of the layer. Figure 2 illustrates the model that is used for classification of TCGA BRCA data.

We also investigated the impact of using the supervised learning networks with different batch sizes; Table 2 shows that performance grows while increasing the batch sizes up to 40 samples and it is rather stable with more sample. Since the smaller batch sizes are computationally more efficient, we decided to use a batch size of 40. Terminating condition is satisfied either when the number of epochs reach 100 or when the training accuracy becomes more than 99%.

With this size, the ladder network converges in about 4 min of execution time over a dataset of about 1000 gene expression records, with about 20000 genes; execution took place on Nvidia GeForce GTX1060 GPU with 6 GB of RAM with the Tensorflow library [26]. It achieves accuracy of 98.69, sensitivity of 98.64, specificity of 99.13, precision of 99.91, F_1 score of 0.993.

5 Evaluation and Conclusions

In the evaluation we used the stratified k-fold cross validation [27] and it is applied on the data with k is equal to 5. In other words, the data were divided into 5 equal subsets such that the folds contains approximately equal proportion of cancerous and healthy samples. In each round, 4 subsets are used for training and validation and 1 subset is used for testing. The procedure is repeated 5 times, by excluding 1 part of the data for testing. This approach was also employed in [1] and for the evaluation of the conventional machine learning algorithms defined in the previous section.

The confusion matrix of each step was summed up and then we calculated the accuracy, sensitivity, specificity precision and F_1 score, as reported in the last section.

We evaluated our ladder network algorithm by comparing its performance metrics against the results from the Danaee et al.'s study [1]. A direct comparison shows that the SDAE network achieves its best result when coupled to SVM for feature selection and in such case, it achieves an accuracy of 98.04, which is slightly inferior to ours. The ladder network could be directly applied without the need for a preliminary feature reduction and it shows that the network learns the important features and it learns the classes.

As the performance of a learning algorithm does not only depend on the data, but also on the hyper-parameters. We performed hyper-parameter tuning on the support vector classifier along with three different dimensionality reduction algorithms, in order to observe an optimal performance from the support vector classifier. The *GridSearch* functionality of the scikit-learn [28] library was utilized for the hyper-parameter tuning. Subsequently, we compared the resulting performance of the support vector classifiers with the ladder network algorithm and reported on the Table 3.

Table 3. Algorithm comparison table

Features	Model	Accuracy	Sensitivity	Specificity	Precision	F_1 score
All	Ladder network	98.69	98.64	99.13	99.91	0.993
NMF†	SVM	98.60	99.45	90.35	99.01	0.992
PCA†		94.91	94.65	97.37	99.71	0.971
CHI2†		98.28	99.45	86.84	98.65	0.990
SDAE*	ANN	96.95	98.73	95.29	95.42	0.970
	SVM	98.04	97.21	99.11	99.17	0.981
	SVM-RBF	98.26	97.61	99.11	99.17	0.983
DIFFEXP500*	ANN	63.04	60.56	70.76	84.58	0.704
	SVM	57.83	64.06	46.43	70.42	0.618
	SVM-RBF	77.39	86.69	71.29	67.08	0.755
DIFFEXP0.05*	ANN	59.93	59.93	69.95	84.58	0.701
	SVM	68.70	82.73	57.50	65.04	0.637
	SVM-RBF	76.96	87.56	70.48	65.42	0.747
PCA*	ANN	96.52	98.38	95.10	95.00	0.965
	SVM	96.30	94.58	98.61	98.75	0.965
	SVM-RBF	89.13	83.31	99.47	99.58	0.906
KPCA*	ANN	97.39	96.02	99.10	99.17	0.975
	SVM	97.17	96.38	98.20	98.33	0.973
	SVM-RBF	97.32	89.92	99.52	99.58	0.943

†To further evaluate the performance of our ladder network, the hyperparameters of the support vector classifiers along with three different dimensionality reduction algorithms are tuned by an exhaustive search approach.

*The results are taken from Table 1 of Danaee et al. [1].

The table also shows that the ladder network algorithm improves over conventional machine learning algorithms, where the best method is KPCA. We also considered the same machine learning methods and actually found better results than [1], but inferior to the results obtained with the ladder network.

In conclusion, we have shown that a ladder network can be applied to binary classification of RNA-seq expression data, and compares well with state-of-the-art machine learning and with the previous attempt of solving this problem by using deep learning. Although improvements are small, they demonstrate that this deep learning method can be directly applied to datasets having less than a thousand samples. Our results indicate ladder networks are very promising candidates for solving classification problems over gene expression data.

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References

1. Danaee, P., Ghaeini, R., Hendrix, D.A.: A deep learning approach for cancer detection and relevant gene identification. In: Pacific Symposium on Biocomputing, pp. 219–229. World Scientific (2017)
2. Krizhevsky, A., Sutskever, I., Hinton, G.E.: ImageNet classification with deep convolutional neural networks. In: Advances in Neural Information Processing Systems, pp. 1097–1105 (2012)
3. Long, J., Shelhamer, E., Darrell, T.: Fully convolutional networks for semantic segmentation. In: Proceedings of the IEEE Conference on Computer Vision and Pattern Recognition, pp. 3431–3440 (2015)
4. Hinton, G., Deng, L., Yu, D., Dahl, G.E., Mohamed, A.R., Jaitly, N., Senior, A., Vanhoucke, V., Nguyen, P., Sainath, T.N., et al.: Deep neural networks for acoustic modeling in speech recognition: the shared views of four research groups. IEEE Signal Process. Mag. **29**(6), 82–97 (2012)
5. Singh, R., Lanchantin, J., Robins, G., Qi, Y.: DeepChrome: deep-learning for predicting gene expression from histone modifications. Bioinformatics **32**(17), i639–i648 (2016)
6. Chakraborty, S., Ghosh, M., Mallick, B.K.: Bayesian non-linear regression for large p small n problems. J. Am. Stat. Assoc. (2005)
7. Chapelle, O., Schlkopf, B., Zien, A.: Semi-Supervised Learning, 1st edn. The MIT Press, Cambridge (2010)
8. Rasmus, A., Berglund, M., Honkala, M., Valpola, H., Raiko, T.: Semi-supervised learning with ladder networks. In: Advances in Neural Information Processing Systems, pp. 3546–3554 (2015)
9. Masseroli, M., Pinoli, P., Venco, F., Kaitoua, A., Jalili, V., Palluzzi, F., Muller, H., Ceri, S.: GenoMetric Query Language: a novel approach to large-scale genomic data management. Bioinformatics **31**(12), 1881–1888 (2015)
10. Weinstein, J.N., Collisson, E.A., Mills, G.B., Shaw, K.R.M., Ozenberger, B.A., Ellrott, K., Shmulevich, I., Sander, C., Stuart, J.M., Cancer Genome Atlas Research Network, et al.: The cancer genome atlas pan-cancer analysis project. Nat. Genet. **45**(10), 1113–1120 (2013)
11. Cumbo, F., Fiscon, G., Ceri, S., Masseroli, M., Weitschek, E.: TCGA2BED: extracting, extending, integrating, and querying the cancer genome atlas. BMC Bioinform. **18**(1), 6 (2017)
12. Li, B., Dewey, C.N.: RSEM: accurate transcript quantification from RNA-seq data with or without a reference genome. BMC Bioinform. **12**(1), 323 (2011)
13. Forman, G.: An extensive empirical study of feature selection metrics for text classification. J. Mach. Learn. Res. **3**(Mar), 1289–1305 (2003)
14. Jolliffe, I.T.: Principal component analysis and factor analysis. In: Principal Component Analysis, pp. 115–128. Springer, New York (1986). https://doi.org/10.1007/978-1-4757-1904-8_7
15. Schölkopf, B., Smola, A., Müller, K.-R.: Kernel principal component analysis. In: Gerstner, W., Germond, A., Hasler, M., Nicoud, J.-D. (eds.) ICANN 1997. LNCS, vol. 1327, pp. 583–588. Springer, Heidelberg (1997). https://doi.org/10.1007/BFb0020217
16. Brunet, J.P., Tamayo, P., Golub, T.R., Mesirov, J.P.: Metagenes and molecular pattern discovery using matrix factorization. Proc. Nat. Acad. Sci. **101**(12), 4164–4169 (2004)

17. Vapnik, V., Cortes, C.: Support-vector networks. Mach. Learn. **20**(3), 273–297 (1995)
18. Furey, T.S., Cristianini, N., Duffy, N., Bednarski, D.W., Schummer, M., Haussler, D.: Support vector machine classification and validation of cancer tissue samples using microarray expression data. Bioinformatics **16**(10), 906–914 (2000)
19. Tuncel, M.A.: A statistical framework for the analysis of genomic data. Master's thesis, Politechnico di Milano (2017)
20. Vapnik, V.: The Nature of Statistical Learning Theory. Springer Science & Business Media, New York (2000). https://doi.org/10.1007/978-1-4757-3264-1
21. Guyon, I., Weston, J., Barnhill, S., Vapnik, V.: Gene selection for cancer classification using support vector machines. Mach. Learn. **46**(1), 389–422 (2002)
22. Wei, J.S., Greer, B.T., Westermann, F., Steinberg, S.M., Son, C.G., Chen, Q.R., Whiteford, C.C., Bilke, S., Krasnoselsky, A.L., Cenacchi, N., et al.: Prediction of clinical outcome using gene expression profiling and artificial neural networks for patients with neuroblastoma. Cancer Res. **64**(19), 6883–6891 (2004)
23. Khan, J., Wei, J.S., Ringner, M., Saal, L.H., Ladanyi, M., Westermann, F., Berthold, F., Schwab, M., Antonescu, C.R., Peterson, C., et al.: Classification and diagnostic prediction of cancers using gene expression profiling and artificial neural networks. Nat. Med. **7**(6), 673–679 (2001)
24. Vohradsky, J.: Neural network model of gene expression. FASEB J. **15**(3), 846–854 (2001)
25. Deng, L.: The mnist database of handwritten digit images for machine learning research [best of the web]. IEEE Signal Process. Mag. **29**(6), 141–142 (2012)
26. Abadi, M., Agarwal, A., Barham, P., Brevdo, E., Chen, Z., Citro, C., Corrado, G.S., Davis, A., Dean, J., Devin, M., et al.: TensorFlow: large-scale machine learning on heterogeneous systems (2015). tensorflow.org
27. Refaeilzadeh, P., Tang, L., Liu, H.: Cross-validation. In: Encyclopedia of Database Systems, pp. 532–538. Springer, Boston (2009). https://doi.org/10.1007/978-0-387-39940-9
28. Pedregosa, F., Varoquaux, G., Gramfort, A., Michel, V., Thirion, B., Grisel, O., Blondel, M., Prettenhofer, P., Weiss, R., Dubourg, V., et al.: Scikit-learn: machine learning in Python. J. Mach. Learn. Res. **12**(Oct), 2825–2830 (2011)